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UTILITY PATENT APPLICATION TRANSMITTAL

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ADDRESS TO:

Assistant Commissioner for Patents
Box Patent Application
Washington, D.C. 20231

Attorney Docket No. 94,784-L

First Named Inventor Stojiljkovic

Express Mail No. EL028732014US

Total Pages 143

APPLICATION ELEMENTS

1. ☒ Transmittal Form with Fee
2. ☒ Specification (including claims and abstract) [Total Pages 78]
3. ☒ Drawings [Total Sheets 47]
4. ☒ Oath or Declaration [Total Pages 4]
 - a. ☐ Newly executed
 - b. ☒ Copy from prior application
[Note Boxes 5 and 17 below]
 - i. ☐ Deletion of Inventor(s) Signed statement attached deleting inventor(s) named in the prior application
5. ☒ Incorporation by Reference: The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.
6. ☐ Microfiche Computer Program
7. ☒ Nucleotide and/or Amino Acid Sequence Submission
 - a. ☐ Computer Readable Copy
 - b. ☒ Paper Copy
 - c. ☐ Statement verifying above copies

ACCOMPANYING APPLICATION PARTS

8. ☒ Assignment Papers
9. ☐ Power of Attorney
10. ☐ English Translation Document (if applicable)
11. ☐ Information Disclosure Statement (IDS)
 - ☐ PTO-1449 Form
 - ☐ Copies of IDS Citations
12. ☒ Preliminary Amendment
13. ☒ Return Receipt Postcard (Should be specifically itemized)
14. ☒ Small Entity Statement(s)
 - ☐ Enclosed
 - ☒ Statement filed in prior application; status still proper and desired
15. ☐ Certified Copy of Priority Document(s)
16. ☐ Other:

17. ☒ This is a CONTINUING APPLICATION. Please note the following:

- a. ☒ This is a ☒ Continuation ☐ Divisional ☐ Continuation-in-part of prior application U.S. Serial No. 08/537,361, filed October 2, 1995, now U.S. Patent No. 6,121,037, issued September 19, 2000, which is a continuation-in-part of U.S. Serial No. 08/326,670, filed October 18, 1994, now U.S. Patent No. 5,698,438, issued December 16, 1997.
- b. ☐ Cancel in this application original claims ____ of the prior application before calculating the filing fee.
- c. ☐ Amend the specification by inserting before the first line the sentence:
This is a ☐ continuation ☐ divisional ☐ continuation-in-part of application Serial No.
- d. ☒ The prior application is assigned of record to Oregon Health Sciences University.

UTILITY PATENT APPLICATION TRANSMITTAL

Attorney Docket No. 94,784-L

APPLICATION FEES

BASIC FEE				\$ 690.00
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	
Total Claims	6 -20=		x \$18.00	\$ 0.00
Independent Claims	4 -3=		x \$78.00	\$ 78.00
<input type="checkbox"/> Multiple Dependent Claims(s) if applicable			+\$270.00	\$ 0.00
Total of above calculations =				\$ 768.00
Reduction by 50% for filing by small entity =				\$1/2 384.00
<input type="checkbox"/> Assignment fee if applicable			+\$40.00	\$ 0.00
TOTAL =				\$ 384.00

18. ☐ Please charge my Deposit Account No. 13-2490 in the amount of \$19. ☒ A check in the amount of \$ 384.00 is enclosed.

20. The Commissioner is hereby authorized to credit overpayments or charge any additional fees of the following types to Deposit Account No. 13-2490:

- a. ☒ Fees required under 37 CFR 1.16.
- b. ☒ Fees required under 37 CFR 1.17.
- c. ☐ Fees required under 37 CFR 1.18.

21. ☒ The Commissioner is hereby generally authorized under 37 CFR 1.136(a)(3) to treat any future reply in this or any related application filed pursuant to 37 CFR 1.53 requiring an extension of time as incorporating a request therefor, and the Commissioner is hereby specifically authorized to charge Deposit Account No. 13-2490 for any fee that may be due in connection with such a request for an extension of time.

22. CERTIFICATE OF MAILING

I hereby certify that I directed that the correspondence identified above be deposited with the United States Postal Service as "Express Mail Post Office to Addressee" under 37 CFR § 1.10 on the date indicated below and is addressed to the Asst. Commissioner for Patents, Box Patent Application, Washington, DC 20231.

23. USPTO CUSTOMER NUMBER

PATENT & TRADEMARK OFFICE



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24. CORRESPONDENCE ADDRESS

Name	McDonnell Boehnen Hulbert & Berghoff
Address	300 South Wacker Drive
City, State, Zip	Chicago, IL 60606

25. SIGNATURE OF APPLICANT, ATTORNEY, OR AGENT REQUIRED

Name Reg. No.	Kevin E. Noonan, Reg. No. 35,303
Signature	
Date	September 19, 2000

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE
(Case No. 94,784-L)

PATENT

In application of:

Stojiljkovic *et al.*

Serial No. To be assigned

Filed: September 19, 2000

For: Novel Bacterial Hemoglobin Receptor
Genes and Uses

Before the Examiner:

Group Art Unit: 1641

PRELIMINARY AMENDMENT

Assistant Commissioner of Patents
Washington, D.C. 20231

Sir:

Please enter the following amendments to the above-identified divisional application.

AMENDMENT

IN THE SPECIFICATION:

On page 1, line 1, please insert the following:

- This application is a divisional of U.S. Serial No. 08/537,361, filed October 2, 1995, now U.S. Patent No. 6,121,037, issued September 19, 2000, which is a continuation-in-part of U.S. Serial No. 08/326,670, filed October 18, 1994, now U.S. Patent 5,698,438, issued December 16, 1997. The disclosures of each of these prior applications are considered as being part of the disclosure of the application and are explicitly incorporated by reference herein.

On page 6, line 8, please replace "Figure 2" with --Figures 2A-2H--.

On page 6, line 11, please replace "Figure 2" with --Figures 2A-2H--.

On page 6, line 12, please replace "Figure 2" with --Figures 2A-2H--.

On page 6, line 27, please replace "Figure 7" with --Figures 7A-7I--.

On page 7, line 1, please replace "Figure 7" with --Figures 7A-7I--.

On page 7, line 2, please replace "Figure 7" with --Figures 7A-7I--.

On page 7, line 17, please replace "Figure 8" with --Figures 8A-8I--.

On page 7, line 20, please replace "Figure 8" with --Figures 8A-8I--.

On page 7, line 21, please replace "Figure 8" with --Figures 8A-8I--.

On page 8, line 8, please replace "Figure 9" with --Figures 9A-9I--.

On page 8, line 10, please replace "Figure 9" with --Figures 9A-9I--.

On page 8, line 12, please replace "Figure 9" with --Figures 9A-9I--.

On page 8, line 26, please replace "Figure 2" with --Figures 2A-2H--.

Please indent page 8, line 27.

On page 9, line 1, please replace "Figure 7" with --Figures 7A-7I--.

On page 9, line 5, please replace "Figure 8" with --Figures 8A-8I--.

On page 9, line 8, please replace "Figure 9" with --Figures 9A-9I--.

On page 9, line 24, please replace "Figure 2" with --Figures 2A-2H--.

On page 9, line 27, please replace "Figure 7" with --Figures 7A-7I--.

On page 10, line 1, please replace "Figure 8" with --Figures 8A-8I--.

On page 10, line 5, please replace "Figure 9" with --Figures 9A-9I--.

On page 11, line 9, please replace "at" with --that--.

On page 12, line 5, please replace "Figure 2" with --Figures 2A-2H--.

On page 12, line 8, please replace "Figure 7" with --Figures 7A-7I--.

On page 12, line 12, please replace "Figure 8" with --Figures 8A-8I--.

On page 12, line 16, please replace "Figure 9" with --Figures 9A-9I--.

On page 12, line 20, please delete "each".

On page 13, line 18, please replace "Figure 2" with --Figures 2A-2H--.

On page 13, line 26, please replace "Figure 7" with --Figures 7A-7I--.

On page 14, line 4, please replace "Figure 8" with --Figures 8A-8I--.

On page 14, line 12, please replace "Figure 9" with --Figures 9A-9I--.

On page 14, line 19, please delete the comma after "human".

On page 15, line 15, please replace "Figure 2" with --Figures 2A-2H--.

On page 15, line 23, please replace "Figure 7" with --Figures 7A-7I--.

On page 16, line 1, please replace "Figure 8" with --Figures 8A-8I--.

On page 16, line 11, please replace "Figure 9" with --Figures 9A-9I--.

On page 16, line 18, please insert a comma after "agonists".

On page 16, line 22, please replace "known or unknown" with --recognized or unrecognized--.

On page 17, line 17, please replace "Figure 2" with --Figures 2A-2H--.

On page 17, line 24, please replace "Figure 4" with --Figures 4A-4C--.

On page 18, line 9, please replace "Figure 7" with --Figures 7A-7I--.

On page 18, line 12, please replace "Figure 8" with --Figures 8A-8I--.

On page 18, line 15, please replace "Figure 9" with --Figures 9A-9I--.

On page 18, line 26, please replace "Figure 11" with --Figures 11A-11D--.

On page 19, line 6, please delete the comma after "by".

On page 19, line 7, please replace "Figure 2" with --Figures 2A-2H--.

On page 19, line 7, please replace "7" with --Figures 7A-7I--.

On page 19, line 7, please replace "8" with --Figures 8A-8I--.

On page 19, line 7, please replace "9" with --Figures 9A-9I--.

On page 19, line 10, please replace "Figure 2" with --Figures 2A-2H--.

On page 19, line 10, please replace "7" with --Figures 7A-7I--.

On page 19, line 10, please replace "8" with --Figures 8A-8I--.

On page 19, line 10, please replace "9" with --Figures 9A-9I--.

On page 19, line 19, please replace "Figure 2" with --Figures 2A-2H--.

On page 19, line 19, please replace "7" with --Figures 7A-7I--.

On page 19, line 19, please replace "8" with --Figures 8A-8I--.

On page 19, line 20, please replace "9" with --Figures 9A-9I--.

On page 20, line 15, please insert --can be prepared-- after "protein".

On page 23, line 12, please replace "manitol" with --mannitol--.

On page 23, line 17, please delete the comma after "salts".

On page 24, line 12, please replace "attenuated" with --attenuated--.

On page 27, line 16, please replace "The" with --Preferred--.

On page 28, line 6, please delete "also".

On page 28, line 26, please replace "a pathogenic" with --pathogenic--.

On page 31, line 2, please replace "hemoglobin" with --Hemoglobin--.

On page 31, line 5, please replace "haemin" with --hemin--.

On page 31, line 5, please replace "haemoglobin" with --hemoglobin--.

On page 31, line 19, please replace "onselective" with --on selective--.

On page 31, line 21, please replace "haemin" with --hemin--.

On page 33, line 2, please replace "Chemicals" with --Chemical--.

On page 33, line 13, please replace "ClaIfragment" with --ClaI fragment--.

On page 35, line 11, please replace "Figure 2" with --Figures 2A-2H--.

On page 35, line 19, please replace "Figure 2" with --Figures 2A-2H--.

On page 36, line 5, please replace "fur-type" with --Fur-type--.

On page 36, line 11, please replace "Figure 2" with --Figures 2A-2H--.

On page 36, line 18, please replace "Figure 2" with --Figures 2A-2H--.

On page 37, line 2, please replace "Figure 4" with --Figures 4A-4C--.

On page 37, line 15, please insert a comma after "Postle".

On page 38, line 12, please replace "Table 2" with --Table II--.

On page 38, line 13, please replace "Table 2" with --Table II--.

On page 39, line 16, please replace "our" with --the--.

On page 39, line 16, please replace "probe ," with --probe,--.

On page 42, line 11, please replace "Figures 7" with --Figures 7A-7I--.

On page 42, line 11, please replace "8" with --Figures 8A-8I--.

On page 42, line 11, please replace "9" with --Figures 9A-9I--.

On page 42, line 16, please replace "Figures 10" with --Figure 10--.

On page 42, line 16, please replace "11" with --Figures 11A-11D--.

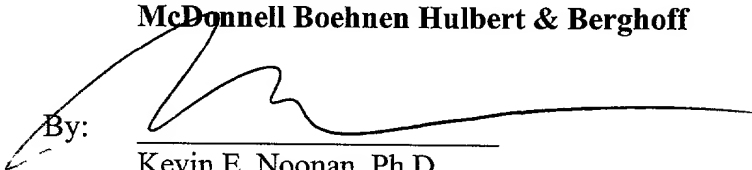
On page 42, line 24, please replace "Figure 11" with --Figures 11A-11D--.

If the Examiner in charge of this application believes it to be helpful, she is invited to contact the undersigned attorney by telephone at (312) 913-0001.

Respectfully submitted,
McDonnell Boehnen Hulbert & Berghoff

Date: September 16, 1999

By:


Kevin E. Noonan, Ph.D.
Reg. No. 35,303

NOVEL BACTERIAL HEMOGLOBIN RECEPTOR GENES AND USES

This application is a continuation-in-part of U.S. patent application Serial No. 08/326,670, filed October 18, 1994.

This invention was made with government support under National Institute of Health grants R01 AI32493 and R01 AI22933. The government has certain rights to this invention.

BACKGROUND OF THE INVENTION

1. Field of the Invention

This invention relates to hemoglobin receptor genes and the proteins encoded therefrom of certain bacterial species, particularly species of *Neisseria* bacteria. More particularly, this invention relates to hemoglobin receptor genes, polypeptides and peptides useful for preparing vaccines and antibodies against *Neisseria*, and methods and means for producing such peptides and polypeptides *in vitro*. Also provided are diagnostic and therapeutic methods and reagents useful in detecting and treating *Neisseria* infection and methods for developing novel and effective anti-*Neisseria* agents.

2. Background of the Invention

The *Neisseriae* comprise a genus of bacteria that includes two gram-negative species of pyogenic cocci pathogenic for humans: *Neisseria meningitidis* and *Neisseria gonorrhoeae*. *N. meningitidis* is a major cause of bacterial meningitis in humans, especially children. The disease characteristically proceeds from asymptomatic carriage of the bacterium in the nasopharynx to invasion of the bloodstream and cerebrospinal fluid in susceptible individuals.

Neisseria meningitidis is one of the leading causes of bacterial meningitis in children and healthy adults in the world. The severity of the disease is evidenced by the ability of meningococci to cause the death of previously healthy individuals in less than 24 hours. *N. meningitidis* has a polysaccharide capsule whose diversity of component antigenic polysaccharide molecules has resulted in the classification of ten different serogroups. Of these, group A strains are the classic epidemic strains; group B and C are generally endemic strains, but C occasionally causes an epidemic outbreak. All known group A strains have the same protein antigens on their

outer membranes, while group B strains have a dozen serotypes or groupings based on the presence of principal outer membrane protein antigens (as opposed to polysaccharides).

Survival of a pathogen such as *N. meningitidis* in a host depends on its ability to overcome a battery of host defense mechanisms. One nonspecific host defense mechanism against microbial intruders is to limit the availability of iron in tissues (Weinberg, 1984, *Physiological. Rev.* 64: 65-102), because iron is a necessary nutrient for most microbial pathogens. The vast majority of iron in the human adult is located intracellularly in the form of hemoglobin (76%) or ferritin (23%). The remainder can be found extracellularly bound to host iron-binding proteins such as transferrin and lactoferrin (Otto *et al.*, 1992, *Crit. Rev. Microbiol.* 18: 217-233).

Pathogenic bacteria have adapted to this iron-limiting environment by developing highly specific and effective iron assimilation systems. A large number of these bacteria secrete siderophores, small, non-protein iron chelators which, due to their extremely high affinity for iron (III), scavenge trace amounts of iron(III) from the environment and shuttle the iron back to the bacterial cell (Baggs and Neilands, 1987, *Microbiol. Rev.* 51: 509-518; Braun and Hantke, 1991, in Winkelmann (ed.), *Handbook of Microbial Iron Chelates*, CRC Press: Boca Raton, Fla., pp. 107-138.).

Alternatively, some bacterial pathogens, like *Neisseriae* species (Archibald and DeVoe, 1979, *FEMS Microbiol. Lett.* 6: 159-162; Mickelson *et al.*, 1982, *Infect. Immun.* 35: 915-920; Dyer *et al.*, 1987, *Infect. Immun.* 55: 2171-2175), *Haemophilus influenzae* (Coulton and Pang, 1983, *Curr. Microbiol.* 9: 93-98; Schryvers, 1988, *Mol. Microbiol.* 2: 467-472; Jarosik *et al.*, 1994, *Infect. Immun.* 62: 2470-2477), *Vibrio cholerae* (Stoebner and Payne, 1988, *Infect. Immun.* 56: 2891-2895; Henderson and Payne, 1994, *J. Bacteriol.* 176: 3269-3277), *Yersiniae* (Stojiljkovic and Hantke, 1992, *EMBO J.* 11: 4359-4367) and *Actinobacillus pleuropneumoniae* (Gerlach *et al.*, 1992, *Infect. Immun.* 60: 3253-3261) have evolved more sophisticated mechanisms to sequester iron from the host. These pathogens can directly bind host's iron-binding proteins such as lactoferrin, transferrin, and heme-containing compounds, and use them as sole sources of iron.

5 The importance of iron in the virulence of *N. meningitidis* was demonstrated by *in vivo* studies using mice as the animal model system (Calver *et al.*, 1976, *Can. J. Microbiol.* 22: 832-838; Holbien *et al.*, 1981, *Infect. Immun.* 34: 120-125). Specific iron-regulated outer membrane receptors have been shown to be involved in the binding and the utilization of lactoferrin- and transferrin-iron in *Neisseriae* (Schryvers and Morris, 1988, *Infect. Immun.* 56: 1144-1149 and *Mol. Microbiol.* 2: 281-288; Legrain *et al.*, 1993, *Gene* 130: 81-90; Pettersson *et al.*, 1993, *Infect. Immun.* 61: 4724-4733 and 1994, *J. Bacteriol.* 176: 1764-1766). These receptors share significant amino acid similarity and, most probably, also the mechanism of iron internalization, with receptors for siderophores and vitamin B12 of other Gram-negative bacteria (Cornelissen *et al.*, 1993, *J. Bacteriol.* 174: 5788-5797). In contrast, the mechanism by which *Neisseriae* utilize hemoglobin- and hemin-iron as well as the components involved have so far not been described.

15 Recently, several proteins with hemoglobin-binding and/or hemin-binding activities have been identified in total membranes of iron-limited *N. meningitidis* and *N. gonorrhoeae*.

Lee and Hill, 1992, *J. gen. Microbiol.* 138: 2647-2656 disclose the specific hemoglobin binding by isolated outer membranes of *N. meningitidis*.

Martek and Lee, 1994, *Infect. Immun.* 62: 700-703 disclosed that acquisition of heme iron by *N. meningitidis* does not involve meningococcal transferrin-binding proteins.

20 Lee, 1994, *Microbiol.* 140: 1473-1480 describes the biochemical isolation and characterization of hemin binding proteins from *N. meningitidis*.

The precise role of these proteins in hemin and/or hemoglobin utilization remains unclear at present, although these proteins are likely to be components of a hemin-utilization system in *N. meningitidis*.

25 The dependence on host iron stores for *Neisseria* growth is a potentially useful route towards the development of novel and effective therapeutic intervention strategies. Historically, infections of both *N. meningitidis* and *N. gonorrhoeae* were treated chemoprophylactically with sulfonamide drugs. However, with the development of sulfonamide-resistant strains came the necessity of using alternative modes of therapy such as antibiotic treatment. More recently, the drug treatment of choice includes the administration of high grade penicillin. However, the

success of antimicrobial treatment is decreased if therapy is not initiated early after infection.

Gonococcal infection has also been treated with penicillin, ampicillin, or amoxicillin, tetracycline hydrochloride, and spectinomycin. Unfortunately, because the incidence of infections due to penicillinase-producing bacteria has increased, several new, more expensive β -lactam antibiotics have been used in treatment. Despite the fact that existing antibiotics have decreased the serious consequences of gonorrhea, their use has not lowered the incidence of the infection in the general population.

Prevention of meningococcal disease has been attempted by chemoprophylaxis and immunoprophylaxis. At present, rifampin and minocycline are used, but only for humans in close contact with an infected person as this treatment has a number of disadvantages. The only commercially available vaccine against meningococcal meningitis has as its major component the bacterial polysaccharide capsule. In adults this vaccine protects against serogroups A, C, Y and W135. It is not effective against serogroup B, and is ineffective in children against serogroup C. Thus far, immunoprophylactic preventive treatment has not been available for *N. gonorrhoeae*.

Thus, what is needed are better preventative therapies for meningococcal meningitis and gonorrhea including more effective, longer lasting vaccines which protect across all of the serogroups of *N. meningitidis* and all the serotypes of *N. gonorrhoeae*. In addition, better methods are need to treat meningococcal and gonococcal infection.

SUMMARY OF THE INVENTION

The present invention relates to the cloning, expression and functional characterization of genes encoding bacterial hemoglobin receptor proteins. Specifically, the invention relates to genes encoding hemoglobin receptor proteins from *Neisseria* species, in particular *Neisseria meningitidis* and *N. gonorrhoeae*. The invention comprises species of nucleic acids having a nucleotide sequence encoding novel bacterial hemoglobin receptor proteins. Also provided by this invention is the deduced amino acid sequence of the cognate hemoglobin receptor proteins of these bacterial genes.

5 The invention provides nucleic acids, nucleic acid hybridization probes, recombinant expression constructs capable of expressing the hemoglobin receptor protein of the invention in cultures of transformed cells, preferably bacterial cells, and such cultures of transformed bacterial cells that express the hemoglobin receptor proteins of the invention. The invention also provides gene knockout vectors for inactivating the hemoglobin receptor protein gene in cells, particularly cells of *Neisseria* species, *via*, for example, homologous recombination and other mechanisms, and cultures of such hemoglobin receptor protein null mutant cells.

10 The invention also provides homogeneous preparations of the bacterial hemoglobin receptor proteins of the invention, as well as antibodies against and epitopes of the hemoglobin receptor protein. Methods for characterizing this receptor protein and methods for using the protein in the development of agents having pharmacological uses related to this receptor, particularly bactericidal and bacteriostatic uses, are also provided by the invention.

15 In other embodiments of this invention are provided diagnostic methods and reagents encompassing the use of the anti-*Neisseria* hemoglobin receptor protein antibodies of the invention. Still further embodiments provided herein include therapeutic methods and reagents encompassing the use of the anti-*Neisseria* hemoglobin receptor protein antibodies of the invention. Even more embodiments include diagnostic methods and reagents encompassing the use of the *Neisseria* hemoglobin receptor protein-encoding nucleic acids of the invention, as sensitive probes for the presence of *Neisseria* infection using nucleic acid hybridization techniques and/or *in vitro* amplification methodologies. Yet additional embodiments of the invention include therapeutic methods and reagents encompassing the use of the *Neisseria* hemoglobin receptor protein-encoding nucleic acids of the invention, comprising recombinant expression constructs engineered to produce antisense transcripts of the *Neisseria* hemoglobin receptor gene and fragments thereof, as well as recombinant knockout vectors of the invention. 20 The invention also provides the *Neisseria* hemoglobin receptor protein and epitopes thereof as components of vaccines for the development of non-disease associated immunity to pathological infection with bacteria of *Neisseria* species. 25

In a first aspect, the invention provides a nucleic acid having a nucleotide sequence encoding a bacterial hemoglobin receptor protein gene. In a preferred embodiment, the bacterial

hemoglobin receptor protein gene is isolated from bacteria of *Neisseria* species. In a particularly preferred embodiment, the hemoglobin receptor protein gene is isolated from *Neisseria meningitidis*, serotype C. In a particular example of this embodiment, the nucleic acid comprises a 3.3 kilobase (kb) *Bam*HI/*Hind*III fragment of *N. meningitidis* genomic DNA. In this embodiment, the nucleotide sequence comprises an open reading frame of 2376 nucleotides of *N. meningitidis* genomic DNA encoding 792 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the nucleotide sequence of the *N. meningitidis* hemoglobin receptor gene is the sequence depicted in Figure 2 (SEQ ID No:1). It will be understood that the *N. meningitidis* gene as disclosed herein is defined, insofar as is necessary, by the amino acid sequence of the protein encoded therein, said amino acid sequence being represented in Figure 2 (SEQ. ID No.:2). Thus, it will be understood that the particular nucleotide sequence depicted in Figure 2 (SEQ. ID. No.:1) is but one of a number of equivalent nucleotide sequences that encode the hemoglobin receptor protein, due to the degeneracy of the genetic code, and that all such alternative, equivalent nucleotide sequences are hereby explicitly encompassed within the disclosed nucleotide sequences of the invention. Also included herein are any mutant or allelic variations of this nucleotide sequence, either naturally occurring or the product of *in vitro* chemical or genetic modification. Each such variant will be understood to have essentially the same nucleotide sequence as the nucleotide sequence of the corresponding *N. meningitidis* hemoglobin receptor protein disclosed herein.

In another particularly preferred embodiment of this aspect of the invention, the hemoglobin receptor protein gene is isolated from *Neisseria meningitidis*, serotype A. In a particular example of this embodiment, the nucleic acid comprises a 2373 basepair (bp) polymerase chain reaction-amplified fragment of *N. meningitidis*, serotype A genomic DNA. In this embodiment, the nucleotide sequence comprises an open reading frame of 2373 nucleotides of *N. meningitidis* genomic DNA encoding 790 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the nucleotide sequence of the *N. meningitidis* hemoglobin receptor gene is the sequence depicted in Figure 7 (SEQ ID No:3). It will be understood that the *N. meningitidis* gene as disclosed herein is defined, insofar as is necessary, by the amino acid sequence of the protein encoded therein, said amino acid sequence

being represented in Figure 7 (SEQ. ID No.:4). Thus, it will be understood that the particular nucleotide sequence depicted in Figure 7 (SEQ. ID. No.:3) is but one of a number of equivalent nucleotide sequences that encode the hemoglobin receptor protein, due to the degeneracy of the genetic code, and that all such alternative, equivalent nucleotide sequences are hereby explicitly encompassed within the disclosed nucleotide sequences of the invention. Also included herein are any mutant or allelic variations of this nucleotide sequence, either naturally occurring or the product of *in vitro* chemical or genetic modification. Each such variant will be understood to have essentially the same nucleotide sequence as the nucleotide sequence of the corresponding *N. meningitidis* hemoglobin receptor protein disclosed herein.

In another particularly preferred embodiment of this aspect of the invention, the hemoglobin receptor protein gene is isolated from *Neisseria meningitidis*, serotype B. In a particular example of this embodiment, the nucleic acid comprises a 2376 basepair (bp) polymerase chain reaction-amplified fragment of *N. meningitidis*, serotype A genomic DNA. In this embodiment, the nucleotide sequence comprises an open reading frame of 2373 nucleotides of *N. meningitidis* genomic DNA encoding 791 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the nucleotide sequence of the *N. meningitidis* hemoglobin receptor gene is the sequence depicted in Figure 8 (SEQ ID No:5). It will be understood that the *N. meningitidis* gene as disclosed herein is defined, insofar as is necessary, by the amino acid sequence of the protein encoded therein, said amino acid sequence being represented in Figure 8 (SEQ. ID No.:6). Thus, it will be understood that the particular nucleotide sequence depicted in Figure 8 (SEQ. ID. No.:5) is but one of a number of equivalent nucleotide sequences that encode the hemoglobin receptor protein, due to the degeneracy of the genetic code, and that all such alternative, equivalent nucleotide sequences are hereby explicitly encompassed within the disclosed nucleotide sequences of the invention. Also included herein are any mutant or allelic variations of this nucleotide sequence, either naturally occurring or the product of *in vitro* chemical or genetic modification. Each such variant will be understood to have essentially the same nucleotide sequence as the nucleotide sequence of the corresponding *N. meningitidis* hemoglobin receptor protein disclosed herein.

5 In yet other preferred embodiments, the invention provides nucleic acid encoding a hemoglobin receptor protein gene isolated from *Neisseria gonorrhoeae*. In a particular example of this embodiment, the nucleic acid comprises a 2378 basepair (bp) polymerase chain reaction-amplified fragment of *N. gonorrhoeae* genomic DNA. In this embodiment, the nucleotide sequence comprises an open reading frame of 2373 nucleotides of *N. gonorrhoeae* genomic DNA encoding 791 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the nucleotide sequence of the *N. gonorrhoeae* hemoglobin receptor gene is the sequence depicted in Figure 9 (SEQ ID No:7). It will be understood that the *N. gonorrhoeae* gene as disclosed herein is defined, insofar as is necessary, by the amino acid sequence of the protein encoded therein, said amino acid sequence being represented in Figure 9 (SEQ. ID No.:8). Thus, it will be understood that the particular nucleotide sequence depicted in Figure 9 (SEQ. ID. No.:7) is but one of a number of equivalent nucleotide sequences that encode the hemoglobin receptor protein, due to the degeneracy of the genetic code, and that all such alternative, equivalent nucleotide sequences are hereby explicitly encompassed within the disclosed nucleotide sequences of the invention. Also included herein are any mutant or allelic variations of this nucleotide sequence, either naturally occurring or the product of *in vitro* chemical or genetic modification. Each such variant will be understood to have essentially the same nucleotide sequence as the nucleotide sequence of the corresponding *N. gonorrhoeae* hemoglobin receptor protein disclosed herein.

20 The invention also provides bacterial hemoglobin receptor proteins. In a preferred embodiment, the bacterial hemoglobin receptor protein is isolated from bacteria of *Neisseria* species. In a particularly preferred embodiment, the hemoglobin receptor protein is isolated from *Neisseria meningitidis*. In a particular example of this embodiment, the protein is derived from *N. meningitidis*, serotype C and comprises an amino acid sequence of 792 amino acids. In this embodiment of the invention, the amino acid sequence of the *N. meningitidis*, serotype C hemoglobin receptor protein is the sequence depicted in Figure 2 (SEQ ID No:2). In another example of this embodiment, the protein is derived from *N. meningitidis*, serotype A and comprises an amino acid sequence of 790 amino acids. In this embodiment of the invention, the amino acid sequence of the *N. meningitidis*, serotype A hemoglobin receptor

protein is the sequence depicted in Figure 7 (SEQ ID No:4). In yet another example of this embodiment, the protein is derived from *N. meningitidis*, serotype B and comprises an amino acid sequence of 791 amino acids. In this embodiment of the invention, the amino acid sequence of the *N. meningitidis*, serotype B hemoglobin receptor protein is the sequence depicted in Figure 8 (SEQ ID No:6). The invention also provides hemoglobin receptor protein derived from *N. gonorrhoeae*. In this embodiment of the invention, the protein comprises an amino acid sequence of 791 amino acids, and the amino acid sequence of the *N. gonorrhoeae* hemoglobin receptor protein is the sequence depicted in Figure 9 (SEQ ID No:8). Also explicitly encompassed within the scope of this invention are related bacterial hemoglobin receptor proteins, particularly such proteins isolated from *Neisseria* species, having essentially the same amino acid sequence and substantially the same biological properties as the hemoglobin receptor protein encoded by the *N. meningitidis* and *N. gonorrhoeae* nucleotide sequences described herein.

In another aspect, the invention provides a homogeneous preparation of an approximately 85.5 kiloDalton (kD) bacterial hemoglobin receptor protein or derivative thereof, said size being understood to be the size of the protein before any post-translational modifications thereof. Also provided is a 90kD embodiment of the receptor as determined by sodium dodecyl sulfate/polyacrylamide gel electrophoresis under reducing conditions. In a preferred embodiment, the bacterial hemoglobin receptor protein is isolated from bacteria of *Neisseria* species. In a particularly preferred embodiment, the hemoglobin receptor protein is isolated from *Neisseria meningitidis*. In one embodiment of this aspect of the invention, the protein is isolated from *N. meningitidis*, serotype C and the amino acid sequence of the bacterial hemoglobin receptor protein or derivative thereof preferably is the amino acid sequence of the hemoglobin receptor protein shown in Figure 2 (SEQ ID No:2). In a second embodiment of this aspect of the invention, the protein is isolated from *N. meningitidis*, serotype A and the amino acid sequence of the bacterial hemoglobin receptor protein or derivative thereof preferably is the amino acid sequence of the hemoglobin receptor protein shown in Figure 7 (SEQ ID No:4). In a third embodiment of this aspect of the invention, the protein is isolated from *N. meningitidis*, serotype B and the amino acid sequence of the bacterial hemoglobin receptor protein or derivative thereof

preferably is the amino acid sequence of the hemoglobin receptor protein shown in Figure 8 (SEQ ID No:6). The invention also provides a homogeneous preparation of a bacterial hemoglobin receptor protein isolated from *N. gonorrhoeae*. In a preferred embodiment, the amino acid sequence of the bacterial hemoglobin receptor protein or derivative thereof preferably is the amino acid sequence of the hemoglobin receptor protein shown in Figure 9 (SEQ ID No:8).

This invention provides nucleotide probes derived from the nucleotide sequences herein provided. The invention includes probes isolated from either complementary DNA (cDNA) copies of bacterial messenger RNA (mRNA) or bacterial genomic DNA (gDNA), as well as probes made synthetically or by *in vitro* amplification methods using the sequence information provided herein. The invention specifically includes but is not limited to oligonucleotide, nick-translated, random primed, or *in vitro* amplified probes made using cDNA or genomic clones embodying the invention, and oligonucleotide and other synthetic probes synthesized chemically using the nucleotide sequence information of cDNA or genomic clone embodiments of the invention.

It is a further object of this invention to provide such nucleic acid hybridization probes to detect the presence of bacteria of *Neisseria* species, particularly *N. meningitidis* and *N. gonorrhoeae*, in a biological sample in the diagnosis of a *Neisseria* infection in a human. Such a biological sample preferably includes blood, urine, semen, mucus, cerebrospinal fluid, peritoneal fluid and ascites fluids, as well as cell scrapings from the epithelium of the mouth, urethra, anus and rectum, and other organs.

The present invention also includes peptides encoded by the nucleotide sequences comprising the nucleic acid embodiments of the invention. The invention includes either naturally occurring or synthetic peptides which may be used as antigens for the production of hemoglobin receptor protein-specific antibodies. The invention also comprises such antibodies, preferably monoclonal antibodies, and cells and cultures of cells producing such antibodies.

Thus, the invention also provides antibodies against and epitopes of bacterial hemoglobin receptor proteins of the invention. It is an object of the present invention to provide antibodies that are immunologically reactive to the bacterial hemoglobin receptor proteins of the invention.

It is a particular object to provide monoclonal antibodies against these bacterial hemoglobin receptor proteins. In a preferred embodiment, antibodies provided are raised against bacterial hemoglobin receptor protein isolated from bacteria of *Neisseria* species. In a particularly preferred embodiment, such antibodies are specific for the hemoglobin receptor protein isolated from *Neisseria meningitidis* serotypes A, B or C. In additional particularly preferred embodiment, such antibodies are specific for the hemoglobin receptor protein isolated from *Neisseria gonorrhoeae*.

Hybridoma cell lines producing such antibodies are also objects of the invention. It is envisioned that such hybridoma cell lines may be produced as the result of fusion between a non-immunoglobulin producing mouse myeloma cell line and spleen cells derived from a mouse immunized with purified hemoglobin receptor protein or a cell expressing antigens or epitopes of bacterial hemoglobin receptor proteins of the invention. The present invention also provides hybridoma cell lines that produce such antibodies, and can be injected into a living mouse to provide an ascites fluid from the mouse that is comprised of such antibodies. In a preferred embodiment, antibodies provided are raised against bacterial hemoglobin receptor protein isolated from bacteria of *Neisseria* species. In a particularly preferred embodiment, such antibodies are specific for the hemoglobin receptor protein isolated from *Neisseria meningitidis*, serotypes A, B or C. In additional particularly preferred embodiment, such antibodies are specific for the hemoglobin receptor protein isolated from *Neisseria gonorrhoeae*.

It is a further object of the invention to provide immunologically-active epitopes of the bacterial hemoglobin receptor proteins of the invention. Chimeric antibodies immunologically reactive against the bacterial hemoglobin receptor proteins of the invention are also within the scope of this invention. In a preferred embodiment, antibodies and epitopes provided are raised against or derived from bacterial hemoglobin receptor protein isolated from bacteria of *Neisseria* species. In a particularly preferred embodiment, such antibodies and epitopes are specific for the hemoglobin receptor protein isolated from *Neisseria meningitidis*, serotypes A, B or C. In additional particularly preferred embodiment, such antibodies and epitopes are specific for the hemoglobin receptor protein isolated from *Neisseria gonorrhoeae*.

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The present invention provides recombinant expression constructs comprising a nucleic acid encoding a bacterial hemoglobin receptor protein wherein the construct is capable of expressing the encoded hemoglobin receptor protein in cultures of cells transformed with the construct. Preferred embodiments of such constructs comprise the *N. meningitidis*, serotype C hemoglobin receptor gene depicted in Figure 2 (SEQ ID No.:1), such constructs being capable of expressing the bacterial hemoglobin receptor protein encoded therein in cells transformed with the construct. Additional preferred embodiments of such constructs comprise the *N. meningitidis*, serotype A hemoglobin receptor gene depicted in Figure 7 (SEQ ID No.:3), such constructs being capable of expressing the bacterial hemoglobin receptor protein encoded therein in cells transformed with the construct. Further additional preferred embodiments of such constructs comprise the *N. meningitidis*, serotype B hemoglobin receptor gene depicted in Figure 8 (SEQ ID No.:5), such constructs being capable of expressing the bacterial hemoglobin receptor protein encoded therein in cells transformed with the construct. The invention also provides recombinant expression constructs encoding a hemoglobin receptor protein gene isolated from *ZN. gonorrhoeae*. In a particularly preferred embodiment, such constructs comprise the *N. gonorrhoeae* hemoglobin receptor gene depicted in Figure 9 (SEQ ID No.:7), the constructs being capable of expressing the bacterial hemoglobin receptor protein encoded therein in cells transformed with the construct.

The invention also provides cultures of cells, preferably bacterial cells, having been transformed with the recombinant expression constructs of the invention, each such cultures being capable of and in fact expressing the bacterial hemoglobin receptor protein encoded in the transforming construct.

The present invention also includes within its scope protein preparations of prokaryotic cell membranes containing the bacterial hemoglobin receptor protein of the invention, derived from cultures of prokaryotic cells transformed with the recombinant expression constructs of the invention.

The invention also provides diagnostic reagents and methods for using such reagents for detecting the existence of an infection in a human, with bacteria of a *Neisseria* species. In preferred embodiments, such diagnostic reagents comprise antibodies that are immunologically

reactive with a bacterial hemoglobin receptor protein. In a preferred embodiment, such antibodies are raised against a bacterial hemoglobin receptor protein isolated from bacteria of *Neisseria* species. In a particularly preferred embodiment, such antibodies are specific for the hemoglobin receptor protein isolated from *Neisseria meningitidis*, serotypes A, B or C. In
5 additional particularly preferred embodiments, such antibodies are specific for the hemoglobin receptor protein isolated from *Neisseria gonorrhoeae*.

In yet another embodiment of this aspect of the invention are provided diagnostic reagents and methods for using such reagents wherein said reagents are nucleic acid hybridization probes comprising a bacterial hemoglobin receptor gene. In a preferred embodiment, the bacterial
10 hemoglobin receptor protein gene is isolated from bacteria of *Neisseria* species. In a particularly preferred embodiment, the hemoglobin receptor protein gene is isolated from *Neisseria meningitidis*. In particular examples of this embodiment of the invention, the nucleic acid probes comprise a specifically-hybridizing fragment of a 3.3 kilobase (kb) *Bam*HI/*Hind*III fragment of *N. meningitidis*, serotype C genomic DNA. In this embodiment, the nucleotide sequence
15 comprises all or a specifically-hybridizing fragment of an open reading frame of 2376 nucleotides of *N. meningitidis*, serotype C genomic DNA encoding 792 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the nucleotide sequence of the *N. meningitidis*, serotype C hemoglobin receptor gene is the sequence depicted in Figure 2 (SEQ ID No:1). In another example of this embodiment of the invention, the nucleic acid
20 probes comprise a specifically-hybridizing fragment of a 2373bp, polymerase chain reaction-amplified fragment of *N. meningitidis*, serotype A genomic DNA. In this embodiment, the nucleotide sequence comprises all or a specifically-hybridizing fragment of an open reading frame of 2370 nucleotides of *N. meningitidis*, serotype A genomic DNA encoding 790 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the
25 nucleotide sequence of the *N. meningitidis*, serotype A hemoglobin receptor gene is the sequence depicted in Figure 7 (SEQ ID No:3). In yet another example of this embodiment of the invention, the nucleic acid probes comprise a specifically-hybridizing fragment of a 2376bp, polymerase chain reaction-amplified fragment of *N. meningitidis*, serotype B genomic DNA. In this embodiment, the nucleotide sequence comprises all or a specifically-hybridizing fragment

of an open reading frame of 2373 nucleotides of *N. meningitidis*, serotype B genomic DNA encoding 791 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the nucleotide sequence of the *N. meningitidis*, serotype B hemoglobin receptor gene is the sequence depicted in Figure 8 (SEQ ID No:5). The invention also provides nucleic acid hybridization probes comprising a bacterial hemoglobin receptor gene isolated from *N. gonorrhoeae*. In a preferred embodiment of this aspect of the invention, the nucleic acid probes comprise a specifically-hybridizing fragment of a 2378bp, polymerase chain reaction-amplified fragment of *N. gonorrhoeae* genomic DNA. In this embodiment, the nucleotide sequence comprises all or a specifically-hybridizing fragment of an open reading frame of 2373 nucleotides of *N. gonorrhoeae* genomic DNA encoding 791 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the nucleotide sequence of the *N. gonorrhoeae* hemoglobin receptor gene is the sequence depicted in Figure 9 (SEQ ID No:7). It will be understood that the term "specifically-hybridizing" when used to describe a fragment of a nucleic acid encoding a bacterial hemoglobin receptor gene is intended to mean that nucleic acid hybridization of such a fragment is stable under high stringency conditions of hybridization and washing as the term "high stringency" would be understood by those having skill in the molecular biological arts.

Also provided by the invention are therapeutic agents and methods for using such agents for treating the an infection in a human, with bacteria of a *Neisseria* species. In preferred embodiments, such agents comprise antibodies that are immunologically reactive with a bacterial hemoglobin receptor protein. In a preferred embodiment, such antibodies are raised against a bacterial hemoglobin receptor protein isolated from bacteria of *Neisseria* species. In a particularly preferred embodiment, such antibodies are specific for the hemoglobin receptor protein isolated from *Neisseria meningitidis*, serotypes A, B or C. In additional preferred embodiments, such antibodies are specific for the hemoglobin receptor protein isolated from *Neisseria gonorrhoeae*. Therapeutic agents provided in this aspect of the invention comprise such antibodies in a pharmaceutically-acceptable carrier, along with appropriate adjuvants and the like. In additional embodiments, such antibodies are covalently conjugated to a bactericidal

or bacteriostatic agent effective against bacteria of *Neisseria* species, preferably *N. meningitidis* and *N. gonorrhoeae*.

In yet another embodiment of this aspect of the invention are provided therapeutic reagents and methods for using such reagents wherein said reagents comprise recombinant expression constructs of the invention, or a homologue thereof that expresses the nucleic acid encoding a hemoglobin receptor in an antisense orientation. In a preferred embodiment, the bacterial hemoglobin receptor protein gene is isolated from bacteria of *Neisseria* species. In a particularly preferred embodiment, the hemoglobin receptor protein gene is isolated from *Neisseria meningitidis*. In particular examples of this embodiment of the invention, the nucleic acids comprise a specifically-hybridizing fragment of a 3.3 kilobase (kb) *Bam*HI/*Hind*III fragment of *N. meningitidis*, serotype C genomic DNA. In this embodiment, the nucleotide sequence comprises all or a specifically-hybridizing fragment of an open reading frame of 2376 nucleotides of *N. meningitidis*, serotype C genomic DNA encoding 792 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the nucleotide sequence of the *N. meningitidis*, serotype C hemoglobin receptor gene is the sequence depicted in Figure 2 (SEQ ID No:1). In another example of this embodiment of the invention, the nucleic acid probes comprise a specifically-hybridizing fragment of a 2373bp, polymerase chain reaction-amplified fragment of *N. meningitidis*, serotype A genomic DNA. In this embodiment, the nucleotide sequence comprises all or a specifically-hybridizing fragment of an open reading frame of 2370 nucleotides of *N. meningitidis*, serotype A genomic DNA encoding 790 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the nucleotide sequence of the *N. meningitidis*, serotype A hemoglobin receptor gene is the sequence depicted in Figure 7 (SEQ ID No:3). In yet another example of this embodiment of the invention, the nucleic acid probes comprise a specifically-hybridizing fragment of a 2376bp, polymerase chain reaction-amplified fragment of *N. meningitidis*, serotype B genomic DNA. In this embodiment, the nucleotide sequence comprises all or a specifically-hybridizing fragment of an open reading frame of 2373 nucleotides of *N. meningitidis*, serotype B genomic DNA encoding 791 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the nucleotide sequence of the *N. meningitidis*, serotype B hemoglobin receptor gene

is the sequence depicted in Figure 8 (SEQ ID No:5). The invention also provides recombinant expression constructs of the invention, or a homologue thereof that expresses the nucleic acid encoding a hemoglobin receptor in an antisense orientation, wherein the nucleic acid encodes a bacterial hemoglobin receptor gene isolated from *N. gonorrhoeae*. In a preferred embodiment of this aspect of the invention, the nucleic acid probes comprise a specifically-hybridizing fragment of a 2378bp, polymerase chain reaction-amplified fragment of *N. gonorrhoeae* genomic DNA. In this embodiment, the nucleotide sequence comprises all or a specifically-hybridizing fragment of an open reading frame of 2373 nucleotides of *N. gonorrhoeae* genomic DNA encoding 791 amino acids comprising the hemoglobin receptor gene. In this embodiment of the invention, the nucleotide sequence of the *N. gonorrhoeae* hemoglobin receptor gene is the sequence depicted in Figure 9 (SEQ ID No:7).

The invention also provides a method for screening compounds for their ability to inhibit, facilitate or modulate the biochemical activity of a bacterial hemoglobin receptor protein of the invention, for use in the *in vitro* screening of novel agonist and antagonist compounds and novel bactericidal and bacteriostatic agents specific for the hemoglobin receptor protein. In preferred embodiments, cells transformed with a recombinant expression construct of the invention are contacted with such a compound, and the binding capacity of the compounds, as well as the effect of the compound on binding of other, known hemoglobin receptor agonists such as hemoglobin and hemin, and antagonists, is assayed. Additional preferred embodiments comprise quantitative analyses of such effects.

The present invention is also useful for the detection of bactericidal and/or bacteriostatic analogues, agonists or antagonists, known or unknown, of a bacterial hemoglobin receptor protein, preferably derived from bacteria of *Neisseria* species, most preferably isolated from *N. meningitidis*, wherein such compounds are either naturally occurring or embodied as a drug.

The invention also provides vaccines for immunizing a human against infection with pathogenic bacteria of *Neisseria* species, the vaccines comprising the hemoglobin binding proteins of the invention or antigenic fragments thereof. In a preferred embodiment, the vaccines of the invention comprise cells expressing a hemoglobin receptor binding protein of the invention, or an antigenic fragment thereof, preferably wherein said cells are attenuated varieties

of cells adapted for growth in humans, *i.e.*, wherein such cells are non-pathogenic and do not cause bacteremia, endotoxemia or sepsis. Examples of such attenuated varieties of cells include attenuated strains of *Salmonella* species, for example *Salmonella typhi* and *Salmonella typhimurium*, as well as other attenuated bacterial species. Also provided by the invention are recombinant expression constructs as disclosed herein useful *per se* as vaccines, for introduction into an animal and production of an immunologic response to bacterial hemoglobin receptor protein antigens encoded therein.

Specific preferred embodiments of the present invention will become evident from the following more detailed description of certain preferred embodiments and the claims.

DESCRIPTION OF THE DRAWINGS

The foregoing and other objects of the present invention, the various features thereof, as well as the invention itself may be more fully understood from the following description, when read together with the accompanying drawings in which:

Figure 1 is a schematic drawing of the restriction enzyme digestion map of a *N. meningitidis* cosmid clone and subclones thereof derived as described in Example 2.

Figure 2 illustrates the nucleotide (SEQ ID No.:1) and deduced amino acid (SEQ ID No.:2) sequences of the *N. meningitidis* hemoglobin receptor protein encoded in a 3.3 kb *Bam*HI/*Hind*III DNA fragment.

Figure 3 presents a photograph of a stained SDS/ 10% PAGE electrophoresis gel showing the results of *in vitro* expression of the *N. meningitidis* hemoglobin receptor gene product as an approximately 90 kilodalton protein, and β -lactamase protein having a molecular weight of about 30.0 kilodaltons used as a molecular weight marker.

Figure 4 presents an amino acid sequence comparison between portions of the *N. meningitidis* transferrin receptor Tbp1 (SEQ ID No.:9), the *N. meningitidis* lactoferrin receptor LbpA (SEQ ID No.:10), and *N. meningitidis* hemoglobin receptor HmbR (SEQ ID No.:2).

Figure 5 illustrates Southern hybridization analysis of chromosomal DNA from *N. meningitidis* 8013 and the MC8013*hmbR* mutant using a *Bam*HI-*Sal*I fragment of the *hmb* gene as probe labeled using a DIG nonradioactive DNA labelling and detection kit (Boehringer

Mannheim Biochemicals, Indianapolis, IN). Lane 1 contains DNA from *N. meningitidis* strain MC8013, digested with *Cla*I; lane 2 is MC8031*hmbR* DNA digested with *Cla*I; lane 3, is MC8013 DNA digested with *Bam*HI and *Sa*II; and lane 4 is MC8013*hmbR* DNA digested with *Bam*HI and *Sa*II.

5 Figure 6 is a graph describing the course of infection using *N. meningitidis* wild type (MC8013) and *hmbR* mutant strains in an *in vivo* rat infant infection model. Each strain was injected intraperitoneally (2×10^6 CFU) into three infant inbred Lewis rats. The results represent the average of two similarly-performed experiments.

10 Figure 7 illustrates the nucleotide (SEQ ID No.:3) and deduced amino acid (SEQ ID No.:4) sequences of the *N. meningitidis*, serotype A hemoglobin receptor protein encoded on a 2373bp polymerase chain reaction-amplified DNA fragment.

Figure 8 illustrates the nucleotide (SEQ ID No.:5) and deduced amino acid (SEQ ID No.:6) sequences of the *N. meningitidis*, serotype B hemoglobin receptor protein encoded on a 2376bp polymerase chain reaction-amplified DNA fragment.

15 Figure 9 illustrates the nucleotide (SEQ ID No.:7) and deduced amino acid (SEQ ID No.:8) sequences of the *N. gonorrhoeae* hemoglobin receptor protein encoded on a 2376bp polymerase chain reaction-amplified DNA fragment.

20 Figure 10 represents a schematic of a nucleic acid sequence comparison between the hemoglobin receptor proteins derived from *N. meningitidis*, serotypes A (SEQ ID No.:3), B (SEQ ID No.:5) and C (SEQ ID No.:1) and from *N. gonorrhoeae* (SEQ ID No.:7), wherein the direction of trascription of the genes is in the direction of the arrow, and the following abbreviations refer to restriction endonuclease sites: H represents *Hind*III; N represents *Not*I; Bg represents *Bgl*I; Bs represents *Bss*HI; Nr represents *Nru*I; Cl represents *Cla*I; P represents *Pst*I; Sa represents *Sac*I; Av represents *Ava*I; B represents *Bam*HI; S represents *Sa*II; EV represents *Eco*RV; Sh represents *Sph*I; and Sy represents *Sty*I.

25 Figure 11 presents an amino acid sequence comparison between the hemoglobin receptor proteins derived from *N. meningitidis*, serotypes A (SEQ ID No.:4), B (SEQ ID No.:6) and C (SEQ ID No.:2) and from *N. gonorrhoeae* (SEQ ID No.:8).

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The term "bacterial hemoglobin receptor" as used herein refers to bacterial proteins comprising the outer membrane of Gram negative bacteria, which specifically mediate transit of hemoglobin-derived heme, as well as heme from other sources, through the outer membrane of such bacteria and into the periplasmic space. The bacterial hemoglobin receptor proteins of the invention are characterized by, first, an amino acid sequence that is essentially the sequence depicted in Figures 2 (SEQ ID No.:2), 7 (SEQ ID No.:4), 8 (SEQ ID No.:6) and 9 (SEQ ID No.:8). The bacterial hemoglobin receptor proteins of the invention are further characterized by having substantially the same biological activity as a protein having the amino acid sequence depicted in Figures 2 (SEQ ID No.:2), 7 (SEQ ID No.:4), 8 (SEQ ID No.:6) and 9 (SEQ ID No.:8). This definition is intended to encompass naturally-occurring variants and mutant proteins, as well as genetically engineered variants made by man.

Cloned, isolated and purified nucleic acid provided by the present invention may encode a bacterial hemoglobin receptor protein of any *Neisseria* species of origin, including, most preferably, *Neisseria meningitidis* species and serotypes thereof and *Neisseria gonorrhoeae* species.

The nucleic acid hybridization probes provided by the invention comprise DNA or RNA having all or a specifically-hybridizing fragment of the nucleotide sequence of the hemoglobin receptor protein as depicted in Figures 2 (SEQ ID No.:1), 7 (SEQ ID No.:3), 8 (SEQ ID No.:5) and 9 (SEQ ID No.:7), or any portion thereof effective in nucleic acid hybridization. Mixtures of such nucleic acid hybridization probes are also within the scope of this embodiment of the invention. Nucleic acid probes as provided herein are useful for detecting the presence of a bacteria, *inter alia*, in a human as the result of an infection, in contaminated biological samples and specimens, in foodstuffs and water supplies, or in any substance that may come in to contact with the human. Specific hybridization will be understood to mean that the nucleic acid probes of the invention are capable of forming stable, specific hybridization to bacterially-derived DNA or RNA under conditions of high stringency, as the term "high stringency" would be understood by those with skill in the art (*see, for example*, Sambrook *et al.*, 1989, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. and Hames and Higgins, eds., 1985, Nucleic Acid Hybridization, IRL Press, Oxford, U.K.).

Hybridization will be understood to be accomplished using well-established techniques, including but not limited to Southern blot hybridization, Northern blot hybridization, *in situ* hybridization and Southern hybridization to polymerase chain reaction product DNAs. The invention will thus be understood to provide oligonucleotides, specifically, pairs of oligonucleotides, for use as primers in support of *in vitro* amplification of bacterial hemoglobin receptor genes and mRNA transcripts.

The production of proteins such as bacterial hemoglobin receptor proteins from cloned genes by genetic engineering means is well known in this art. The discussion which follows is accordingly intended as an overview of this field, and is not intended to reflect the full state of the art. It will be understood from the following discussion that the hemoglobin receptor protein genes of this invention are particularly advantageous, since expression of such proteins by bacteria, including non-*Neisseria* species of bacteria, can complement certain auxotrophic mutants of said transformed bacteria otherwise unable to subsist absent supplementation of the growth media with iron (III).

DNA encoding a bacterial hemoglobin receptor protein, in view of the instant disclosure, by chemical synthesis, by screening reverse transcripts of mRNA from appropriate cells, by screening genomic libraries from appropriate cells, or by combinations of these procedures, as illustrated below. Screening of mRNA or genomic DNA may be carried out with oligonucleotide probes generated from the nucleic acid sequence information from the bacterial hemoglobin receptor protein disclosed herein. Probes may be labeled with a detectable group such as a fluorescent group, a radioactive atom or a chemiluminescent group in accordance with known procedures and used in conventional hybridization assays, as described in greater detail in the Examples below. In the alternative, bacterial hemoglobin receptor protein-encoding nucleic acids may be obtained by use of the polymerase chain reaction (PCR) procedure, using appropriate pairs of PCR oligonucleotide primers corresponding to nucleic acid sequence information derived from a bacterial hemoglobin receptor protein as provided herein. See U.S. Patent Nos. 4,683,195 to Mullis *et al.* and 4,683,202 to Mullis, as specifically disclosed herein in Example 9 below. In another alternative, such bacterial hemoglobin receptor protein-encoding

nucleic acids may be isolated from auxotrophic cells transformed with a bacterial hemoglobin receptor protein gene, thereby relieved of the nutritional requirement for uncomplexed iron (III).

Any bacterial hemoglobin receptor protein of the invention may be synthesized in host cells transformed with a recombinant expression construct comprising a nucleic acid encoding the bacterial hemoglobin receptor protein. Such recombinant expression constructs can also be comprised of a vector that is a replicable DNA construct. Vectors are used herein either to amplify DNA encoding a bacterial hemoglobin receptor protein and/or to express DNA encoding a bacterial hemoglobin receptor protein. For the purposes of this invention, a recombinant expression construct is a replicable DNA construct in which a nucleic acid encoding a bacterial hemoglobin receptor protein is operably linked to suitable control sequences capable of effecting the expression of the bacterial hemoglobin receptor protein in a suitable host cell.

The need for such control sequences will vary depending upon the host cell selected and the transformation method chosen. Generally, bacterial control sequences include a transcriptional promoter, an optional operator sequence to control transcription, a sequence encoding suitable mRNA ribosomal binding sites (the Shine-Delgarno sequence), and sequences which control the termination of transcription and translation. Amplification vectors do not require expression control domains. All that is needed is the ability to replicate in a host, usually conferred by an origin of replication, and a selection gene to facilitate recognition of transformants. *See, Sambrook et al., 1989, ibid.*

Vectors useful for practicing the present invention include plasmids and virus-derived constructs, including phage and particularly bacteriophage, and integratable DNA fragments (i.e., fragments integratable into the host genome by homologous recombination). The vector replicates and functions independently of the host genome, or may, in some instances, integrate into the genome itself. Suitable vectors will contain replicon and control sequences which are derived from species compatible with the intended expression host. A preferred vector is pLAFR2 (*see Riboli et al., 1991, Microb. Pathogen. 10: 393-403*).

Transformed host cells are cells which have been transformed or transfected with recombinant expression constructs made using recombinant DNA techniques and comprising nucleic acid encoding a bacterial hemoglobin receptor protein. Preferred host cells are cells of *Neisseria* species, particularly *N. meningitidis*, as well as *Salmonella typhi* and *Salmonella*

typhimurium species, and *Escherichia coli* auxotrophic mutant cells (*hemA aroB*). Transformed host cells may express the bacterial hemoglobin receptor protein, but host cells transformed for purposes of cloning or amplifying nucleic acid hybridization probe DNA need not express the receptor protein. When expressed, the bacterial hemoglobin receptor protein of the invention will typically be located in the host cell outer membrane. See, Sambrook *et al.*, *ibid*.

Cultures of bacterial cells, particularly cells of *Neisseria* species, and certain *E. coli* mutants, are a desirable host for recombinant bacterial hemoglobin receptor protein synthesis. In principal, any bacterial cell auxotrophic for uncomplexed iron (III) is useful for selectively growing bacterial hemoglobin receptor protein-transformed cells. However, for this purpose, well-characterized auxotrophs, such as *E. coli hemA aroB* mutants are preferred.

The invention provides homogeneous compositions of a bacterial hemoglobin receptor protein produced by transformed cells as provided herein. Each such homogeneous composition is intended to be comprised of a bacterial hemoglobin receptor protein that comprises at least 90% of the protein in such a homogenous composition. The invention also provides membrane preparations from cells expressing a bacterial hemoglobin receptor protein as the result of transformation with a recombinant expression construct of the invention, as described herein.

Bacterial hemoglobin receptor proteins, peptide fragments thereof and membranes derived from cells expressing such proteins in accordance with the present invention may be used for the production of vaccines effective against bacterial infections in a human, with pathogenic microorganisms expressing such bacterial hemoglobin receptor proteins. Such vaccines preferably would be effective in raising an immunological response against bacteria of *Neisseria* species, most preferably *N. meningitidis* and *N. gonorrhoeae*. Also encompassed within the vaccines provided by the invention are recombinant expression constructs as disclosed herein useful *per se* as vaccines, for introduction into an animal and production of an immunologic response to bacterial hemoglobin receptor protein antigens encoded therein.

Preparation of vaccines which contain polypeptide or polynucleotide sequences as active ingredients is well understood in the art. Typically, such vaccines are prepared as injectables, either as liquid solutions or suspensions. However, solid forms suitable for solution in, or suspension in, liquid prior to injection may also be prepared. The preparation may also be

emulsified. The active immunogenic ingredient is often mixed with excipients which are pharmaceutically acceptable and compatible with the active ingredient. Suitable excipients are, for example, water, saline, dextrose, glycerol, ethanol, or the like and combinations thereof. In addition, if desired, the vaccine may contain minor amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents, or adjuvants which enhance the effectiveness of the vaccine. The vaccines are conventionally administered parenterally, by injection, for example, either subcutaneously or intramuscularly. Additional formulations which are suitable for other modes of administration include suppositories and, in some cases, oral formulations. For suppositories, traditional binders and carriers may include, for example, polyalkylene glycols or triglycerides; such suppositories may be formed from mixtures containing the active ingredient in the range of 0.5% to 10%, preferably 1 to 2%. Oral formulations include such normally employed excipients as, for example, pharmaceutical grades of manitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate and the like. These compositions take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain 10% to 95% of active ingredient, preferably 25 to 70%.

The polypeptides of the invention may be formulated into the vaccine as neutral or salt forms. Pharmaceutically acceptable salts, include the acid addition salts (formed with the free amino groups of the peptide) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups may also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, 2-ethylamino ethanol, histidine, procaine, and the like.

In another embodiment, such vaccines are provided wherein the bacterial hemoglobin receptor proteins or peptide fragments thereof are present in the intact cell membranes of cells expressing such proteins in accordance with the present invention. In preferred embodiments, cells useful in these embodiments include attenuated varieties of cells adapted to growth in humans. Most preferably, said cells are attenuated varieties of cells adapted for growth in humans, *i.e.*, wherein such cells do not cause frank disease or other pathological conditions, such as bacteremia, endotoxemia or sepsis. For the purposes of this invention, "attenuated" cells

will be understood to encompass prokaryotic and eukaryotic cells that do not cause infection, disease, septicemia, endotoxic shock, pyrogenic shock, or other serious and adverse reactions to administration of vaccines to an animal, most preferably a human, when such cells are introduced into the animal, whether such cells are viable, living, heat-, chemically- or genetically attenuated or inactivated, or dead. It will be appreciated by those with skill in this art that certain minor side-effects of vaccination, such as short-term fever, muscle discomfort, general malaise, and other well-known reactions to vaccination using a variety of different types of vaccines, can be anticipated as accompanying vaccination of an animal, preferably a human, using the vaccines of the invention. Such acute, short-term and non-life-threatening side effects are encompassed in the instant definition of the vaccines of the invention, and vaccines causing such side-effects fall within the definition of "attenuated" presented herein. Preferred examples of such attenuated cells include attenuated varieties of *Salmonella* species, preferably *Salmonella typhi* and *Salmonella typhimurium*, as well as other attenuated bacterial species. It will be specifically understood that these embodiments of the vaccines of the invention encompass so-called "live" attenuated cell preparations as well as heat- or chemically-inactivated cell preparations.

In other embodiments of the invention are provided vaccines that are DNA vaccines, comprising the nucleic acids of the invention in recombinant expression constructs competent to direct expression of hemoglobin receptor proteins when introduced into an animal. In preferred embodiments, such DNA vaccines comprise recombinant expression constructs wherein the hemoglobin receptor-encoding nucleic acids of the invention are operably linked to promoter elements, most preferably the early gene promoter of cytomegalovirus or the early gene promoter of simian virus 40. DNA vaccines of the invention are preferably administered by intramuscular injection, but any appropriate route of administration, including oral, transdermal, rectal, nasal, aerosol administration into lung, or any other clinically-acceptable route of administration can be used by those with skill in the art.

In general, the vaccines of the invention are administered in a manner compatible with the dosage formulation, and in such amount as will be therapeutically effective and immunogenic. The quantity to be administered depends on the subject to be treated, capacity of the subject's immune system to synthesize antibodies, and the degree of protection desired. Precise amounts of active ingredient required to be administered depend on the judgment of the practitioner and are peculiar to each individual. However, suitable dosage ranges are of the

order of several hundred micrograms active ingredient per individual. Suitable regimes for initial administration and booster shots are also variable, but are typified by an initial administration followed in one or two week intervals by a subsequent injection or other administration.

5 The recombinant expression constructs of the present invention are also useful in molecular biology to transform bacterial cells which do not ordinarily express a hemoglobin receptor protein to thereafter express this receptor. Such cells are useful, *inter alia*, as intermediates for making cell membrane preparations useful for receptor binding activity assays, vaccine production, and the like, and in certain embodiments may themselves be used, *inter alia*,
10 as vaccines or components of vaccines, as described above. The recombinant expression constructs of the present invention thus provide a method for screening potentially useful bactericidal and bacteriostatic drugs at advantageously lower cost than conventional screening protocols. While not completely eliminating the need for ultimate *in vivo* activity and toxicology assays, the constructs and cultures of the invention provide an important first screening step for
15 the vast number of potentially useful bactericidal and bacteriostatic drugs synthesized, discovered or extracted from natural sources each year. In addition, such bactericidal or bacteriostatic drugs would be selected to utilize a nutritional pathway associated with infectious virulence in these types of bacteria, as disclosed in more detail below, thus selectively targeting bacteria associated with the development of serious infections *in vivo*.

20 Also, the invention provides both functional bacterial hemoglobin receptor proteins, membranes comprising such proteins, cells expressing such proteins, and the amino acid sequences of such proteins. This invention thereby provides sufficient structural and functional activity information to enable rational drug design of novel therapeutically-active antibacterial drugs using currently-available techniques (*see* Walters, "Computer-Assisted Modeling of
25 Drugs", in Klegerman & Groves, eds., 1993, Pharmaceutical Biotechnology, Interpharm Press: Buffalo Grove, IL, pp. 165-174).

30 Nucleic acids and oligonucleotides of the present invention are useful as diagnostic tools for detecting the existence of a bacterial infection in a human, caused by a hemoglobin receptor protein-expressing pathological organism of *Neisseria* species. Such diagnostic reagents comprise nucleic acid hybridization probes of the invention and encompass paired oligonucleotide PCR primers, as described above. Methods provided by the invention include blot hybridization, *in situ* hybridization and *in vitro* amplification techniques for detecting the

presence of pathogenic bacteria in a biological sample. Appropriate biological samples advantageously screened using the methods described herein include plasma, serum, lymph, cerebrospinal fluid, seminal fluid, mucosal tissue samples, biopsy samples, and other potential sites of bacterial infection. It is also envisioned that the methods of the invention may be used to screen water, foodstuffs, pharmaceuticals, and other potential sources of infection.

The invention also provides antibodies that are immunologically reactive to a bacterial hemoglobin receptor protein or epitopes thereof provided by the invention. The antibodies provided by the invention may be raised, using methods well known in the art, in animals by inoculation with cells that express a bacterial hemoglobin receptor protein or epitopes thereof, cell membranes from such cells, whether crude membrane preparations or membranes purified using methods well known in the art, or purified preparations of proteins, including fusion proteins, particularly fusion proteins comprising epitopes of a bacterial hemoglobin receptor protein of the invention fused to heterologous proteins and expressed using genetic engineering means in bacterial, yeast or eukaryotic cells, said proteins being isolated from such cells to varying degrees of homogeneity using conventional biochemical means. Synthetic peptides made using established synthetic means *in vitro* and optionally conjugated with heterologous sequences of amino acids, are also encompassed in these methods to produce the antibodies of the invention. Animals that are used for such inoculations include individuals from species comprising cows, sheep, pigs, mice, rats, rabbits, hamsters, goats and primates. Preferred animals for inoculation are rodents (including mice, rats, hamsters) and rabbits. The most preferred animal is the mouse.

Cells that can be used for such inoculations, or for any of the other means used in the invention, include any cell that naturally expresses a bacterial hemoglobin receptor protein as provided by the invention, or any cell or cell line that expresses a bacterial hemoglobin receptor protein of the invention, or any epitope thereof, as a result of molecular or genetic engineering, or that has been treated to increase the expression of an endogenous or heterologous bacterial hemoglobin receptor protein by physical, biochemical or genetic means. Preferred cells are *E. coli* auxotrophic mutant *hemA aroB* cells transformed with a recombinant expression construct of the invention and grown in media supplemented with hemin or hemoglobin as the sole iron (III) source, and cells of *Neisseria* species.

5 The present invention also provides monoclonal antibodies that are immunologically reactive with an epitope of a bacterial hemoglobin receptor protein of the invention, or fragment thereof, present on the surface of such cells, preferably *E. coli* cells. Such antibodies are made using methods and techniques well known to those of skill in the art. Monoclonal antibodies provided by the present invention are produced by hybridoma cell lines, that are also provided by the invention and that are made by methods well known in the art (*see* Harlow and Lane, 1988, Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.).

10 Hybridoma cell lines are made by fusing individual cells of a myeloma cell line with spleen cells derived from animals immunized with a homogeneous preparation of a bacterial hemoglobin receptor protein, membranes comprised thereof, cells expressing such protein, or epitopes of a bacterial hemoglobin receptor protein, used *per se* or comprising a heterologous or fusion protein construct, as described above. The myeloma cell lines used in the invention include lines derived from myelomas of mice, rats, hamsters, primates and humans. Preferred myeloma cell lines are from mouse, and the most preferred mouse myeloma cell line is P3X63-
15 Ag8.653. The animals from whom spleens are obtained after immunization are rats, mice and hamsters, preferably mice, most preferably Balb/c mice. Spleen cells and myeloma cells are fused using a number of methods well known in the art, including but not limited to incubation with inactivated Sendai virus and incubation in the presence of polyethylene glycol (PEG). The most preferred method for cell fusion is incubation in the presence of a solution of 45% (w/v) PEG-1450. Monoclonal antibodies produced by hybridoma cell lines can be harvested from cell culture supernatant fluids from *in vitro* cell growth; alternatively, hybridoma cells can be injected subcutaneously and/or into the peritoneal cavity of an animal, most preferably a mouse, and the monoclonal antibodies obtained from blood and/or ascites fluid.

25 Monoclonal antibodies provided by the present invention are also produced by recombinant genetic methods well known to those of skill in the art, and the present invention encompasses antibodies made by such methods that are immunologically reactive with an epitope of a bacterial hemoglobin receptor protein of the invention. The present invention also encompasses fragments, including but not limited to F(ab) and F(ab)₂ fragments, of such
30 antibody. Fragments are produced by any number of methods, including but not limited to

proteolytic cleavage, chemical synthesis or preparation of such fragments by means of genetic engineering technology. The present invention also encompasses single-chain antibodies that are immunologically reactive with an epitope of a bacterial hemoglobin receptor protein, made by methods known to those of skill in the art.

5 The antibodies and fragments used herein can be labeled preferably with radioactive labels, by a variety of techniques. For example, the biologically active molecules can also be labeled with a radionucleotide via conjugation with the cyclic anhydride of diethylenetriamine penta-acetic acid (DPTA) or bromoacetyl aminobenzyl ethylamine diamine tetra-acidic acid (BABE). See Hnatowich *et al.* (1983, *Science* 220: 613-615) and Meares *et al.* (1984, *Anal.*
10 *Biochem.* 142: 68-78, both references incorporated by reference) for further description of labeling techniques.

15 The present invention also encompasses an epitope of a bacterial hemoglobin receptor protein of the invention, comprised of sequences and/or a conformation of sequences present in the receptor molecule. This epitope may be naturally occurring, or may be the result of proteolytic cleavage of a receptor molecule and isolation of an epitope-containing peptide or may be obtained by synthesis of an epitope-containing peptide using methods well known to those skilled in the art. The present invention also encompasses epitope peptides produced as a result of genetic engineering technology and synthesized by genetically engineered prokaryotic or eukaryotic cells.

20 The invention also includes chimeric antibodies, comprised of light chain and heavy chain peptides immunologically reactive to a bacterial hemoglobin receptor protein-derived epitope. The chimeric antibodies embodied in the present invention include those that are derived from naturally occurring antibodies as well as chimeric antibodies made by means of genetic engineering technology well known to those of skill in the art.

25 Also provided by the present invention are diagnostic and therapeutic methods of detecting and treating an infection in a human, by a pathogenic organisms expressing a bacterial hemoglobin receptor protein. Diagnostic reagents for use in such methods include the antibodies, most preferably monoclonal antibodies, of the invention. Such antibodies are used in conventional immunological techniques, including but not limited to enzyme-linked
30 immunosorbent assay (ELISA), radioimmune assay (RIA), Western blot assay, immunological

titration assays, immunological diffusion assays (such as the Ouchterlony assay), and others known to those of skill in the art. Also provided are epitopes derived from a bacterial hemoglobin receptor protein of the invention and immunologically cross-reactive to said antibodies, for use in any of the immunological techniques described herein.

5 Additional diagnostic assays include nucleic acid hybridization assays, using the nucleic acids of the invention or specifically-hybridizing fragments thereof, for sensitive detection of bacterial genomic DNA and/or mRNA. Such assays include various blot assays, such as Southern blots, Northern blots, dot blots, slot blots and the like, as well as *in vitro* amplification assays, such as the polymerase chain reaction assay (PCR), reverse transcriptase-polymerase
10 chain reaction assay (RT-PCR), ligase chain reaction assay (LCR), and others known to those skilled in the art. Specific restriction endonuclease digestion of diagnostic fragments detected using any of the methods of the invention, analogous to restriction fragment linked polymorphism assays (RFLP) are also within the scope of this invention.

15 The invention also provides therapeutic methods and reagents for use in treating infections in a human, cause by a microorganism expressing a bacterial hemoglobin receptor protein of the invention, most preferably a bacteria of *Neisseria* species. Therapeutic reagents for use in such methods include the antibodies, most preferably monoclonal antibodies, of the invention, either *per se* or conjugated to bactericidal or bacteriostatic drugs or other antibiotic compounds effective against the infectious microorganism. In such embodiments, the antibodies
20 of the invention comprise pharmaceutical compositions, additionally comprising appropriate pharmaceutically-acceptable carriers and adjuvants or other ancillary components where necessary. Suitable carriers are, for example, water, saline, dextrose, glycerol, ethanol, or the like and combinations thereof. In addition, if desired, the pharmaceutical formulation may contain minor amounts of auxiliary substances such as wetting or emulsifying agents, pH
25 buffering agents, or other compounds which enhance the effectiveness of the antibody. In these embodiments, it will be understood that the therapeutic agents of the invention serve to target the infectious bacteria, either by immunologically "tagging" the bacteria with an antibody of the invention for recognition by cytotoxic cells of a human's immune system, or by specifically delivering an antimicrobial drug to the infectious microorganism *via* the bacterial hemoglobin
30 receptor protein.

Additional therapeutic reagents include the nucleic acids of the invention or fragments thereof, specifically antisense embodiments of such nucleic acids. Such antisense nucleic acids may be used themselves or embodied in a recombinant expression construct specific for antisense expression, wherein said construct is genetically engineered to co-opt a portion of the genome of a bacterial virus, preferably a bacteriophage, infectious for the bacterial pathogen responsible for the infection. In these embodiments, introduction of the antisense nucleic acids of the invention into the bacterial cell inhibits, attenuates or abolishes expression of the bacterial hemoglobin receptor, thereby reducing the virulence of the bacterial infection and enabling more effective antibacterial interventions. In additional embodiments, bacteriophage are provided bearing "knockout" copies of a bacterial hemoglobin receptor gene, whereby the phage achieves genetic mutation of the endogenous hemoglobin receptor gene in the infectious bacteria *via, for example*, homologous recombination of the exogenous knockout copy of the bacterial hemoglobin receptor gene with the endogenous hemoglobin receptor gene in the infectious microorganism.

The Examples which follow are illustrative of specific embodiments of the invention, and various uses thereof. They set forth for explanatory purposes only, and are not to be taken as limiting the invention.

EXAMPLE 1

Plasmids, bacteria, and media

Plasmids and bacteria used herein are listed on Table 1. *E. coli* strains were routinely grown in Luria-Bertani (LB) broth supplemented with 5-aminolevulinic acid and 50mg/L hemin chloride as necessary. *N. meningitidis* 8013 is a serogroup C clinical isolate (Nassif *et al.*, 1993, *Mol. Microbiol.* 8: 719-725). The meningococci were routinely grown on GCB agar (Difco) supplemented as described by Kellogg *et al.* (1963, *J. Bacteriol* 85: 1274-1279), and incubated at 37°C under a 5% CO₂ atmosphere. Transformation of meningococci was performed as described by Nassif *et al.* (1992, *Mol. Microbiol.* 6: 591-597). When necessary, the following antibiotics were used with *E. coli*: rifampicin, 100 mg/L; tetracycline, 15 mg/L; kanamycin, 30 mg/L; chloramphenicol, 20 mg/L; carbenicillin, 100 mg/L. For *Neisseriae*, kanamycin at 100 mg/L was used when needed.

EXAMPLE 2

Auxotroph Complementation Cloning of a hemoglobin Receptor Gene from *Neisseria meningitidis*

In order to identify *N. meningitidis* outer membrane receptor(s) involved in the uptake of haemin and/or haemoglobin iron, an auxotroph complementation cloning strategy was used, similar to the approach previously taken to identify the *Y. enterocolitica* and *V. cholerae* hemin receptors (see Stojiljkovic and Hantke, 1992, *EMBO J.* **11**: 4359-4367; Henderson and Payne, 1994, *J. Bacteriol.* **176**: 3269-3277). This strategy is based on the fact that the outer membrane of Gram-negative bacteria is impermeable to hemin (McConville and Charles, 1979, *J. Microbiol.* **113**: 165-168) and therefore *E. coli* porphyrin biosynthesis mutants cannot grow on exogenously supplied hemin. If provided with the *N. meningitidis* outer membrane hemin receptor gene, the *E. coli* porphyrin mutant would be able to use exogenously supplied hemin as its porphyrin source.

A cosmid bank of *N. meningitidis* 8013 clone 6 DNA was prepared using conventional cosmid cloning methodologies (Sambrook *et al.*, 1989, *ibid.*). *N. meningitidis* bacterial DNA was partially digested by *MboI*, size fractionated on sucrose gradients and cloned into the *Bam*HI site of the cosmid vector pLAFR2 (Riboli *et al.*, 1991, *Microb. Pathogen.* **10**: 393-403). This cosmid bank was mobilized into the *E. coli hema aroB Rif^r* recipient strain by triparental matings using a conjugal plasmid pRK2013::Tn9. The mating mixture was plated onselective plates containing hemin chloride (50mg/L), 0.1 mM 2,2'-dipyridil and rifampicin (100 mg/L). Several clones growing on exogenously supplied haemin were isolated after an overnight incubation.

The hemin utilization phenotype of these transformants was tested by re-introduction of the cosmids into naive *E. coli hema aroB* cells and by monitoring the growth on hemin-supplemented plates. The ability of *E. coli* strains to utilize heme or hemoglobin as the sole iron source was tested as previously described (Stojiljkovic and Hantke, 1992, *ibid.*). Cells were grown on LB agar supplemented with 50 μ M deferoxamine mesylate (an iron chelating agent, obtained from Sigma Chemical Co., St. Louis, MO). Filter discs (1/4 inches, Schleicher & Schuell, Inc., Keene, NH.) impregnated with the test compounds (20 μ L of 5 mg/ml stock solutions unless otherwise stated) were placed on these plates. After overnight growth at 37°C

TABLE I

<u>STRAIN</u>	<u>GENOTYPE</u>
<i>E. coli</i> K12	
EB53	<i>hemA, aroB, rpoB</i>
KP1041	MC4100 <i>tonB::Km^r</i>
H1388	<i>exbB::Tn10 Δlac pro</i>
TSM348	<i>endA, hsdR, pro, supF, pRK2013::Tn9</i>
IR754	EB53, <i>tonB::Km^r</i>
IR736	EB53, <i>exbB::Tn10</i>
DH5α	<i>recA, gyrB</i>
<i>N. meningitidis</i>	
ATCC 13077	Serotype A
--	Serotype B*
MC8013	clone 6, wild type
MChmbR	<i>hmbR::aphA-3</i>
<i>N. gonorrhoeae</i> MS11A	
<u>PLASMIDS</u>	
pSUSK	pA15 replicon, chloramphenicol ^r
pHEM22	pLAFR2, hemoglobin-utilizing cosmid
pHEM44	pLAFR2, hemin-utilizing cosmid
pIRS508	6kb <i>ClaI</i> , pSUSK
pIRS523	3kb <i>BamHI/SalI</i> , pUC19
pIRS525	1.2kb <i>aphA-3</i> , in <i>NotI</i> site of pIRS523
pIRS527	4kb <i>BamHI/ClaI</i> , pBluescript
pIRS528	0.7kb <i>NotI/BamHI</i> , pBluescript
pIRS692	3.3kb <i>BamHI/HindIII</i> , SU(SK)

* Laboratory collection

with 5% CO₂, zones of growth around the discs were monitored. The iron-bound proteins tested in this assay (all obtained from Sigma Chemicals Co.) were hemoglobin from human, baboon, bovine and mouse sources, bovine hemin, human lactoferrin (90% iron saturated), and human transferrin (90% iron saturated, obtained from Boehringer Mannheim Biochemicals, Indianapolis, IN). A total of six hemin utilization positive cosmids were obtained using this protocol. Results using such assays are shown in Table II.

EXAMPLE 3

Restriction Enzyme Digestion Mapping of Hemin Utilization Positive Cosmids

Cosmid DNA from six hemin-utilization positive cosmids obtained as described in Example 2 were digested with *Cla*I, and the resulting fragments were cloned into *Cla*I-digested pSU(SK) vector (obtained from Stratagene, LaJolla, CA). One subclone, containing a 6 kb *Cla*I fragment from cosmid cos22 (the resultant plasmid was designated pIRS508), was determined to allow utilization of hemin and hemoglobin by *E. coli hema aroB* assayed as described in Example 2. Another such clone, containing an 11 kb *Cla*I fragment from cos44 was also determined to allow hemin utilization in these auxotrophic mutant cells. Restriction analysis and Southern hybridization indicated that the DNA fragments originating from cos22 and cos44 are unrelated.

The deduced restriction enzyme digestion map of cosmid clone pIRS508 is shown in Figure 1. Plasmid pIRS508 enabled *E. coli hema aroB* to use both hemin and bovine hemoglobin as iron sources although growth on hemoglobin was somewhat weaker than on hemin (Table II). Further subcloning localized the hemin/hemoglobin utilization locus to the *Bam*HI/*Hind*III fragment of the insert. In addition to sequences encoding the hemoglobin receptor gene (designated *hmbR*), sequences for a *Neisseria* insertion element (IS1106) and a portion of a *Neisseria* small repetitive element (IR1) are also represented in the Figure.

Table II

STRAIN	ϕ -TYPE	HEMIN IRON	PORPHYRIN	Hb IRON
<i>N. meningitidis</i>				
MC8013	wild type	+++	N.T.	+++
MChmbR	Hb ^R mutant	+++	N.T.	-
<i>E. coli</i>				
EB53	iron utilization ⁻	-	-	-
EB53 (pIRS508)	<i>tonB</i> ⁺ , <i>exbB</i> ⁺ , <i>hmbR</i> ⁺	+++	+++	+
IR754(pIRS508)	<i>tonB</i> ⁺ , <i>exbB</i> ⁺ , <i>hmbR</i> ⁺	-	-	-
IR736(pIRS508)	<i>tonB</i> ⁺ , <i>exbB</i> ⁺ , <i>hmbR</i> ⁺	-	-	-

N.T.-not tested. Use of hemin/hemoglobin as a porphyrin source was tested by scoring for growth of strains around hemin (5mg/mL) or hemoglobin (for *E. coli*, 10 mg/mL; for *N. meningitidis*, 5 mg/mL) discs on LB plates. The use of the hemin/hemoglobin as an iron source was tested similarly except NBD plates supplemented with 50 μ L of 5 g/L delta-aminolevulinic acid were used (GCB plates supplemented with the 50 μ M Desferal in the case of *N. meningitidis*).

-: indicates no growth; +: less than 100 mm of growth zone around the disc; +++: ± 15 mm of growth zone around the disc.

EXAMPLE 4
Nucleotide Sequence Analysis of a Cosmid Clone Encoding
a *Neisseria* Hemoglobin Receptor Gene

The nucleotide sequence of the 3.3 kb *Bam*HI-*Hind*III DNA fragment carrying the *hmbR* gene and its promoter region was determined using the dideoxy chain termination method using a Sequenase 2.0 kit (obtained from U.S. Biochemicals, Cleveland, OH) and analyzed using a BioRad electrophoresis system, an AutoRead kit (obtained from Pharmacia, Uppsala, SE) and an ALF-370 automatic sequenator (Pharmacia, Uppsala, Sweden). Plasmid subclones for sequencing were produced by a nested deletion approach using Erase-a-Base kit (obtained from Promega Biotech, Madison, WI) using different restriction sites in the *hmbR* gene. The nucleotide and predicted amino acid sequences of the *hmbR* gene are shown in Figure 2

An open reading frame (ORF) encoding the *N. meningitidis*, serotype C hemoglobin receptor protein begins at position 470 of the sequence and encodes a protein having an amino acid sequence of 792 amino acids, with a calculated molecular weight of 85.5 kDa. A Shine-Delgarno sequence (SD) is found at position 460. The HmbR receptor protein contains a signal peptidase I recognition sequence at residues 22 to 24 of the protein (underlined), consistent with the fact that it is an outer membrane protein.

A typical Fur binding nucleotide sequence (designated "Fur box") was found in the promoter region of the *hmbR* gene (Figure 2). Like hemin utilization in *Yersinia* and *Vibrio*, hemin and hemoglobin utilization in *Neisseria* are known to be iron-inducible phenotypes (West and Sparling, 1985, *Infect. Immun.* 47: 388-394; Dyer *et al.*, 1987, *Infect. Immun.* 55: 2171-2175). In Gram-negative bacteria, conditional expression of many iron utilization genes is regulated by the Fur repressor, which recognizes a 19 bp imperfect dyad repeat (Fur-box) in the promoter regions of Fur-repressed genes. Recently, a genetic screen (FURTA) for the identification of Fur-regulated genes from different Gram-negative bacteria was described (Stojiljkovic *et al.*, 1994, *J. Mol. Biol.* 236: 531-545), and this assay was used to test whether *hmbR* expression was controlled in this way. Briefly, a plasmid carrying a Fur-box sequence is transformed into an *E. coli* strain (H1717) which possesses a Fur-regulated *lac* fusion in the chromosome. Expression of this Fur-regulated *lac* fusion is normally repressed. Introduction of a multicopy Fur-box sequence on the plasmid titrates the available Fur repressor thus allowing

expression of the Fur-regulated *lac* fusion (this phenotype is termed FURTA positive). Using this screen, the smallest insert fragment from cosmid pIRS508 that produced a FURTA positive result was a 0.7 kb *Bam*HI-*Not*I DNA fragment carried on plasmid pIRS528 (see Figure 1). This result indicated that the 0.7 kb *Bam*HI-*Not*I fragment carries a Fur-box and that gene expression from the *hmbR* promoter is controlled by a fur-type operon.

N. meningitidis, serotype C hemoglobin receptor protein was expressed *in vitro* using an *E. coli* S30 extract system from Promega Biotech (Madison, WI). The 3.3 kb *Bam*HI-*Hind*III fragment, expressed *in vitro*, encoded a 90kDa protein which corresponds in size to the predicted molecular weight of the unprocessed HmbR receptor. SDS/ 10% PAGE analysis showing the observed M_r of 90K is shown in Figure 3.

Immediately downstream of the *hmbR* gene (at positions 2955 to 3000 bp in Figure 2) was found a short nucleotide sequence that is 99% identical to the flanking sequence of the PIII gene of *N. gonorrhoeae* (Gotschlich *et al.*, 1987, *J. Exp. Med.* 165: 471-482). The first 26 bp of this sequence represents one half of the inverted repeat (IR1) of the *N. gonorrhoeae* small repetitive element. This element is found in approximately 20 copies in both *N. gonorrhoeae* and *N. meningitidis* (Correia *et al.*, 1988, *J. Biol. Chem.* 263: 12194-12198). The analysis of the nucleotide sequence from position 3027 to the *Cla*I (3984) restriction site (only the nucleotide sequence from *Bam*HI (1) to *Hind*III (3370) is shown in Figure 2) indicated the presence of an IS1106 element (Knight *et al.*, 1992, *Mol. Microbiol.* 6: 1565-1573). Interestingly, no nucleotide sequence similar to the IS1106 inverted repeat was found between the IR1 element and the beginning of the homology to IS1106.

These results were consistent with the cloning and identification of a novel hemoglobin receptor protein gene from *N. meningitidis*, embodied in a 3.3kb *Bam*HI/*Hind*III fragment of *N. meningitidis* genomic DNA.

EXAMPLE 5

Amino Acid Sequence Comparison of the *N. meningitidis* Hemoglobin Receptor Protein and *Neisseria* Lactoferrin and Transferrin Receptor Proteins

A comparison of the transferrin (Tbp1; Legrain *et al.*, 1993, *Gene* 130: 81-90), lactoferrin (LbpA; Pettersson *et al.*, 1993, *Infect. Immun.* 61: 4724-4733, and 1994, *J.*

Bacteriol. 176: 1764-1766) and hemoglobin receptors (HmbR) from *N. meningitidis* is shown in Figure 4. The comparison was done with the CLASTAL program from the PC/GENE program package (Intelligenetics, Palo Alto, CA). Only the amino-terminal and carboxyl terminal segments of the proteins are shown. An asterisk indicates identity and a point indicates similarity at the amino acid level. Lactoferrin and transferrin receptors were found to share 44.4% identity in amino acid sequence. In contrast, homology between these proteins and the hemoglobin receptor disclosed herein was found to be significantly weaker (22% amino acid sequence identity with lactoferrin and 21% with transferrin receptor).

EXAMPLE 6

TonB/ExbBD-Dependence of Hemin Transport by the *N. meningitidis* Hemoglobin Receptor

It was known that the transport of iron-containing siderophores, some colicins and vitamin B12 across the outer membrane of *E. coli* depends on three cytoplasmic membrane proteins: TonB, ExbB and ExbD (Postle 1990, *Mol. Microbiol.* 133: 891-898; Braun and Hantke, 1991, in Winkelmann, (ed.), Handbook of Microbial Iron Chelates, CRC Press, Boca Raton, Fla., pp. 107-138). In *Yersinia* and *Hemophilus*, hemin uptake was shown to be a TonB-dependent process (Stojiljkovic and Hantke, 1992, *ibid.*; Jarosik *et al.*, 1994, *Infect. Immun.* 62: 2470-2477). Through direct interaction between the outer membrane receptors and the TonB cytoplasmic machinery, the substrate bound to the receptor is internalized into the periplasm (Heller *et al.*, 1988, *Gene* 64: 147-153; Schoffler and Braun, 1989, *Molec. Gen. Genet.* 217: 378-383). This direct interaction has been associated with a particular amino acid sequence in membrane proteins associated with the TonB machinery.

All TonB-dependent receptors in Gram-negative bacteria contain several regions of high homology in their primary structures (Lundrigan and Kadner, 1986, *J. Biol. Chem.* 261: 10797-10801). In the amino acid sequence comparison described in Example 5, putative TonB-boxes of all three proteins are underlined. The carboxyl terminal end of the HmbR receptor contains the highly conserved terminal phenylalanine and position 782 arginine residues thought to be part of an outer membrane localization signal (Struyve *et al.*, 1991, *J. Mol. Biol.* 218: 141-148; Koebnik, 1993, *Trends Microbiol.* 1: 201). At residue 6 of the mature HmbR protein, an amino

acid sequence - ETTPVKA - is similar in sequence to the so called TonB-boxes of several Gram-negative receptors (Heller *et al.*, 1988, *ibid.*). Interestingly, the putative TonB-box of HmbR has more homology to the TonB-box of the *N. gonorrhoeae* transferrin receptor (Cornelissen *et al.*, 1992, *J. Bacteriol.* 174: 5788-5797) than to the TonB-boxes of *E. coli* siderophore receptors. When the sequence of the HmbR receptor was compared with other TonB-dependent receptors, the highest similarity was found with *Y. enterocolitica* HemR receptor although the similarity was not as high as to the *Neisseria* receptors.

In order to prove the TonB-dependent nature of the *N. meningitidis*, serotype C hemoglobin receptor, *hmbR* was introduced into *exbB* and *tonB* mutants of *E. coli* EB53, and the ability of the strains to utilize hemin and hemoglobin as porphyrin and iron sources was assessed. In these assays, both mutants of *E. coli* EB53 were unable to use hemin either as a porphyrin source or as an iron source in the presence of a functional *hmbR* (Table 2). The usage of hemoglobin as an iron source was also affected (Table 2). These results are consistent with the notion that the *hmbR* gene product, the *N. meningitidis* hemoglobin receptor protein of the invention, is TonB-dependent, since expression of this gene in TonB wild type *E. coli* supported the use of hemin and hemoglobin as sole iron source in the experiments disclosed in Example 2.

EXAMPLE 7

Functional Demonstration that the *hmbR* Gene Product is the Hemoglobin Receptor Protein in *N. meningitidis*

As shown in the data presented in Table II, *hmbR* mediated both hemin and hemoglobin utilization when expressed in *E. coli*, but hemoglobin utilization was less vigorous than hemin utilization. To determine if the HmbR receptor has the same specificity in *N. meningitidis*, *hmbR* was inactivated with a 1.2kb kanamycin cassette (*aphA-3*; Nassif *et al.*, 1991, *ibid.*) and transformed into wild-type *N. meningitidis* 8013 clone 6 (serotype C) cells. The inactivation of the chromosomal *hmbR* copy of the Km-resistant transformants was confirmed by Southern hybridization, as shown in Figure 5. As can be seen from Figure 5, wild-type *N. meningitidis* genomic DNA contains only one copy of the *hmbR* gene (lanes 1 and 3). In the Km^r transformants, the size of the DNA fragments containing the wild-type gene has increased by

1.2 kb, which is the size of the Kan cassette (Figure 5, lanes 2 and 4). When tested for its ability to utilize different iron-containing compounds, these mutant cells were found to be unable to use hemoglobin-bound iron, regardless of the source (human, bovine, baboon, mouse). The ability of the mutant to utilize hemoglobin-haptoglobin was not tested because the wild-type *N. meningitidis* strain is unable to use haptoglobin-haemoglobin complex as an iron source. However, the mutant was still able to use hemin iron, lactoferrin- and transferrin-bound iron as well as citrate-iron (Table II). As the iron-containing component of hemoglobin is hemin, a hemoglobin receptor would be expected to be capable of transporting hemin into the periplasm. Indeed, the cloning strategy disclosed herein depended on the ability of the cloned meningococcal receptor to transport hemin into the periplasm of *E. coli*. These results strongly suggest that *N. meningitidis* has at least two functional receptors that are involved in the internalization of hemin-containing compounds. One is the hemoglobin receptor described herein, which allows the utilization of both hemin and hemoglobin as iron sources. The other putative receptor in *N. meningitidis* is a hemin receptor which allows utilization of only hemin. This schema is also consistent with the isolation of several cosmid clones that allow *E. coli* EB53 to utilize hemin. DNAs from these cosmids do not hybridize with our *hmbR* probe, indicating that these clones encode a structurally-distinct receptor protein capable of transporting hemin into the periplasm of *N. meningitidis* cells.

EXAMPLE 8

Attenuation of Virulence in *hmbR* Mutant *N. meningitidis* Cells *In Vivo*

In order to test the importance of hemoglobin and hemin scavenging systems of *N. meningitidis* *in vivo*, the *hmbR* -mutant and the wild type strain of *N. meningitidis*, serotype C were inoculated into 5 day old infant rats and the numbers of bacteria recovered from blood and cerebrospinal fluid were followed. In these experiments, the method for the assessing *N. meningitidis*, serotype C virulence potential was essentially the same as described by Nassif *et al.* (1992, *ibid.*) using infant inbred Lewis rats (Charles River, Saint Aubin les Elbeufs, France). Inbred rats were used to minimize individual variations. Briefly, the 8013 strain was reactivated by 3 animal passages. After the third passage, bacteria were kept frozen in aliquots at -80° C.

To avoid the possibility that modifications in the course of infection could result from selection of one spontaneous avirulent variant, one aliquot from the animal-passed frozen stock of 8013 was transformed with chromosomal DNA from the *hmbR* mutant, the resultant Kan^r transformants were pooled without further purification and kept frozen at -80°C. For each experiment, all infant rats were from the same litter. *N. meningitidis* 8013 was grown overnight and 2 X 10⁶ bacteria injected intraperitoneally into the infant rat. Three rats were used for each meningococcal strain. The course of infection was followed over a 24 hours time period with blood collected at the indicated times. At the 24 h time period, the rats were sacrificed, the cerebrospinal fluid (CSF) collected and the number of colony-forming units (CFU) determined. Each experiment was performed in replicate; similar results were obtained both times.

The results of these experiments are shown in Figure 6. The *hmbR* strain, which is unable to use hemoglobin as an iron source, was recovered from the blood of infected animals in significantly lower numbers when compared with the wild type strain. Both the mutant and the wild type strain were still able to cross the blood-brain barrier as indicated by the isolation of bacteria from the cerebrospinal fluid. These results indicate that hemoglobin represents an important iron source for *N. meningitidis* during growth *in vivo*.

EXAMPLE 9

Polymerase Chain Reaction Amplification of Hemoglobin Receptor Genes from *N. meningitidis* Serotypes and *N. gonorrhoeae*

From the nucleotide sequence of the 3.3 kb *Bam*HI-*Hind*III DNA fragment carrying the *hmbR* gene and its promoter region was determined specific oligonucleotide promoters for *in vitro* amplification of the homologous hemoglobin receptor protein genes from *N. meningitidis* serotypes A and B and *N. gonorrhoeae* MS11A as follows.

The following oligonucleotide primers were developed for *in vitro* amplification reactions using the polymerase chain reaction (PCR; Saiki *et al.*, 1988, *Science* 230: 1350-1354):

5'-AAACAGGTCTCGGCATAG-3' (sense primer) (SEQ ID No.:11)

5'-CGCGAATTCAAACAGGTCTCGGCATAG-3' (antisense primer) (SEQ ID No.:12)

for amplifying the hemoglobin receptor protein from *N. meningitidis*, serotype A;

5'-CGCGAATTCAAAACTTCCATTCCAGCGATACG-3' (sense primer) (SEQ ID No.:13)

5'-TAAAACTTCCATTCCAGCGATACG-3' (antisense primer) (SEQ ID No.:14)

for amplifying the hemoglobin receptor protein from *N. meningitidis*, serotype B;

5'-AAACAGGTCTCGGCATAG-3' (sense primer) (SEQ ID No.:15)

or

5 5'-CGCGAATTCAAACAGGTCTCGGCATAG-3' (sense primer) (SEQ ID No.:16)

and

5'-CGCGAATTCAAAAACCTTCCATTCCAGCGATACG-3' (SEQ ID No.:17)
(antisense primer)

or

10 5'-TAAAACTTCCATTCCAGCGATACG-3' (antisense primer) (SEQ ID No.:18)

for amplifying the hemoglobin receptor protein from *N. gonorrhoeae* MS11A.

Genomic DNA from *N. meningitidis* serotype A or B or *N. gonorrhoeae* species was prepared using standard techniques (see Sambrook, *et al.*, *ibid.*), including enzymatic degradation of bacterial cell walls, protoplast lysis, protease and RNase digestion, extraction with organic solvents such as phenol and/or chloroform, and ethanol precipitation. Crude DNA preparations were also used. An amount (typically, about 0.1 μ g) of genomic DNA was used for each amplification reaction. A PCR amplification reaction consisted of *Pfu* polymerase (Stratagene, LaJolla, CA) and/or *Taq* polymerase (Boehringer Mannheim, Germany) in the appropriate buffer including about 20picomoles of each amplification primer and 200nanomoles of each deoxynucleoside triphosphate. Amplification reactions were performed according to the following scheme:

First cycle	5 min at 95°C
	2 min at 51°C
	6 min at 72°C
Cycles 2-13	45 sec at 95°C
	35 sec at 49°C
	10 min at 72°C

Cycles 14-30 25 sec at 95°C
 35 sec at 47°C
 10 min at 72°C

Upon completion of the amplification reaction, DNA fragments were cloned either blunt-ended or, after *EcoRI* digestion, into *EcoRI* digested pSUKS or pWKS30 vectors and transformed into bacteria. Positively-selected clones were then analyzed for the presence of recombinant inserts, which were sequenced as described above in Example 4.

As a result of these experiments, three clones encoding the hemoglobin receptor genes from *N. meningitidis* serotypes A and B and *N. gonorrhoeae* MS11A were cloned and the sequence of these genes determined. The nucleic acid sequence for each of these genes are shown in Figures 7 (*N. meningitidis*, serotype A), 8 (*N. meningitidis*, serotype A) and 9 (*N. gonorrhoeae* MS11A).

The degree of homology between the cloned hemoglobin receptors from the different *N. meningitidis* serotypes and *N. gonorrhoeae* MS11A was assessed by nucleic acid and amino acid sequence comparison, as described in Example 5 above. The results of these comparisons are shown in Figures 10 and 11, respectively. Hemoglobin receptor genes from the three *N. meningitidis* serotypes and *N. gonorrhoeae* MS11A were found to be from 86.5% to 93.4% homologous; the most homologous nucleic acids were *N. meningitidis* serotypes B and C, and the most divergent nucleic acids were *N. meningitidis* serotype B and *N. gonorrhoeae* MS11A (Figure 10 and Table III). Hemoglobin receptor proteins from all four *Neisseria* species showed a high degree of homology to the other members of the group, ranging from 87% homology between the hemoglobin receptor proteins from *N. gonorrhoeae* MS11A and *N. meningitidis* serotype B to 93% homology between hemoglobin receptor proteins from *N. meningitidis* serotypes A and B (Figure 11). In this comparison, all four receptors were found to share 84.7% amino acid sequence identity, and up to 11.6% sequence similarity (*i.e.*, chemically-related amino acid residues at homologous sites within the amino acid sequence). The non-conserved amino acids were found clustered in the regions of the amino acid sequence corresponding to the external loops in the predicted topographical structure of the hemoglobin receptor proteins.

TABLE III

*	A	B	C	MS11
A	X	92.2%	93.0%	90.4%
B	93.3%	X	93.4%	86.5%
C	93.2%	93%	X	90.4%
MS11	91.1%	86.8%	91.4%	X

* The numbers in the upper quadrant of the Table (in **boldface**) represent nucleic acid sequence homology between the different hemoglobin receptor genes of the invention, while the numbers in the lower quadrant of the Table represent amino acid sequence homology between the different hemoglobin receptor proteins

It should be understood that the foregoing disclosure emphasizes certain specific embodiments of the invention and that all modifications or alternatives equivalent thereto are within the spirit and scope of the invention as set forth in the appended claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Stojiljkovic, Igor
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Heffron, Fred
Nassif, Xavier
- (ii) TITLE OF INVENTION: Novel Bacterial Hemoglobin Receptor
Genes and Uses
- (iii) NUMBER OF SEQUENCES: 14
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- (v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/537,361
(B) FILING DATE: 02-OCT-1995
(C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Noonan, Kevin E
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(C) REFERENCE/DOCKET NUMBER: 94,784-A
- (ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: 312-913-0001
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(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 3319 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 471..2848

006760" 885960

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

AGAACTAGTG GATCCAATTT GGGCGCGGCG TTTTGTTC	AACACGCCCA AAAACTCGAT	60
TACAACGGCG AACACGGCGC GCGCCACCTC GCTCCGCATC	CCGACGGGCC GCGGCAAACA	120
CTGGCGCGCC TTCGTCGAGC ATCTTGAACG CTTTGAACCT	GACTCCCGAA GCCGAAGCGG	180
AAGCCATTCA AGGCGCGCGC GAAGCCTTTG CATTCTACAA	AGTCGTGTTG CGCGAAACCT	240
TCGGCTTGGC AGCCGATGCC GAAGCCCCCG AAGGTATGAT	GCCGCACAGG CACTAAAAAA	300
TAATCGAACC AAATAAACAA GGTCTCGGCA TAGCTGTTTG	CAGGGACCTT TAATTACAG	360
GCGCGGCTTT GTTTACATGG ATTACTGTCT TATTAAATAT	TAATGATTAT CATAAAATCT	420
ATTATTCGCT AACCGATGGA TGAACAATCC ATACATCTTG	AGTTGATAAT ATG AAA	476
	Met Lys	
	1	
CCA TTA CAA ATG CTC CCT ATC GCC GCG CTG GTC	GGC AGT ATT TTC GGC	524
Pro Leu Gln Met Leu Pro Ile Ala Ala Leu Val	Gly Ser Ile Phe Gly	
5 10 15		
AAT CCG GTC TTT GCG GCA GAT GAA GCT GCA ACT	GAA ACC ACA CCC GTT	572
Asn Pro Val Phe Ala Ala Asp Glu Ala Ala Thr	Glu Thr Thr Pro Val	
20 25 30		
AAG GCA GAG GTA AAA GCA GTG CGC GGT AAA GGC	CAG CGC AAT GCG CCT	620
Lys Ala Glu Val Lys Ala Val Arg Gly Lys Gly	Gln Arg Asn Ala Pro	
35 40 45 50		
GCG GCT GTG GAA CGC GTC AAC CTT AAC CGT ATC	AAA CAA GAA ATG ATA	668
Ala Ala Val Glu Arg Val Asn Leu Asn Arg Ile	Lys Gln Glu Met Ile	
55 60 65		
CGC GAC AAC AAA GAC TTG GTG CGC TAT TCC ACC	GAT GTC GGC TTG AGC	716
Arg Asp Asn Lys Asp Leu Val Arg Tyr Ser Thr	Asp Val Gly Leu Ser	
70 75 80		
GAC AGC GGC CGC CAT CAA AAA GGC TTT GCT GTT	CGC GGC GTG GAA GGC	764
Asp Ser Gly Arg His Gln Lys Gly Phe Ala Val	Arg Gly Val Glu Gly	
85 90 95		
AAC CGT GTC GGC GTG AGC ATA GAC GGC GTA AAC	CTG CCT GAT TCC GAA	812
Asn Arg Val Gly Val Ser Ile Asp Gly Val Asn	Leu Pro Asp Ser Glu	
100 105 110		
GAA AAC TCG CTG TAC GCC CGT TAT GGC AAC TTC	AAC AGC TCG CGT CTG	860
Glu Asn Ser Leu Tyr Ala Arg Tyr Gly Asn Phe	Asn Ser Ser Arg Leu	
115 120 125 130		
TCT ATC GAC CCC GAA CTC GTG CGC AAC ATC GAC	ATC GTA AAA GGG GCG	908
Ser Ile Asp Pro Glu Leu Val Arg Asn Ile Asp	Ile Val Lys Gly Ala	
135 140 145		
GAC TCT TTC AAT ACC GGC AGC GGC GCC TTG GGC	GGC GGT GTG AAT TAC	956
Asp Ser Phe Asn Thr Gly Ser Gly Ala Leu Gly	Gly Gly Val Asn Tyr	

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150	155	160	
CAA ACC CTG CAA GGA CGT GAC TTA CTG TTG CCT GAA CGG CAG TTC GGC Gln Thr Leu Gln Gly Arg Asp Leu Leu Leu Pro Glu Arg Gln Phe Gly 165 170 175			1004
GTG ATG ATG AAA AAC GGT TAC AGC ACG CGT AAC CGT GAA TGG ACA AAT Val Met Met Lys Asn Gly Tyr Ser Thr Arg Asn Arg Glu Trp Thr Asn 180 185 190			1052
ACC CTC GGT TTC GGC GTG AGC AAC GAC CGC GTG GAT GCC GCT TTG CTG Thr Leu Gly Phe Gly Val Ser Asn Asp Arg Val Asp Ala Ala Leu Leu 195 200 205 210			1100
TAT TCG CAA CGG CGC GGC CAT GAA ACT GAA AGC GCG GGC AAG CGT GGT Tyr Ser Gln Arg Arg Gly His Glu Thr Glu Ser Ala Gly Lys Arg Gly 215 220 225			1148
TAT CCG GTA GAG GGT GCT GGT AGC GGA GCG AAT ATC CGT GGT TCT GCG Tyr Pro Val Glu Gly Ala Gly Ser Gly Ala Asn Ile Arg Gly Ser Ala 230 235 240			1196
CGC GGT ATT CCT GAT CCG TCC CAA CAC AAA TAC CAC AGC TTC TTG GGT Arg Gly Ile Pro Asp Pro Ser Gln His Lys Tyr His Ser Phe Leu Gly 245 250 255			1244
AAG ATT GCT TAT CAA ATC AAC GAC AAC CAC CGC ATC GGC GCA TCG CTC Lys Ile Ala Tyr Gln Ile Asn Asp Asn His Arg Ile Gly Ala Ser Leu 260 265 270			1292
AAC GGT CAG CAG GGG CAT AAT TAC ACG GTT GAA GAG TCT TAC AAC CTG Asn Gly Gln Gln Gly His Asn Tyr Thr Val Glu Glu Ser Tyr Asn Leu 275 280 285 290			1340
CTT GCT TCT TAT TGG CGT GAA GCT GAC GAT GTC AAC AGA CGG CGT AAC Leu Ala Ser Tyr Trp Arg Glu Ala Asp Asp Val Asn Arg Arg Arg Asn 295 300 305			1388
ACC AAC CTC TTT TAC GAA TGG ACG CCG GAA TCC GAC CGG TTG TCT ATG Thr Asn Leu Phe Tyr Glu Trp Thr Pro Glu Ser Asp Arg Leu Ser Met 310 315 320			1436
GTA AAA GCG GAT GTC GAT TAT CAA AAA ACC AAA GTA TCT GCG GTC AAC Val Lys Ala Asp Val Asp Tyr Gln Lys Thr Lys Val Ser Ala Val Asn 325 330 335			1484
TAC AAA GGT TCG TTC CCG ATA GAG GAT TCT TCC ACC TTG ACA CGT AAC Tyr Lys Gly Ser Phe Pro Ile Glu Asp Ser Ser Thr Leu Thr Arg Asn 340 345 350			1532
TAC AAT CAA AAG GAC TTG GAT GAA ATC TAC AAC CGC AGT ATG GAT ACC Tyr Asn Gln Lys Asp Leu Asp Glu Ile Tyr Asn Arg Ser Met Asp Thr 355 360 365 370			1580
CGC TTC AAA CGC ATT ACC CTG CGT TTG GAC AGC CAT CCG TTG CAA CTC Arg Phe Lys Arg Ile Thr Leu Arg Leu Asp Ser His Pro Leu Gln Leu 375 380 385			1628
GGG GGG GGG CGA CAC CGC CTG TCG TTT AAA ACT TTC GCC AGC CGC CGT			1676

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Gly	Gly	Gly	Arg	His	Arg	Leu	Ser	Phe	Lys	Thr	Phe	Ala	Ser	Arg	Arg	
			390					395					400			
GAT	TTT	GAA	AAC	CTA	AAC	CGC	GAC	GAT	TAT	TAC	TTC	AGC	GGC	CGT	GTT	1724
Asp	Phe	Glu	Asn	Leu	Asn	Arg	Asp	Asp	Tyr	Tyr		Phe	Ser	Gly	Arg	Val
		405					410					415				
GTT	CGA	ACC	ACC	AGC	AGT	ATC	CAG	CAT	CCG	GTG	AAA	ACC	ACC	AAC	TAC	1772
Val	Arg	Thr	Thr	Ser	Ser	Ile	Gln	His	Pro	Val	Lys	Thr	Thr	Asn	Tyr	
	420					425					430					
GGT	TTC	TCA	CTG	TCT	GAC	CAA	ATT	CAA	TGG	AAC	GAC	GTG	TTC	AGT	AGC	1820
Gly	Phe	Ser	Leu	Ser	Asp	Gln	Ile	Gln	Trp	Asn	Asp	Val	Phe	Ser	Ser	
435					440					445					450	
CGC	GCA	GGT	ATC	CGT	TAC	GAT	CAT	ACC	AAA	ATG	ACG	CCT	CAG	GAA	TTG	1868
Arg	Ala	Gly	Ile	Arg	Tyr	Asp	His	Thr	Lys	Met	Thr	Pro	Gln	Glu	Leu	
				455					460					465		
AAT	GCC	GAG	TGT	CAT	GCT	TGT	GAC	AAA	ACA	CCG	CCT	GCA	GCC	AAC	ACT	1916
Asn	Ala	Glu	Cys	His	Ala	Cys	Asp	Lys	Thr	Pro	Pro	Ala	Ala	Asn	Thr	
			470				475						480			
TAT	AAA	GGC	TGG	AGC	GGT	TTT	GTC	GGC	TTG	GCG	GCG	CAA	CTG	AAT	CAG	1964
Tyr	Lys	Gly	Trp	Ser	Gly	Phe	Val	Gly	Leu	Ala	Ala	Gln	Leu	Asn	Gln	
		485					490					495				
GCT	TGG	CGT	GTC	GGT	TAC	GAC	ATT	ACT	TCC	GGC	TAC	CGT	GTC	CCC	AAT	2012
Ala	Trp	Arg	Val	Gly	Tyr	Asp	Ile	Thr	Ser	Gly	Tyr	Arg	Val	Pro	Asn	
	500					505					510					
GCG	TCC	GAA	GTG	TAT	TTC	ACT	TAC	AAC	CAC	GGT	TCG	GGT	AAT	TGG	CTG	2060
Ala	Ser	Glu	Val	Tyr	Phe	Thr	Tyr	Asn	His	Gly	Ser	Gly	Asn	Trp	Leu	
515					520					525					530	
CCC	AAT	CCC	AAC	CTG	AAA	GCC	GAG	CGC	ACG	ACC	ACC	CAC	ACC	CTC	TCT	2108
Pro	Asn	Pro	Asn	Leu	Lys	Ala	Glu	Arg	Thr	Thr	Thr	His	Thr	Leu	Ser	
				535					540					545		
CTG	CAA	GGC	CGC	AGC	GAA	AAA	GGT	ACT	TTG	GAT	GCC	AAC	CTG	TAT	CAA	2156
Leu	Gln	Gly	Arg	Ser	Glu	Lys	Gly	Thr	Leu	Asp	Ala	Asn	Leu	Tyr	Gln	
			550				555						560			
AGC	AAT	TAC	CGC	AAT	TTC	CTG	TCT	GAA	GAG	CAG	AAG	CTG	ACC	ACC	AGC	2204
Ser	Asn	Tyr	Arg	Asn	Phe	Leu	Ser	Glu	Glu	Gln	Lys	Leu	Thr	Thr	Ser	
		565					570					575				
GGC	GAT	GTC	AGC	TGT	ACT	CAG	ATG	AAT	TAC	TAC	TAC	GGT	ATG	TGT	AGC	2252
Gly	Asp	Val	Ser	Cys	Thr	Gln	Met	Asn	Tyr	Tyr	Tyr	Gly	Met	Cys	Ser	
	580					585					590					
AAT	CCT	TAT	TCC	GAA	AAA	CTG	GAA	TGG	CAG	ATG	CAA	AAT	ATC	GAC	AAG	2300
Asn	Pro	Tyr	Ser	Glu	Lys	Leu	Glu	Trp	Gln	Met	Gln	Asn	Ile	Asp	Lys	
595					600					605				610		
GCC	AGA	ATC	CGC	GGT	ATC	GAG	CTG	ACG	GGC	CGT	CTG	AAT	GTG	GAC	AAA	2348
Ala	Arg	Ile	Arg	Gly	Ile	Glu	Leu	Thr	Gly	Arg	Leu	Asn	Val	Asp	Lys	
				615					620					625		

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GTA GCG TCT TTT GTT CCT GAG GGC TGG AAA CTG TTC GGC TCG CTG GGT Val Ala Ser Phe Val Pro Glu Gly Trp Lys Leu Phe Gly Ser Leu Gly 630 635 640	2396
TAT GCG AAA AGC AAA CTG TCG GGC GAC AAC AGC CTG CTG TTC ACC CAG Tyr Ala Lys Ser Lys Leu Ser Gly Asp Asn Ser Leu Leu Phe Thr Gln 645 650 655	2444
CCG TTG AAA GTG ATT GCC GGT ATC GAC TAT GAA AGT CCG AGC GAA AAA Pro Leu Lys Val Ile Ala Gly Ile Asp Tyr Glu Ser Pro Ser Glu Lys 660 665 670	2492
TGG GGC GTG TTC TCC CGC CTG ACC TAT CTG GGC GCG AAA AAG GTC AAA Trp Gly Val Phe Ser Arg Leu Thr Tyr Leu Gly Ala Lys Lys Val Lys 675 680 685 690	2540
GAC GCG CAA TAC ACC GTT TAT GAA AAC AAG GGC TGG GGT ACG CCT TTG Asp Ala Gln Tyr Thr Val Tyr Glu Asn Lys Gly Trp Gly Thr Pro Leu 695 700 705	2588
CAG AAA AAG GTA AAA GAT TAC CCG TGG CTG AAC AAG TCG GCT TAT GTG Gln Lys Lys Val Lys Asp Tyr Pro Trp Leu Asn Lys Ser Ala Tyr Val 710 715 720	2636
TTC GAT ATG TAC GGC TTC TAC AAA CCG GTG AAA AAC CTG ACT TTG CGT Phe Asp Met Tyr Gly Phe Tyr Lys Pro Val Lys Asn Leu Thr Leu Arg 725 730 735	2684
GCA GGC GTA TAT AAT GTG TTC AAC CGC AAA TAC ACC ACT TGG GAT TCC Ala Gly Val Tyr Asn Val Phe Asn Arg Lys Tyr Thr Thr Trp Asp Ser 740 745 750	2732
CTG CGC GGC CTG TAT AGC TAC AGC ACC ACC AAC TCG GTC GAC CGC GAT Leu Arg Gly Leu Tyr Ser Tyr Ser Thr Thr Asn Ser Val Asp Arg Asp 755 760 765 770	2780
GGC AAA GGC TTA GAC CGC TAC CGC GCC CCA AGC CGT AAT TAC GCC GTA Gly Lys Gly Leu Asp Arg Tyr Arg Ala Pro Ser Arg Asn Tyr Ala Val 775 780 785	2828
TCG CTG GAA TGG AAG TTT TA ATCTGGTATT ATTGAATTAA TCGCCTTGTT Ser Leu Glu Trp Lys Phe 790	2878
GAAAATTAAA GCCGTCCGAA TTGTGTTCAA GAACTCATTC GGACGGTTTT TACCGAATCT	2938
GTGTGTGGGT TTATAGTGGA TTAACAAAAA TCAGGACAAG GCGACGAAGC CGCAGACAGT	2998
ACAGATAGTA CGGAACCGAT TCACTTGGTG AGACCTTTGC AAAATTCCTT TCCCTCCCGA	3058
CAGCCGAAAC CCAAACACAG GTTTTCGGCT GTTTTCGCCC CAAATACCTC CTAATTCTAC	3118
CCAAATACCC CCTTAATCCT CCCCATAACC CGATAATCAG GCATCCGGCG CCTTTAGGCG	3178
GCAGCGGGCG CACTTAACCT GTTGGCGGCT TTCAAAAGGT TCAAACACAT CGCCTTCAGG	3238
TGCCTTTGCG CACTCACTTT AATCAGTCCG AAATAGGCCG CCCGCGCATA GCAGAACTTA	3298
CGGTGCAGCG TACCGAAGCT T	3319

006760-856960

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 792 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Lys Pro Leu Gln Met Leu Pro Ile Ala Ala Leu Val Gly Ser Ile
1 5 10 15
Phe Gly Asn Pro Val Phe Ala Ala Asp Glu Ala Ala Thr Glu Thr Thr
20 25 30
Pro Val Lys Ala Glu Val Lys Ala Val Arg Gly Lys Gly Gln Arg Asn
35 40 45
Ala Pro Ala Ala Val Glu Arg Val Asn Leu Asn Arg Ile Lys Gln Glu
50 55 60
Met Ile Arg Asp Asn Lys Asp Leu Val Arg Tyr Ser Thr Asp Val Gly
65 70 75 80
Leu Ser Asp Ser Gly Arg His Gln Lys Gly Phe Ala Val Arg Gly Val
85 90 95
Glu Gly Asn Arg Val Gly Val Ser Ile Asp Gly Val Asn Leu Pro Asp
100 105 110
Ser Glu Glu Asn Ser Leu Tyr Ala Arg Tyr Gly Asn Phe Asn Ser Ser
115 120 125
Arg Leu Ser Ile Asp Pro Glu Leu Val Arg Asn Ile Asp Ile Val Lys
130 135 140
Gly Ala Asp Ser Phe Asn Thr Gly Ser Gly Ala Leu Gly Gly Gly Val
145 150 155 160
Asn Tyr Gln Thr Leu Gln Gly Arg Asp Leu Leu Leu Pro Glu Arg Gln
165 170 175
Phe Gly Val Met Met Lys Asn Gly Tyr Ser Thr Arg Asn Arg Glu Trp
180 185 190
Thr Asn Thr Leu Gly Phe Gly Val Ser Asn Asp Arg Val Asp Ala Ala
195 200 205
Leu Leu Tyr Ser Gln Arg Arg Gly His Glu Thr Glu Ser Ala Gly Lys
210 215 220
Arg Gly Tyr Pro Val Glu Gly Ala Gly Ser Gly Ala Asn Ile Arg Gly
225 230 235 240
Ser Ala Arg Gly Ile Pro Asp Pro Ser Gln His Lys Tyr His Ser Phe

006760"833300

			245					250					255		
Leu	Gly	Lys	Ile 260	Ala	Tyr	Gln	Ile	Asn 265	Asp	Asn	His	Arg	Ile 270	Gly	Ala
Ser	Leu	Asn 275	Gly	Gln	Gln	Gly	His 280	Asn	Tyr	Thr	Val	Glu 285	Glu	Ser	Tyr
Asn	Leu 290	Leu	Ala	Ser	Tyr	Trp 295	Arg	Glu	Ala	Asp	Asp 300	Val	Asn	Arg	Arg
Arg 305	Asn	Thr	Asn	Leu	Phe 310	Tyr	Glu	Trp	Thr	Pro 315	Glu	Ser	Asp	Arg	Leu 320
Ser	Met	Val	Lys	Ala 325	Asp	Val	Asp	Tyr	Gln 330	Lys	Thr	Lys	Val	Ser 335	Ala
Val	Asn	Tyr	Lys 340	Gly	Ser	Phe	Pro	Ile 345	Glu	Asp	Ser	Ser	Thr 350	Leu	Thr
Arg	Asn	Tyr 355	Asn	Gln	Lys	Asp	Leu 360	Asp	Glu	Ile	Tyr	Asn 365	Arg	Ser	Met
Asp	Thr 370	Arg	Phe	Lys	Arg	Ile 375	Thr	Leu	Arg	Leu	Asp 380	Ser	His	Pro	Leu
Gln 385	Leu	Gly	Gly	Gly	Arg 390	His	Arg	Leu	Ser	Phe 395	Lys	Thr	Phe	Ala	Ser 400
Arg	Arg	Asp	Phe	Glu 405	Asn	Leu	Asn	Arg	Asp 410	Asp	Tyr	Tyr	Phe	Ser 415	Gly
Arg	Val	Val	Arg 420	Thr	Thr	Ser	Ser	Ile 425	Gln	His	Pro	Val	Lys 430	Thr	Thr
Asn	Tyr	Gly 435	Phe	Ser	Leu	Ser	Asp 440	Gln	Ile	Gln	Trp	Asn 445	Asp	Val	Phe
Ser	Ser 450	Arg	Ala	Gly	Ile	Arg 455	Tyr	Asp	His	Thr	Lys 460	Met	Thr	Pro	Gln
Glu 465	Leu	Asn	Ala	Glu	Cys 470	His	Ala	Cys	Asp	Lys 475	Thr	Pro	Pro	Ala	Ala 480
Asn	Thr	Tyr	Lys	Gly 485	Trp	Ser	Gly	Phe	Val 490	Gly	Leu	Ala	Ala	Gln 495	Leu
Asn	Gln	Ala	Trp 500	Arg	Val	Gly	Tyr	Asp 505	Ile	Thr	Ser	Gly	Tyr 510	Arg	Val
Pro	Asn	Ala 515	Ser	Glu	Val	Tyr	Phe 520	Thr	Tyr	Asn	His	Gly 525	Ser	Gly	Asn
Trp	Leu 530	Pro	Asn	Pro	Asn	Leu 535	Lys	Ala	Glu	Arg	Thr 540	Thr	Thr	His	Thr
Leu 545	Ser	Leu	Gln	Gly	Arg 550	Ser	Glu	Lys	Gly	Thr 555	Leu	Asp	Ala	Asn	Leu 560

Tyr Gln Ser Asn Tyr Arg Asn Phe Leu Ser Glu Glu Gln Lys Leu Thr
 565 570 575
 Thr Ser Gly Asp Val Ser Cys Thr Gln Met Asn Tyr Tyr Tyr Gly Met
 580 585 590
 Cys Ser Asn Pro Tyr Ser Glu Lys Leu Glu Trp Gln Met Gln Asn Ile
 595 600 605
 Asp Lys Ala Arg Ile Arg Gly Ile Glu Leu Thr Gly Arg Leu Asn Val
 610 615 620
 Asp Lys Val Ala Ser Phe Val Pro Glu Gly Trp Lys Leu Phe Gly Ser
 625 630 635 640
 Leu Gly Tyr Ala Lys Ser Lys Leu Ser Gly Asp Asn Ser Leu Leu Phe
 645 650 655
 Thr Gln Pro Leu Lys Val Ile Ala Gly Ile Asp Tyr Glu Ser Pro Ser
 660 665 670
 Glu Lys Trp Gly Val Phe Ser Arg Leu Thr Tyr Leu Gly Ala Lys Lys
 675 680 685
 Val Lys Asp Ala Gln Tyr Thr Val Tyr Glu Asn Lys Gly Trp Gly Thr
 690 695 700
 Pro Leu Gln Lys Lys Val Lys Asp Tyr Pro Trp Leu Asn Lys Ser Ala
 705 710 715 720
 Tyr Val Phe Asp Met Tyr Gly Phe Tyr Lys Pro Val Lys Asn Leu Thr
 725 730 735
 Leu Arg Ala Gly Val Tyr Asn Val Phe Asn Arg Lys Tyr Thr Thr Trp
 740 745 750
 Asp Ser Leu Arg Gly Leu Tyr Ser Tyr Ser Thr Thr Asn Ser Val Asp
 755 760 765
 Arg Asp Gly Lys Gly Leu Asp Arg Tyr Arg Ala Pro Ser Arg Asn Tyr
 770 775 780
 Ala Val Ser Leu Glu Trp Lys Phe
 785 790

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2376 base pairs
- (B) TYPE: nucleic acid

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(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 1..2373

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ATG AAA CCA TTA CAA ATG CCC CCT ATC GCC GCG CTG CTC GGC AGT ATT	48
Met Lys Pro Leu Gln Met Pro Pro Ile Ala Ala Leu Leu Gly Ser Ile	
1 5 10 15	
TTC GGC AAT CCG GTC TTT GCG GCA GAT GAA GCT GCA ACT GAA ACC ACA	96
Phe Gly Asn Pro Val Phe Ala Ala Asp Glu Ala Ala Thr Glu Thr Thr	
20 25 30	
CCC GTT AAG GCA GAG GTA AAA GCA GTG CGC GTT AAA GGT CAG CGC AAT	144
Pro Val Lys Ala Glu Val Lys Ala Val Arg Val Lys Gly Gln Arg Asn	
35 40 45	
GCG CCT GCG GCT GTG GAA CGC GTC AAC CTT AAC CGT ATC AAA CAA GAA	192
Ala Pro Ala Ala Val Glu Arg Val Asn Leu Asn Arg Ile Lys Gln Glu	
50 55 60	
ATG ATA CGC GAC AAT AAA GAC TTG GTG CGC TAT TCC ACC GAT GTC GGC	240
Met Ile Arg Asp Asn Lys Asp Leu Val Arg Tyr Ser Thr Asp Val Gly	
65 70 75 80	
TTG AGC GAC AGG AGC CGT CAT CAA AAA GGC TTT GCC ATT CGC GGC GTG	288
Leu Ser Asp Arg Ser Arg His Gln Lys Gly Phe Ala Ile Arg Gly Val	
85 90 95	
GAA GGC GAC CGT GTC GGC GTT AGT ATT GAC GGC GTA AAC CTG CCT GAT	336
Glu Gly Asp Arg Val Gly Val Ser Ile Asp Gly Val Asn Leu Pro Asp	
100 105 110	
TCC GAA GAA AAC TCG CTG TAC GCC CGT TAT GGC AAC TTC AAC AGC TCG	384
Ser Glu Glu Asn Ser Leu Tyr Ala Arg Tyr Gly Asn Phe Asn Ser Ser	
115 120 125	
CGT CTG TCT ATC GAC CCC GAA CTC GTG CGC AAC ATC GAC ATC GTA AAA	432
Arg Leu Ser Ile Asp Pro Glu Leu Val Arg Asn Ile Asp Ile Val Lys	
130 135 140	
GGG GCG GAC TCT TTC AAT ACC GGC AGC GGC GCC TTG GGC GGC GGT GTG	480
Gly Ala Asp Ser Phe Asn Thr Gly Ser Gly Ala Leu Gly Gly Gly Val	
145 150 155 160	
AAT TAC CAA ACC CTG CAA GGA CGT GAC TTA CTG TTG CCT GAA CGG CAG	528
Asn Tyr Gln Thr Leu Gln Gly Arg Asp Leu Leu Leu Pro Glu Arg Gln	
165 170 175	
TTC GGC GTG ATG ATG AAA AAC GGT TAC AGC ACG CGT AAC CGT GAA TGG	576
Phe Gly Val Met Met Lys Asn Gly Tyr Ser Thr Arg Asn Arg Glu Trp	
180 185 190	

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ACA	AAT	ACC	CTC	GGT	TTC	GGC	GTG	AGC	AAC	GAC	CGC	GTG	GAT	GCC	GCT	624
Thr	Asn	Thr	Leu	Gly	Phe	Gly	Val	Ser	Asn	Asp	Arg	Val	Asp	Ala	Ala	
	195						200					205				
TTG	CTG	TAT	TCG	CAA	CGG	CGC	GGC	CAT	GAA	ACT	GAA	AGC	GCG	GGC	AAG	672
Leu	Leu	Tyr	Ser	Gln	Arg	Arg	Gly	His	Glu	Thr	Glu	Ser	Ala	Gly	Lys	
	210					215					220					
CGT	GGT	TAT	CCG	GTA	GAG	GGT	GCT	GGT	AGC	GGA	GCG	AAT	ATC	CGT	GGT	720
Arg	Gly	Tyr	Pro	Val	Glu	Gly	Ala	Gly	Ser	Gly	Ala	Asn	Ile	Arg	Gly	
225					230					235					240	
TCT	GCG	CGC	GGT	ATT	CCT	GAT	CCG	TCC	CAA	CAC	AAA	TAC	CAC	AGC	TTC	768
Ser	Ala	Arg	Gly	Ile	Pro	Asp	Pro	Ser	Gln	His	Lys	Tyr	His	Ser	Phe	
			245						250					255		
TTG	GGT	AAG	ATT	GCT	TAT	CAA	ATC	AAC	GAC	AAC	CAC	CGC	ATC	GGC	GCA	816
Leu	Gly	Lys	Ile	Ala	Tyr	Gln	Ile	Asn	Asp	Asn	His	Arg	Ile	Gly	Ala	
			260					265					270			
TCG	CTC	AAC	GGT	CAG	CAG	GGG	CAT	AAT	TAC	ACG	GTT	GAA	GAG	TCT	TAC	864
Ser	Leu	Asn	Gly	Gln	Gln	Gly	His	Asn	Tyr	Thr	Val	Glu	Glu	Ser	Tyr	
		275					280					285				
AAC	CTG	CTT	GCT	TCT	TAT	TGG	CGT	GAA	GCT	GAC	GAT	GTC	AAC	AGA	CGG	912
Asn	Leu	Leu	Ala	Ser	Tyr	Trp	Arg	Glu	Ala	Asp	Asp	Val	Asn	Arg	Arg	
	290					295					300					
CGT	AAC	ACC	PAC	CTC	TTT	TAC	GAA	TGG	ACG	CCG	GAA	TCC	GAC	CGG	TTG	960
Arg	Asn	Thr	Asn	Leu	Phe	Tyr	Glu	Trp	Thr	Pro	Glu	Ser	Asp	Arg	Leu	
305					310					315					320	
TCT	ATG	GTA	AAA	GCG	GAT	GTC	GAT	TAT	CAA	AAA	ACC	AAA	GTA	TCT	GCG	1008
Ser	Met	Val	Lys	Ala	Asp	Val	Asp	Tyr	Gln	Lys	Thr	Lys	Val	Ser	Ala	
			325						330					335		
GTC	AAC	TAC	AAA	GGT	TCG	TTC	CCG	ACG	AAT	TAC	ACC	ACA	TGG	GAA	ACC	1056
Val	Asn	Tyr	Lys	Gly	Ser	Phe	Pro	Thr	Asn	Tyr	Thr	Thr	Trp	Glu	Thr	
			340					345					350			
GAG	TAC	CAT	AAA	AAG	GAA	GTT	GGC	GAA	ATC	TAT	AAC	CGC	AGC	ATG	GAT	1104
Glu	Tyr	His	Lys	Lys	Glu	Val	Gly	Glu	Ile	Tyr	Asn	Arg	Ser	Met	Asp	
		355					360					365				
ACA	ACC	TTC	AAA	CGT	ATT	ACG	CTG	CGT	ATG	GAC	AGC	CAT	CCG	TTG	CAA	1152
Thr	Thr	Phe	Lys	Arg	Ile	Thr	Leu	Arg	Met	Asp	Ser	His	Pro	Leu	Gln	
	370					375					380					
CTC	GGG	GGG	GGG	CGA	CAC	CGC	CTG	TCG	TTT	AAA	ACC	TTT	GCC	GGG	CAG	1200
Leu	Gly	Gly	Gly	Arg	His	Arg	Leu	Ser	Phe	Lys	Thr	Phe	Ala	Gly	Gln	
385					390					395					400	
CGT	GAT	TTT	GAA	AAC	TTA	AAC	CGC	GAC	GAT	TAC	TAC	TTC	AGC	GGC	CGT	1248
Arg	Asp	Phe	Glu	Asn	Leu	Asn	Arg	Asp	Asp	Tyr	Tyr	Phe	Ser	Gly	Arg	
				405					410					415		
GTT	GTT	CGA	ACC	ACC	AAC	AGT	ATC	CAG	CAT	CCG	GTG	AAA	ACC	ACC	AAC	1296
Val	Val	Arg	Thr	Thr	Asn	Ser	Ile	Gln	His	Pro	Val	Lys	Thr	Thr	Asn	
			420					425					430			

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TAC	GGT	TTC	TCG	CTG	TCC	GAC	CAA	ATC	CAA	TGG	AAC	GAC	GTG	TTC	AGT	1344
Tyr	Gly	Phe	Ser	Leu	Ser	Asp	Gln	Ile	Gln	Trp	Asn	Asp	Val	Phe	Ser	
		435					440					445				
AGC	CGC	GCA	GGT	ATC	CGT	TAC	GAC	CAC	ACC	AAA	ATG	ACG	CCT	CAG	GAA	1392
Ser	Arg	Ala	Gly	Ile	Arg	Tyr	Asp	His	Thr	Lys	Met	Thr	Pro	Gln	Glu	
	450					455					460					
TTG	AAT	GCC	GAC	TGT	CAT	GCT	TGT	GAC	AAA	ACA	CCG	CCT	GCA	GCC	AAC	1440
Leu	Asn	Ala	Asp	Cys	His	Ala	Cys	Asp	Lys	Thr	Pro	Pro	Ala	Ala	Asn	
465					470					475					480	
ACT	TAT	AAA	GGC	TGG	AGC	GGA	TTT	GTC	GGC	TTG	GCG	GCG	CAG	CTG	AGC	1488
Thr	Tyr	Lys	Gly	Trp	Ser	Gly	Phe	Val	Gly	Leu	Ala	Ala	Gln	Leu	Ser	
			485						490					495		
CAA	ACA	TGG	CGT	TTG	GGT	TAC	GAT	GTG	ACC	TCA	GGT	TTC	CGC	GTG	CCG	1536
Gln	Thr	Trp	Arg	Leu	Gly	Tyr	Asp	Val	Thr	Ser	Gly	Phe	Arg	Val	Pro	
			500					505					510			
AAT	GCG	TCT	GAA	GTG	TAT	TTC	ACT	TAC	AAC	CAC	GGT	TCG	GGC	ACT	TGG	1584
Asn	Ala	Ser	Glu	Val	Tyr	Phe	Thr	Tyr	Asn	His	Gly	Ser	Gly	Thr	Trp	
		515					520					525				
AAG	CCT	AAT	CCT	AAT	TTG	AAG	GCA	GAA	CGC	AGC	ACC	ACC	CAC	ACC	CTG	1632
Lys	Pro	Asn	Pro	Asn	Leu	Lys	Ala	Glu	Arg	Ser	Thr	Thr	His	Thr	Leu	
	530					535					540					
TCC	TTG	CAG	GGG	CGC	GGC	GAC	AAA	GGG	ACA	CTG	GAT	GCC	AAC	CTG	TAT	1680
Ser	Leu	Gln	Gly	Arg	Gly	Asp	Lys	Gly	Thr	Leu	Asp	Ala	Asn	Leu	Tyr	
545					550					555					560	
CAA	AGC	AAT	TAC	CGA	AAC	TTC	CTG	TCG	GAA	GAG	CAG	AAT	CTG	ACT	GTC	1728
Gln	Ser	Asn	Tyr	Arg	Asn	Phe	Leu	Ser	Glu	Glu	Gln	Asn	Leu	Thr	Val	
				565					570					575		
AGC	GGC	ACA	CCC	GGC	TGT	ACT	GAG	GAG	GAT	GCT	TAC	TAC	TAT	AGA	TGC	1776
Ser	Gly	Thr	Pro	Gly	Cys	Thr	Glu	Glu	Asp	Ala	Tyr	Tyr	Tyr	Arg	Cys	
			580					585					590			
AGC	GAC	CCC	TAC	AAA	GAA	AAA	CTG	GAT	TGG	CAG	ATG	AAA	AAT	ATC	GAC	1824
Ser	Asp	Pro	Tyr	Lys	Glu	Lys	Leu	Asp	Trp	Gln	Met	Lys	Asn	Ile	Asp	
		595					600					605				
AAG	GCC	AGA	ATC	CGC	GGT	ATC	GAG	TTG	ACA	GGC	CGT	CTG	AAT	GTG	GAC	1872
Lys	Ala	Arg	Ile	Arg	Gly	Ile	Glu	Leu	Thr	Gly	Arg	Leu	Asn	Val	Asp	
	610					615					620					
AAA	GTA	GCG	TCT	TTT	GTT	CCT	GAG	GGT	TGG	AAA	CTG	TTC	GGC	TCG	CTG	1920
Lys	Val	Ala	Ser	Phe	Val	Pro	Glu	Gly	Trp	Lys	Leu	Phe	Gly	Ser	Leu	
625					630					635					640	
GGT	TAT	GCG	AAA	AGC	AAA	CTG	TCG	GGC	GAC	AAC	AGC	CTG	CTG	TCC	ACA	1968
Gly	Tyr	Ala	Lys	Ser	Lys	Leu	Ser	Gly	Asp	Asn	Ser	Leu	Leu	Ser	Thr	
				645					650					655		
CAG	CCG	CTG	AAA	GTG	ATT	GCC	GGT	ATC	GAC	TAT	GAA	AGT	CCG	AGC	GAA	2016
Gln	Pro	Leu	Lys	Val	Ile	Ala	Gly	Ile	Asp	Tyr	Glu	Ser	Pro	Ser	Glu	
			660					665					670			

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AAA TGG GGC GTA TTC TCC CGC CTG ACC TAT CTA GGC GCG AAA AAG GTC	2064
Lys Trp Gly Val Phe Ser Arg Leu Thr Tyr Leu Gly Ala Lys Lys Val	
675 680 685	
AAA GAC GCG CAA TAC ACC GTT TAT GAA AAC AAG GGC TGG GGT ACG CCT	2112
Lys Asp Ala Gln Tyr Thr Val Tyr Glu Asn Lys Gly Trp Gly Thr Pro	
690 695 700	
TTG CAG AAA AAG GTA AAA GAT TAC CCG TGG CTG AAC AAG TCG GCT TAT	2160
Leu Gln Lys Lys Val Lys Asp Tyr Pro Trp Leu Asn Lys Ser Ala Tyr	
705 710 715 720	
GTG TTT GAT ATG TAC GGC TTC TAC AAA CCG GCT AAA AAC CTG ACT TTG	2208
Val Phe Asp Met Tyr Gly Phe Tyr Lys Pro Ala Lys Asn Leu Thr Leu	
725 730 735	
CGT GCA GGC GTG TAC AAC CTG TTC AAC CGC AAA TAC ACC ACT TGG GAT	2256
Arg Ala Gly Val Tyr Asn Leu Phe Asn Arg Lys Tyr Thr Thr Trp Asp	
740 745 750	
TCC CTG CGC GGT TTA TAT AGC TAC AGC ACC ACC AAT GCG GTC GAC CGC	2304
Ser Leu Arg Gly Leu Tyr Ser Tyr Ser Thr Thr Asn Ala Val Asp Arg	
755 760 765	
GAT GGC AAA GGC TTA GAC CGC TAC CGC GCC CCA GGC CGC AAT TAC GCC	2352
Asp Gly Lys Gly Leu Asp Arg Tyr Arg Ala Pro Gly Arg Asn Tyr Ala	
770 775 780	
GTA TCG CTG GAA TGG AAG TTT TAA	2376
Val Ser Leu Glu Trp Lys Phe	
785 790	

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 791 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Lys Pro Leu Gln Met Pro Pro Ile Ala Ala Leu Leu Gly Ser Ile	
1 5 10 15	
Phe Gly Asn Pro Val Phe Ala Ala Asp Glu Ala Ala Thr Glu Thr Thr	
20 25 30	
Pro Val Lys Ala Glu Val Lys Ala Val Arg Val Lys Gly Gln Arg Asn	
35 40 45	
Ala Pro Ala Ala Val Glu Arg Val Asn Leu Asn Arg Ile Lys Gln Glu	
50 55 60	
Met Ile Arg Asp Asn Lys Asp Leu Val Arg Tyr Ser Thr Asp Val Gly	
65 70 75 80	

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Leu	Ser	Asp	Arg	Ser 85	Arg	His	Gln	Lys	Gly 90	Phe	Ala	Ile	Arg	Gly 95	Val
Glu	Gly	Asp	Arg	Val 100	Gly	Val	Ser	Ile 105	Asp	Gly	Val	Asn	Leu 110	Pro	Asp
Ser	Glu	Glu	Asn	Ser 115	Leu	Tyr	Ala 120	Arg	Tyr	Gly	Asn	Phe 125	Asn	Ser	Ser
Arg	Leu	Ser	Ile	Asp 130	Pro	Glu 135	Leu	Val	Arg	Asn	Ile 140	Asp	Ile	Val	Lys
Gly 145	Ala	Asp	Ser	Phe 150	Asn	Thr	Gly	Ser	Gly	Ala 155	Leu	Gly	Gly	Gly	Val 160
Asn	Tyr	Gln	Thr	Leu 165	Gln	Gly	Arg	Asp 170	Leu	Leu	Leu	Pro	Glu	Arg 175	Gln
Phe	Gly	Val	Met 180	Met	Lys	Asn	Gly	Tyr 185	Ser	Thr	Arg	Asn	Arg 190	Glu	Trp
Thr	Asn	Thr 195	Leu	Gly	Phe	Gly	Val 200	Ser	Asn	Asp	Arg	Val 205	Asp	Ala	Ala
Leu 210	Leu	Tyr	Ser	Gln	Arg	Arg 215	Gly	His	Glu	Thr	Glu 220	Ser	Ala	Gly	Lys
Arg 225	Gly	Tyr	Pro	Val 230	Glu	Gly	Ala	Gly	Ser	Gly 235	Ala	Asn	Ile	Arg	Gly 240
Ser	Ala	Arg	Gly	Ile 245	Pro	Asp	Pro	Ser	Gln 250	His	Lys	Tyr	His	Ser 255	Phe
Leu	Gly	Lys	Ile 260	Ala	Tyr	Gln	Ile	Asn 265	Asp	Asn	His	Arg	Ile 270	Gly	Ala
Ser	Leu	Asn 275	Gly	Gln	Gln	Gly	His 280	Asn	Tyr	Thr	Val	Glu 285	Glu	Ser	Tyr
Asn 290	Leu	Leu	Ala	Ser	Tyr	Trp 295	Arg	Glu	Ala	Asp	Asp 300	Val	Asn	Arg	Arg
Arg 305	Asn	Thr	Asn	Leu 310	Phe	Tyr	Glu	Trp	Thr	Pro 315	Glu	Ser	Asp	Arg	Leu 320
Ser	Met	Val	Lys 325	Ala	Asp	Val	Asp	Tyr	Gln 330	Lys	Thr	Lys	Val	Ser 335	Ala
Val	Asn	Tyr	Lys 340	Gly	Ser	Phe	Pro	Thr 345	Asn	Tyr	Thr	Thr	Trp 350	Glu	Thr
Glu	Tyr	His 355	Lys	Lys	Glu	Val	Gly 360	Glu	Ile	Tyr	Asn	Arg 365	Ser	Met	Asp
Thr	Thr 370	Phe	Lys	Arg	Ile	Thr 375	Leu	Arg	Met	Asp	Ser 380	His	Pro	Leu	Gln
Leu 385	Gly	Gly	Gly	Arg 390	His	Arg	Leu	Ser	Phe	Lys 395	Thr	Phe	Ala	Gly	Gln 400

Arg Asp Phe Clu Asn Leu Asn Arg Asp Asp Tyr Tyr Phe Ser Gly Arg
405 410 415

Val Val Arg Thr Thr Asn Ser Ile Gln His Pro Val Lys Thr Thr Asn
420 425 430

Tyr Gly Phe Ser Leu Ser Asp Gln Ile Gln Trp Asn Asp Val Phe Ser
435 440 445

Ser Arg Ala Gly Ile Arg Tyr Asp His Thr Lys Met Thr Pro Gln Glu
450 455 460

Leu Asn Ala Asp Cys His Ala Cys Asp Lys Thr Pro Pro Ala Ala Asn
465 470 475 480

Thr Tyr Lys Gly Trp Ser Gly Phe Val Gly Leu Ala Ala Gln Leu Ser
485 490 495

Gln Thr Trp Arg Leu Gly Tyr Asp Val Thr Ser Gly Phe Arg Val Pro
500 505 510

Asn Ala Ser Glu Val Tyr Phe Thr Tyr Asn His Gly Ser Gly Thr Trp
515 520 525

Lys Pro Asn Pro Asn Leu Lys Ala Glu Arg Ser Thr Thr His Thr Leu
530 535 540

Ser Leu Gln Gly Arg Gly Asp Lys Gly Thr Leu Asp Ala Asn Leu Tyr
545 550 555 560

Gln Ser Asn Tyr Arg Asn Phe Leu Ser Glu Glu Gln Asn Leu Thr Val
565 570 575

Ser Gly Thr Pro Gly Cys Thr Glu Glu Asp Ala Tyr Tyr Tyr Arg Cys
580 585 590

Ser Asp Pro Tyr Lys Glu Lys Leu Asp Trp Gln Met Lys Asn Ile Asp
595 600 605

Lys Ala Arg Ile Arg Gly Ile Glu Leu Thr Gly Arg Leu Asn Val Asp
610 615 620

Lys Val Ala Ser Phe Val Pro Glu Gly Trp Lys Leu Phe Gly Ser Leu
625 630 635 640

Gly Tyr Ala Lys Ser Lys Leu Ser Gly Asp Asn Ser Leu Leu Ser Thr
645 650 655

Gln Pro Leu Lys Val Ile Ala Gly Ile Asp Tyr Glu Ser Pro Ser Glu
660 665 670

Lys Trp Gly Val Phe Ser Arg Leu Thr Tyr Leu Gly Ala Lys Lys Val
675 680 685

Lys Asp Ala Gln Tyr Thr Val Tyr Glu Asn Lys Gly Trp Gly Thr Pro
690 695 700

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Leu Gln Lys Lys Val Lys Asp Tyr Pro Trp Leu Asn Lys Ser Ala Tyr
705 710 715 720

Val Phe Asp Met Tyr Gly Phe Tyr Lys Pro Ala Lys Asn Leu Thr Leu
725 730 735

Arg Ala Gly Val Tyr Asn Leu Phe Asn Arg Lys Tyr Thr Thr Trp Asp
740 745 750

Ser Leu Arg Gly Leu Tyr Ser Tyr Ser Thr Thr Asn Ala Val Asp Arg
755 760 765

Asp Gly Lys Gly Leu Asp Arg Tyr Arg Ala Pro Gly Arg Asn Tyr Ala
770 775 780

Val Ser Leu Glu Trp Lys Phe
785 790

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2379 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 1..2376

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

ATG AAA CCA TTA CAA ATG CTC CCT ATC GCC GCG CTG GTC GGC AGT ATT	48
Met Lys Pro Leu Gln Met Leu Pro Ile Ala Ala Leu Val Gly Ser Ile	
1 5 10 15	
TTC GGC AAT CCG GTC TTT GCG GCA GAT GAA GCT GCA ACT GAA ACC ACA	96
Phe Gly Asn Pro Val Phe Ala Ala Asp Glu Ala Ala Thr Glu Thr Thr	
20 25 30	
CCC GTT AAG GCA GAG GTA AAA GCA GTG CGC GTT AAA GGC CAG CGC AAT	144
Pro Val Lys Ala Glu Val Lys Ala Val Arg Val Lys Gly Gln Arg Asn	
35 40 45	
GCG CCT GCG GCT GTG GAA CGC GTC AAC CTT AAC CGT ATC AAA CAA GAA	192
Ala Pro Ala Ala Val Glu Arg Val Asn Leu Asn Arg Ile Lys Gln Glu	
50 55 60	
ATG ATA CGC GAC AAC AAA GAC TTG GTG CGC TAT TCC ACC GAT GTC GGC	240
Met Ile Arg Asp Asn Lys Asp Leu Val Arg Tyr Ser Thr Asp Val Gly	
65 70 75 80	
TTG AGC GAC AGC GGC CGC CAT CAA AAA GGC TTT GCC GTT CGC GGC GTG	288
Leu Ser Asp Ser Gly Arg His Gln Lys Gly Phe Ala Val Arg Gly Val	
85 90 95	

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1008

GTC AAC TAC AAA GGT TCG TTC CCG ATA GAG GAT TCT TCC ACC TTG ACA	1056
Val Asn Tyr Lys Gly Ser Phe Pro Ile Glu Asp Ser Ser Thr Leu Thr	
340 345 350	
CGT AAC TAC AAT CAA AAG GAC TTG GAT GAA ATC TAC AAC CGC AGT ATG	1104
Arg Asn Tyr Asn Gln Lys Asp Leu Asp Glu Ile Tyr Asn Arg Ser Met	
355 360 365	
GAT ACC CGC TTC AAA CGT ATT ACG CTG CGT TTG GAC AGC CAT CCG TTG	1152
Asp Thr Arg Phe Lys Arg Ile Thr Leu Arg Leu Asp Ser His Pro Leu	
370 375 380	
CAA CTC GGG GGG GGG CGA CAC CGC CTG TCG TTT AAA ACT TTC GCC AGC	1200
Gln Leu Gly Gly Gly Arg His Arg Leu Ser Phe Lys Thr Phe Ala Ser	
385 390 395 400	
CGC CGT GAT TTT GAA AAC CTA AAC CGC GAC TAT TAC TAC TTC AGC GGC	1248
Arg Arg Asp Phe Glu Asn Leu Asn Arg Asp Tyr Tyr Tyr Phe Ser Gly	
405 410 415	
CGT GTT GTT CGA ACC ACC AGC AGT ATC CAG CAT CCG GTG AAA ACC ACC	1296
Arg Val Val Arg Thr Thr Ser Ser Ile Gln His Pro Val Lys Thr Thr	
420 425 430	
AAC TAC GGT TTC TCA CTG TCT GAC CAA ATT CAA TGG AAC GAC GTG TTC	1344
Asn Tyr Gly Phe Ser Leu Ser Asp Gln Ile Gln Trp Asn Asp Val Phe	
435 440 445	
AGT AGC CGC GCA GGT ATC CGT TAC GAT CAT ACC AAA ATG ACG CCT CAG	1392
Ser Ser Arg Ala Gly Ile Arg Tyr Asp His Thr Lys Met Thr Pro Gln	
450 455 460	
GAA TTG AAT GCC GAG TGT CAT GCT TGT GAC AAA ACA CCG CCT GCA GCC	1440
Glu Leu Asn Ala Glu Cys His Ala Cys Asp Lys Thr Pro Pro Ala Ala	
465 470 475 480	
AAC ACT TAT AAA GGC TGG AGC GGT TTT GTC GGC TTG GCG GCG CAA CTG	1488
Asn Thr Tyr Lys Gly Trp Ser Gly Phe Val Gly Leu Ala Ala Gln Leu	
485 490 495	
AAT CAG GCT TGG CGT GTC GGT TAC GAC ATT ACT TCC GGC TAC CGT GTC	1536
Asn Gln Ala Trp Arg Val Gly Tyr Asp Ile Thr Ser Gly Tyr Arg Val	
500 505 510	
CCC AAT GCG TCC GAA GTG TAT TTC ACT TAC AAC CAC GGT TCG GGT AAT	1584
Pro Asn Ala Ser Glu Val Tyr Phe Thr Tyr Asn His Gly Ser Gly Asn	
515 520 525	
TGG CTG CCC AAT CCC AAC CTG AAA GCC GAG CGC ACG ACC ACC CAC ACC	1632
Trp Leu Pro Asn Pro Asn Leu Lys Ala Glu Arg Thr Thr Thr His Thr	
530 535 540	
CTC TCT CTG CAA GGC CGC AGC GAA AAA GGT ACT TTG GAT GCC AAC CTG	1680
Leu Ser Leu Gln Gly Arg Ser Glu Lys Gly Thr Leu Asp Ala Asn Leu	
545 550 555 560	
TAT CAA AGC AAT TAC CGA AAT TTC CTG TCT GAA GAG CAG AAG CTG ACC	1728
Tyr Gln Ser Asn Tyr Arg Asn Phe Leu Ser Glu Glu Gln Lys Leu Thr	
565 570 575	

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ACC	AGC	GGC	GAT	GTC	AGC	TGT	ACT	CAG	ATG	AAT	TAC	TAC	TAC	GGT	ATG	1776
Thr	Ser	Gly	Asp	Val	Ser	Cys	Thr	Gln	Met	Asn	Tyr	Tyr	Tyr	Gly	Met	
			580					585						590		
TGT	AGC	AAT	CCT	TAT	TCC	GAA	AAA	CTG	GAA	TGG	CAG	ATG	CAA	AAT	ATC	1824
Cys	Ser	Asn	Pro	Tyr	Ser	Glu	Lys	Leu	Glu	Trp	Gln	Met	Gln	Asn	Ile	
		595					600					605				
GAC	AAG	GCC	AGA	ATC	CGC	GGT	ATC	GAG	CTG	ACG	GGC	CGT	CTG	AAT	GTG	1872
Asp	Lys	Ala	Arg	Ile	Arg	Gly	Ile	Glu	Leu	Thr	Gly	Arg	Leu	Asn	Val	
	610					615					620					
GAC	AAA	GTA	GCG	TCT	TTT	GTT	CCT	GAG	GGC	TGG	AAA	CTG	TTC	GGC	TCG	1920
Asp	Lys	Val	Ala	Ser	Phe	Val	Pro	Glu	Gly	Trp	Lys	Leu	Phe	Gly	Ser	
	625				630					635					640	
CTG	GGT	TAT	GCG	AAA	AGC	AAA	CTG	TCG	GGC	GAC	AAC	AGC	CTG	CTG	TCC	1968
Leu	Gly	Tyr	Ala	Lys	Ser	Lys	Leu	Ser	Gly	Asp	Asn	Ser	Leu	Leu	Ser	
				645					650					655		
ACC	CAG	CCG	TTG	AAA	GTG	ATT	GCC	GGT	ATC	GAC	TAT	GAA	AGT	CCG	AGC	2016
Thr	Gln	Pro	Leu	Lys	Val	Ile	Ala	Gly	Ile	Asp	Tyr	Glu	Ser	Pro	Ser	
			660					665					670			
GAA	AAA	TGG	GGC	GTG	TTC	TCC	CGC	CTG	ACC	TAT	CTG	GGC	GCG	AAA	AAG	2064
Glu	Lys	Trp	Gly	Val	Phe	Ser	Arg	Leu	Thr	Tyr	Leu	Gly	Ala	Lys	Lys	
		675					680					685				
GTC	AAA	GAC	GCG	CAA	TAC	ACC	GTT	TAT	GAA	AAC	AAG	GGC	TGG	GGT	ACG	2112
Val	Lys	Asp	Ala	Gln	Tyr	Thr	Val	Tyr	Glu	Asn	Lys	Gly	Trp	Gly	Thr	
	690					695					700					
CCT	TTG	CAG	AAA	AAG	GTA	AAA	GAT	TAC	CCG	TGG	CTG	AAC	AAG	TCG	GCT	2160
Pro	Leu	Gln	Lys	Lys	Val	Lys	Asp	Tyr	Pro	Trp	Leu	Asn	Lys	Ser	Ala	
	705				710					715					720	
TAT	GTG	TTC	GAT	ATG	TAC	GGC	TTC	TAC	AAA	CCG	GTG	AAA	AAC	CTG	ACT	2208
Tyr	Val	Phe	Asp	Met	Tyr	Gly	Phe	Tyr	Lys	Pro	Val	Lys	Asn	Leu	Thr	
				725				730						735		
TTG	CGT	GCA	GGC	GTA	TAT	AAT	GTG	TTC	AAC	CGC	AAA	TAC	ACC	ACT	TGG	2256
Leu	Arg	Ala	Gly	Val	Tyr	Asn	Val	Phe	Asn	Arg	Lys	Tyr	Thr	Thr	Trp	
			740				745					750				
GAT	TCC	CTG	CGC	GGC	CTG	TAT	AGC	TAC	AGC	ACC	ACC	AAC	TCG	GTC	GAC	2304
Asp	Ser	Leu	Arg	Gly	Leu	Tyr	Ser	Tyr	Ser	Thr	Thr	Asn	Ser	Val	Asp	
		755					760					765				
CGC	GAT	GGC	AAA	GGC	TTA	GAC	CGC	TAC	CGC	GCC	CCA	AGC	CGT	AAT	TAC	2352
Arg	Asp	Gly	Lys	Gly	Leu	Asp	Arg	Tyr	Arg	Ala	Pro	Ser	Arg	Asn	Tyr	
	770					775					780					
GCC	GTA	TCG	CTG	GAA	TGG	AAG	TTT	TAA								2379
Ala	Val	Ser	Leu	Glu	Trp	Lys	Phe									
	785					790										

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 792 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Lys Pro Leu Gln Met Leu Pro Ile Ala Ala Leu Val Gly Ser Ile
1 5 10 15
Phe Gly Asn Pro Val Phe Ala Ala Asp Glu Ala Ala Thr Glu Thr Thr
20 25 30
Pro Val Lys Ala Glu Val Lys Ala Val Arg Val Lys Gly Gln Arg Asn
35 40 45
Ala Pro Ala Ala Val Glu Arg Val Asn Leu Asn Arg Ile Lys Gln Glu
50 55 60
Met Ile Arg Asp Asn Lys Asp Leu Val Arg Tyr Ser Thr Asp Val Gly
65 70 75 80
Leu Ser Asp Ser Gly Arg His Gln Lys Gly Phe Ala Val Arg Gly Val
85 90 95
Glu Gly Asn Arg Val Gly Val Ser Ile Asp Gly Val Asn Leu Pro Asp
100 105 110
Ser Glu Glu Asn Ser Leu Tyr Ala Arg Tyr Gly Asn Phe Asn Ser Ser
115 120 125
Arg Leu Ser Ile Asp Pro Glu Leu Val Arg Asn Ile Asp Ile Val Lys
130 135 140
Gly Ala Asp Ser Phe Asn Thr Gly Ser Gly Ala Leu Gly Gly Gly Val
145 150 155 160
Asn Tyr Gln Thr Leu Gln Gly Arg Asp Leu Leu Leu Pro Glu Arg Gln
165 170 175
Phe Gly Val Met Met Lys Asn Gly Tyr Ser Thr Arg Asn Arg Glu Trp
180 185 190
Thr Asn Thr Leu Gly Phe Gly Val Ser Asn Asp Arg Val Asp Ala Ala
195 200 205
Leu Leu Tyr Ser Gln Arg Arg Gly His Glu Thr Glu Ser Ala Gly Lys
210 215 220
Arg Gly Tyr Pro Val Glu Gly Ala Gly Ser Gly Ala Asn Ile Arg Gly
225 230 235 240
Ser Ala Arg Gly Ile Pro Asp Pro Ser Gln His Lys Tyr His Ser Phe
245 250 255
Leu Gly Lys Ile Ala Tyr Gln Ile Asn Asp Asn His Arg Ile Gly Ala

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260					265					270					
Ser	Leu	Asn	Gly	Gln	Gln	Gly	His	Asn	Tyr	Thr	Val	Glu	Glu	Ser	Tyr
		275					280					285			
Asn	Leu	Leu	Ala	Ser	Tyr	Trp	Arg	Glu	Ala	Asp	Asp	Val	Asn	Arg	Arg
	290					295					300				
Arg	Asn	Thr	Asn	Leu	Phe	Tyr	Glu	Trp	Thr	Pro	Glu	Ser	Asp	Arg	Leu
305					310					315					320
Ser	Met	Val	Lys	Ala	Asp	Val	Asp	Tyr	Gln	Lys	Thr	Lys	Val	Ser	Ala
				325					330					335	
Val	Asn	Tyr	Lys	Gly	Ser	Phe	Pro	Ile	Glu	Asp	Ser	Ser	Thr	Leu	Thr
			340					345					350		
Arg	Asn	Tyr	Asn	Gln	Lys	Asp	Leu	Asp	Glu	Ile	Tyr	Asn	Arg	Ser	Met
		355					360					365			
Asp	Thr	Arg	Phe	Lys	Arg	Ile	Thr	Leu	Arg	Leu	Asp	Ser	His	Pro	Leu
	370					375					380				
Gln	Leu	Gly	Gly	Gly	Arg	His	Arg	Leu	Ser	Phe	Lys	Thr	Phe	Ala	Ser
385					390					395					400
Arg	Arg	Asp	Phe	Glu	Asn	Leu	Asn	Arg	Asp	Tyr	Tyr	Tyr	Phe	Ser	Gly
				405					410					415	
Arg	Val	Val	Arg	Thr	Thr	Ser	Ser	Ile	Gln	His	Pro	Val	Lys	Thr	Thr
			420					425					430		
Asn	Tyr	Gly	Phe	Ser	Leu	Ser	Asp	Gln	Ile	Gln	Trp	Asn	Asp	Val	Phe
		435					440					445			
Ser	Ser	Arg	Ala	Gly	Ile	Arg	Tyr	Asp	His	Thr	Lys	Met	Thr	Pro	Gln
	450					455					460				
Glu	Leu	Asn	Ala	Glu	Cys	His	Ala	Cys	Asp	Lys	Thr	Pro	Pro	Ala	Ala
465					470					475					480
Asn	Thr	Tyr	Lys	Gly	Trp	Ser	Gly	Phe	Val	Gly	Leu	Ala	Ala	Gln	Leu
				485					490					495	
Asn	Gln	Ala	Trp	Arg	Val	Gly	Tyr	Asp	Ile	Thr	Ser	Gly	Tyr	Arg	Val
			500					505					510		
Pro	Asn	Ala	Ser	Glu	Val	Tyr	Phe	Thr	Tyr	Asn	His	Gly	Ser	Gly	Asn
		515					520					525			
Trp	Leu	Pro	Asn	Pro	Asn	Leu	Lys	Ala	Glu	Arg	Ser	Thr	Thr	His	Thr
	530					535					540				
Leu	Ser	Leu	Gln	Gly	Arg	Ser	Glu	Lys	Gly	Thr	Leu	Asp	Ala	Asn	Leu
545					550					555					560
Tyr	Gln	Ser	Asn	Tyr	Arg	Asn	Phe	Leu	Ser	Glu	Glu	Gln	Lys	Leu	Thr

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565										570					575				
Thr	Ser	Gly	Asp	Val	Ser	Cys	Thr	Gln	Met	Asn	Tyr	Tyr	Tyr	Gly	Met				
			580					585						590					
Cys	Ser	Asn	Pro	Tyr	Ser	Glu	Lys	Leu	Glu	Trp	Gln	Met	Gln	Asn	Ile				
		595					600					605							
Asp	Lys	Ala	Arg	Ile	Arg	Gly	Ile	Glu	Leu	Thr	Gly	Arg	Leu	Asn	Val				
	610					615					620								
Asp	Lys	Val	Ala	Ser	Phe	Val	Pro	Glu	Gly	Trp	Lys	Leu	Phe	Gly	Ser				
625					630					635					640				
Leu	Gly	Tyr	Ala	Lys	Ser	Lys	Leu	Ser	Gly	Asp	Asn	Ser	Leu	Leu	Ser				
				645					650					655					
Thr	Gln	Pro	Leu	Lys	Val	Ile	Ala	Gly	Ile	Asp	Tyr	Glu	Ser	Pro	Ser				
			660					665					670						
Glu	Lys	Trp	Gly	Val	Phe	Ser	Arg	Leu	Thr	Tyr	Leu	Gly	Ala	Lys	Lys				
		675					680					685							
Val	Lys	Asp	Ala	Gln	Tyr	Thr	Val	Tyr	Glu	Asn	Lys	Gly	Trp	Gly	Thr				
	690					695					700								
Pro	Leu	Gln	Lys	Lys	Val	Lys	Asp	Tyr	Pro	Trp	Leu	Asn	Lys	Ser	Ala				
705					710					715					720				
Tyr	Val	Phe	Asp	Met	Tyr	Gly	Phe	Tyr	Lys	Pro	Val	Lys	Asn	Leu	Thr				
				725					730					735					
Leu	Arg	Ala	Cly	Val	Tyr	Asn	Val	Phe	Asn	Arg	Lys	Tyr	Thr	Thr	Trp				
			740					745					750						
Asp	Ser	Leu	Arg	Gly	Leu	Tyr	Ser	Tyr	Ser	Thr	Thr	Asn	Ser	Val	Asp				
		755					760					765							
Arg	Asp	Gly	Lys	Gly	Leu	Asp	Arg	Tyr	Arg	Ala	Pro	Ser	Arg	Asn	Tyr				
	770					775					780								
Ala	Val	Ser	Leu	Glu	Trp	Lys	Phe												
785					790														

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2378 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 1..2370

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

ATG AAA CCA TTA CAC ATG CTT CCT ATT GCC GCG CTG GTC GGC AGT ATT	48
Met Lys Pro Leu His Met Leu Pro Ile Ala Ala Leu Val Gly Ser Ile	
1 5 10 15	
TTC GGC AAT CCG GTC TTG GCA GCG GAT GAA GCT GCA ACC GAA ACC ACA	96
Phe Gly Asn Pro Val Leu Ala Ala Asp Glu Ala Ala Thr Glu Thr Thr	
20 25 30	
CCC GTT AAA GCA GAG ATA AAA GAA GTG CGC GTT AAA GAC CAG CTT AAT	144
Pro Val Lys Ala Glu Ile Lys Glu Val Arg Val Lys Asp Gln Leu Asn	
35 40 45	
GCG CCT GCA ACC GTG GAA CGT GTC AAC CTC GGC CGC ATT CAA CAG GAA	192
Ala Pro Ala Thr Val Glu Arg Val Asn Leu Gly Arg Ile Gln Gln Glu	
50 55 60	
ATG ATA CGC GAC AAC AAA GAC TTG GTG CGT TAC TCC ACC GAC GTC GGC	240
Met Ile Arg Asp Asn Lys Asp Leu Val Arg Tyr Ser Thr Asp Val Gly	
65 70 75 80	
TTG AGC GAT AGC GGC CGC CAT CAA AAA GGC TTT GCT GTG CGC GGC GTG	288
Leu Ser Asp Ser Gly Arg His Gln Lys Gly Phe Ala Val Arg Gly Val	
85 90 95	
GAA GGC AAC CGT GTC GGT GTC AGC ATT GAC GGC GTG AGC CTG CCT GAT	336
Glu Gly Asn Arg Val Gly Val Ser Ile Asp Gly Val Ser Leu Pro Asp	
100 105 110	
TCG GAA GAA AAC TCA CTG TAT GCA CGT TAT GGC AAC TTC AAC AGC TCG	384
Ser Glu Glu Asn Ser Leu Tyr Ala Arg Tyr Gly Asn Phe Asn Ser Ser	
115 120 125	
CGC CTG TCT ATC GAC CCC GAA CTC GTG CGC AAC ATC GAA ATC GCG AAG	432
Arg Leu Ser Ile Asp Pro Glu Leu Val Arg Asn Ile Glu Ile Ala Lys	
130 135 140	
GGC GCT GAC TCT TTC AAT ACC GGT AGC GGC GCA TTG GGT GGC GGC GTG	480
Gly Ala Asp Ser Phe Asn Thr Gly Ser Gly Ala Leu Gly Gly Gly Val	
145 150 155 160	
AAT TAC CAA ACC CTG CAA GGA CAT GAT TTG CTG TTG GAC GAC AGG CAA	528
Asn Tyr Gln Thr Leu Gln Gly His Asp Leu Leu Leu Asp Asp Arg Gln	
165 170 175	
TTC GGC GTG ATG ATG AAA AAC GGT TAC AGC ACG CGC AAC CGC GAA TGG	576
Phe Gly Val Met Met Lys Asn Gly Tyr Ser Thr Arg Asn Arg Glu Trp	
180 185 190	
ACA AAT ACA CTC GGT TTC GGT GTG AGC AAC GAC CGC GTG GAT GCC GCT	624
Thr Asn Thr Leu Gly Phe Gly Val Ser Asn Asp Arg Val Asp Ala Ala	
195 200 205	
TTG CTG TAT TCG CAA CGT CGC GGT CAT GAG ACC GAA AGC GCG GGC GAG	672
Leu Leu Tyr Ser Gln Arg Arg Gly His Glu Thr Glu Ser Ala Gly Glu	

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210	215	220	
CGT GGC TAT CCG GTA GAG GGT GCT GGC AGC GGA GCA ATT ATC CGT GGT Arg Gly Tyr Pro Val Glu Gly Ala Gly Ser Gly Ala Ile Ile Arg Gly 225 230 235 240			720
TCG TCA CGC GGT ATC CCT GAT CCG TCC AAA CAC AAA TAC CAC AAC TTC Ser Ser Arg Gly Ile Pro Asp Pro Ser Lys His Lys Tyr His Asn Phe 245 250 255			768
TTG GGT AAG ATT GCT TAT CAA ATC AAC GAC AAG CAC CGC ATC GGC CCA Leu Gly Lys Ile Ala Tyr Gln Ile Asn Asp Lys His Arg Ile Gly Pro 260 265 270			816
TCG TTT AAC GGC CAG CAG GGG CAT AAT TAC ACG ATT GAA GAG TCT TAT Ser Phe Asn Gly Gln Gln Gly His Asn Tyr Thr Ile Glu Glu Ser Tyr 275 280 285			864
AAC CTG ACC GCT TCT TCC TGG CGC GAA GCC GAT GAC GTA AAC AGA CGG Asn Leu Thr Ala Ser Ser Trp Arg Glu Ala Asp Asp Val Asn Arg Arg 290 295 300			912
CGC AAT GCC AAC CTC TTT TAC GAA TGG ACG CCT GAT TCA AAT TGG CTG Arg Asn Ala Asn Leu Phe Tyr Glu Trp Thr Pro Asp Ser Asn Trp Leu 305 310 315 320			960
TCG TCT TTG AAG GCG GAT TTC GAT TAT CAG ACA ACC AAA GTG GCG GCG Ser Ser Leu Lys Ala Asp Phe Asp Tyr Gln Thr Thr Lys Val Ala Ala 325 330 335			1008
GTT AAC AAC AAA GGC TCG TTC CCG ACG GAT TAT TCC ACC TTG ACG CGC Val Asn Asn Lys Gly Ser Phe Pro Thr Asp Tyr Ser Thr Leu Thr Arg 340 345 350			1056
AAC TAT AAT CAG AAG GAT TTG GAG AAT ATA TAC AAC CGC AGC ATG GAC Asn Tyr Asn Gln Lys Asp Leu Glu Asn Ile Tyr Asn Arg Ser Met Asp 355 360 365			1104
ACC CGA TTC AAA CGT TTT ACT TTG CGT ATG GAC AGC CAA CCG TTG CAA Thr Arg Phe Lys Arg Phe Thr Leu Arg Met Asp Ser Gln Pro Leu Gln 370 375 380			1152
CTG GGC GGC CAA CAT CGC TTG TCG CTT AAA ACT TTC GCC AGT CGG CGT Leu Gly Gly Gln His Arg Leu Ser Leu Lys Thr Phe Ala Ser Arg Arg 385 390 395 400			1200
GAG TTT GAA AAC TTA AAC CGC GAC GAT TAT TAC TTC AGC GAA AGA GTA Glu Phe Glu Asn Leu Asn Arg Asp Asp Tyr Tyr Phe Ser Glu Arg Val 405 410 415			1248
TCC CGT ACT ACC AGC TCG ATT CAA CAC CCC GTG AAA ACC ACT AAT TAT Ser Arg Thr Thr Ser Ser Ile Gln His Pro Val Lys Thr Thr Asn Tyr 420 425 430			1296
GGT TTC TCA CTG TCT GAT CAA ATC CAA TGG AAC GAC GTG TTC AGC AGC Gly Phe Ser Leu Ser Asp Gln Ile Gln Trp Asn Asp Val Phe Ser Ser 435 440 445			1344
CGT GCA GAT ATC CGT TAC GAT CAT ACC AAA ATG ACG CCT CAG GAA TTG			1392

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Arg	Ala	Asp	Ile	Arg	Tyr	Asp	His	Thr	Lys	Met	Thr	Pro	Gln	Glu	Leu	
450						455					460					
AAT	GCC	GAG	TGT	CAT	GCT	TGT	GAC	AAA	ACA	CCG	CCT	GCA	GCC	AAT	ACT	1440
Asn	Ala	Glu	Cys	His	Ala	Cys	Asp	Lys	Thr	Pro	Pro	Ala	Ala	Asn	Thr	
465					470					475					480	
TAT	AAA	GGC	TGG	AGC	GGA	TTT	GTC	GGT	TTG	GCG	GCG	CAA	CTG	AAT	CAG	1488
Tyr	Lys	Gly	Trp	Ser	Gly	Phe	Val	Gly	Leu	Ala	Ala	Gln	Leu	Asn	Gln	
				485					490					495		
GCT	TGG	CAT	GTC	GGT	TAC	GAC	ATT	ACT	TCC	GGC	TAC	CGT	GTC	CCC	AAT	1536
Ala	Trp	His	Val	Gly	Tyr	Asp	Ile	Thr	Ser	Gly	Tyr	Arg	Val	Pro	Asn	
			500					505					510			
GCG	TCC	GAA	GTG	TAT	TTC	ACT	TAC	AAC	CAC	GGT	TCG	GGT	AAT	TGG	CTG	1584
Ala	Ser	Glu	Val	Tyr	Phe	Thr	Tyr	Asn	His	Gly	Ser	Gly	Asn	Trp	Leu	
		515					520					525				
CCC	AAT	CCC	AAC	CTG	AAA	GCC	GAG	CGC	AGC	ACC	ACC	CAC	ACC	CTG	TCT	1632
Pro	Asn	Pro	Asn	Leu	Lys	Ala	Glu	Arg	Ser	Thr	Thr	His	Thr	Leu	Ser	
	530				535						540					
CTG	CAA	GGC	CGC	AGC	GAA	AAA	GGT	ACT	TTG	GAT	GCC	AAC	CTG	TAT	CAA	1680
Leu	Gln	Gly	Arg	Ser	Glu	Lys	Gly	Thr	Leu	Asp	Ala	Asn	Leu	Tyr	Gln	
545					550				555						560	
AGC	AAT	TAC	CGA	AAC	TTC	TTG	TCT	GAA	GAG	CAG	AAG	CTG	ACC	ACC	AGC	1728
Ser	Asn	Tyr	Arg	Asn	Phe	Leu	Ser	Glu	Glu	Gln	Lys	Leu	Thr	Thr	Ser	
				565				570						575		
GGC	GAT	GTC	GGC	TGT	ACT	CAG	ATG	AAT	TAC	TAC	TAC	GGT	ATG	TGT	AGC	1776
Gly	Asp	Val	Gly	Cys	Thr	Gln	Met	Asn	Tyr	Tyr	Tyr	Gly	Met	Cys	Ser	
			580					585					590			
AAT	CCT	TAT	TCC	GAA	AAA	CCG	GAA	TGG	CAG	ATG	CAA	AAT	ATC	GAT	AAG	1824
Asn	Pro	Tyr	Ser	Glu	Lys	Pro	Glu	Trp	Gln	Met	Gln	Asn	Ile	Asp	Lys	
		595				600					605					
GCC	CGA	ATC	CGT	GGT	CTT	GAG	CTG	ACG	GGC	CGT	CTG	AAT	GTG	ACA	AAA	1872
Ala	Arg	Ile	Arg	Gly	Leu	Glu	Leu	Thr	Gly	Arg	Leu	Asn	Val	Thr	Lys	
	610				615						620					
GTA	GCG	TCT	TTT	GTT	CCT	GAG	GGC	TGG	AAA	TTG	TTC	GGC	TCG	CTG	GGT	1920
Val	Ala	Ser	Phe	Val	Pro	Glu	Gly	Trp	Lys	Leu	Phe	Gly	Ser	Leu	Gly	
625					630				635						640	
TAT	GCG	AAA	AGC	AAA	CTG	TCG	GGC	GAC	AAC	AGC	CTG	CTG	TCC	ACA	CAG	1968
Tyr	Ala	Lys	Ser	Lys	Leu	Ser	Gly	Asp	Asn	Ser	Leu	Leu	Ser	Thr	Gln	
				645				650						655		
CCG	CCG	AAA	GTG	ATT	GCC	GGT	ATC	GAC	TAT	GAA	AGT	CCG	AGC	GAA	AAA	2016
Pro	Pro	Lys	Val	Ile	Ala	Gly	Ile	Asp	Tyr	Glu	Ser	Pro	Ser	Glu	Lys	
			660					665					670			
TGG	GGT	GTG	TTC	TCC	CGC	CTG	ACT	TAT	CTG	GGT	GCG	AAA	AAG	GTC	AAA	2064
Trp	Gly	Val	Phe	Ser	Arg	Leu	Thr	Tyr	Leu	Gly	Ala	Lys	Lys	Val	Lys	
		675					680					685				

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GAC GCG CAA TAC ACC GTT TAT GAA AAC AAG GGC CGG GGT ACG CCT TTG	2112
Asp Ala Gln Tyr Thr Val Tyr Glu Asn Lys Gly Arg Gly Thr Pro Leu	
690 695 700	
CAG AAA AAG GTA AAA GAT TAC CCG TGG CTG AAC AAG TCG GCT TAT GTG	2160
Gln Lys Lys Val Lys Asp Tyr Pro Trp Leu Asn Lys Ser Ala Tyr Val	
705 710 715 720	
TTT GAT ATG TAC GGC TTC TAC AAA CTG GCT AAA AAC CTG ACT TTG CGT	2208
Phe Asp Met Tyr Gly Phe Tyr Lys Leu Ala Lys Asn Leu Thr Leu Arg	
725 730 735	
GCA GGC GTA TAT AAT GTG TTC AAC CGC AAA TAC ACC ACT TGG GAT TCC	2256
Ala Gly Val Tyr Asn Val Phe Asn Arg Lys Tyr Thr Thr Trp Asp Ser	
740 745 750	
CTG CGC GGT TTG TAT AGC TAC ACC ACC ACC AAC GCG GTC GAC CGA GAT	2304
Leu Arg Gly Leu Tyr Ser Tyr Thr Thr Thr Asn Ala Val Asp Arg Asp	
755 760 765	
GGC AAA GGC TTA GAC CGC TAC CGC GCC TCA GGC CGT AAT TAC GCC GTA	2352
Gly Lys Gly Leu Asp Arg Tyr Arg Ala Ser Gly Arg Asn Tyr Ala Val	
770 775 780	
TCG CTG GAT TGG AAG TTT TGAATTCC	2378
Ser Leu Asp Trp Lys Phe	
885 790	

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 790 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met Lys Pro Leu His Met Leu Pro Ile Ala Ala Leu Val Gly Ser Ile	
1 5 10 15	
Phe Gly Asn Pro Val Leu Ala Ala Asp Glu Ala Ala Thr Glu Thr Thr	
20 25 30	
Pro Val Lys Ala Glu Ile Lys Glu Val Arg Val Lys Asp Gln Leu Asn	
35 40 45	
Ala Pro Ala Thr Val Glu Arg Val Asn Leu Gly Arg Ile Gln Gln Glu	
50 55 60	
Met Ile Arg Asp Asn Lys Asp Leu Val Arg Tyr Ser Thr Asp Val Gly	
65 70 75 80	
Leu Ser Asp Ser Gly Arg His Gln Lys Gly Phe Ala Val Arg Gly Val	
85 90 95	
Glu Gly Asn Arg Val Gly Val Ser Ile Asp Gly Val Ser Leu Pro Asp	

006760"091900

100					105					110					
Ser	Glu	Glu	Asn	Ser	Leu	Tyr	Ala	Arg	Tyr	Gly	Asn	Phe	Asn	Ser	Ser
		115					120					125			
Arg	Leu	Ser	Ile	Asp	Pro	Glu	Leu	Val	Arg	Asn	Ile	Glu	Ile	Ala	Lys
	130					135					140				
Gly	Ala	Asp	Ser	Phe	Asn	Thr	Gly	Ser	Gly	Ala	Leu	Gly	Gly	Gly	Val
145					150					155					160
Asn	Tyr	Gln	Thr	Leu	Gln	Gly	His	Asp	Leu	Leu	Leu	Asp	Asp	Arg	Gln
				165					170					175	
Phe	Gly	Val	Met	Met	Lys	Asn	Gly	Tyr	Ser	Thr	Arg	Asn	Arg	Glu	Trp
			180					185					190		
Thr	Asn	Thr	Leu	Gly	Phe	Gly	Val	Ser	Asn	Asp	Arg	Val	Asp	Ala	Ala
			195				200					205			
Leu	Leu	Tyr	Ser	Gln	Arg	Arg	Gly	His	Glu	Thr	Glu	Ser	Ala	Gly	Glu
	210					215					220				
Arg	Gly	Tyr	Pro	Val	Glu	Gly	Ala	Gly	Ser	Gly	Ala	Ile	Ile	Arg	Gly
225					230					235					240
Ser	Ser	Arg	Gly	Ile	Pro	Asp	Pro	Ser	Lys	His	Lys	Tyr	His	Asn	Phe
				245					250					255	
Leu	Gly	Lys	Ile	Ala	Tyr	Gln	Ile	Asn	Asp	Lys	His	Arg	Ile	Gly	Pro
			260					265					270		
Ser	Phe	Asn	Gly	Gln	Gln	Gly	His	Asn	Tyr	Thr	Ile	Glu	Glu	Ser	Tyr
		275					280					285			
Asn	Leu	Thr	Ala	Ser	Ser	Trp	Arg	Glu	Ala	Asp	Asp	Val	Asn	Arg	Arg
	290					295					300				
Arg	Asn	Ala	Asn	Leu	Phe	Tyr	Glu	Trp	Thr	Pro	Asp	Ser	Asn	Trp	Leu
305					310					315					320
Ser	Ser	Leu	Lys	Ala	Asp	Phe	Asp	Tyr	Gln	Thr	Thr	Lys	Val	Ala	Ala
				325					330					335	
Val	Asn	Asn	Lys	Gly	Ser	Phe	Pro	Thr	Asp	Tyr	Ser	Thr	Leu	Thr	Arg
			340				345						350		
Asn	Tyr	Asn	Gln	Lys	Asp	Leu	Glu	Asn	Ile	Tyr	Asn	Arg	Ser	Met	Asp
		355				360						365			
Thr	Arg	Phe	Lys	Arg	Phe	Thr	Leu	Arg	Met	Asp	Ser	Gln	Pro	Leu	Gln
	370					375					380				
Leu	Gly	Gly	Gln	His	Arg	Leu	Ser	Leu	Lys	Thr	Phe	Ala	Ser	Arg	Arg
385					390					395					400
Glu	Phe	Glu	Asn	Leu	Asn	Arg	Asp	Asp	Tyr	Tyr	Phe	Ser	Glu	Arg	Val
			405						410				415		

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Ser	Arg	Thr	Thr	Ser	Ser	Ile	Gln	His	Pro	Val	Lys	Thr	Thr	Asn	Tyr
			420					425						430	
Gly	Phe	Ser	Leu	Ser	Asp	Gln	Ile	Gln	Trp	Asn	Asp	Val	Phe	Ser	Ser
		435					440					445			
Arg	Ala	Asp	Ile	Arg	Tyr	Asp	His	Thr	Lys	Met	Thr	Pro	Gln	Glu	Leu
	450					455					460				
Asn	Ala	Glu	Cys	His	Ala	Cys	Asp	Lys	Thr	Pro	Pro	Ala	Ala	Asn	Thr
465					470					475					480
Tyr	Lys	Gly	Trp	Ser	Gly	Phe	Val	Gly	Leu	Ala	Ala	Gln	Leu	Asn	Gln
				485					490						495
Ala	Trp	His	Val	Gly	Tyr	Asp	Ile	Thr	Ser	Gly	Tyr	Arg	Val	Pro	Asn
			500					505					510		
Ala	Ser	Glu	Val	Tyr	Phe	Thr	Tyr	Asn	His	Gly	Ser	Gly	Asn	Trp	Leu
		515					520					525			
Pro	Asn	Pro	Asn	Leu	Lys	Ala	Glu	Arg	Ser	Thr	Thr	His	Thr	Leu	Ser
	530					535					540				
Leu	Gln	Gly	Arg	Ser	Glu	Lys	Gly	Thr	Leu	Asp	Ala	Asn	Leu	Tyr	Gln
545					550					555					560
Ser	Asn	Tyr	Arg	Asn	Phe	Leu	Ser	Glu	Glu	Gln	Lys	Leu	Thr	Thr	Ser
				565					570					575	
Gly	Asp	Val	Gly	Cys	Thr	Gln	Met	Asn	Tyr	Tyr	Tyr	Gly	Met	Cys	Ser
		580						585					590		
Asn	Pro	Tyr	Ser	Glu	Lys	Pro	Glu	Trp	Gln	Met	Gln	Asn	Ile	Asp	Lys
		595					600					605			
Ala	Arg	Ile	Arg	Gly	Leu	Glu	Leu	Thr	Gly	Arg	Leu	Asn	Val	Thr	Lys
		610				615					620				
Val	Ala	Ser	Phe	Val	Pro	Glu	Gly	Trp	Lys	Leu	Phe	Gly	Ser	Leu	Gly
625					630					635					640
Tyr	Ala	Lys	Ser	Lys	Leu	Ser	Gly	Asp	Asn	Ser	Leu	Leu	Ser	Thr	Gln
				645					650					655	
Pro	Pro	Lys	Val	Ile	Ala	Gly	Ile	Asp	Tyr	Glu	Ser	Pro	Ser	Glu	Lys
			660					665					670		
Trp	Gly	Val	Phe	Ser	Arg	Leu	Thr	Tyr	Leu	Gly	Ala	Lys	Lys	Val	Lys
		675					680					685			
Asp	Ala	Gln	Tyr	Thr	Val	Tyr	Glu	Asn	Lys	Gly	Arg	Gly	Thr	Pro	Leu
	690					695					700				
Gln	Lys	Lys	Val	Lys	Asp	Tyr	Pro	Trp	Leu	Asn	Lys	Ser	Ala	Tyr	Val
705					710					715					720
Phe	Asp	Met	Tyr	Gly	Phe	Tyr	Lys	Leu	Ala	Lys	Asn	Leu	Thr	Leu	Arg

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	725		730		735
Ala Gly Val Tyr Asn Val Phe Asn Arg Lys Tyr Thr Thr Trp Asp Ser	740		745		750
Leu Arg Gly Leu Tyr Ser Tyr Thr Thr Thr Asn Ala Val Asp Arg Asp	755		760		765
Gly Lys Gly Leu Asp Arg Tyr Arg Ala Ser Gly Arg Asn Tyr Ala Val	770		775		780
Ser Leu Asp Trp Lys Phe	785		790		

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 600 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met	Gln	Gln	Gln	His	Leu	Phe	Arg	Leu	Asn	Ile	Leu	Cys	Leu	Ser	Leu
1				5					10					15	
Met	Thr	Ala	Leu	Pro	Val	Tyr	Ala	Glu	Asn	Val	Gln	Ala	Glu	Gln	Ala
			20					25					30		
Gln	Glu	Lys	Gln	Leu	Asp	Thr	Ile	Val	Lys	Ala	Lys	Lys	Gln	Lys	Thr
		35					40					45			
Arg	Arg	Asp	Asn	Glu	Val	Thr	Gly	Leu	Gly	Lys	Leu	Val	Lys	Ser	Ser
	50					55					60				
Asp	Thr	Leu	Ser	Lys	Glu	Gln	Val	Leu	Asn	Ile	Arg	Asp	Leu	Thr	Arg
65					70				75						80
Tyr	Asp	Pro	Gly	Ile	Ala	Val	Val	Glu	Gln	Gly	Arg	Gly	Ala	Ser	Ser
				85					90					95	
Gly	Tyr	Ser	Ile	Arg	Gly	Met	Asp	Lys	Asn	Arg	Val	Ser	Leu	Thr	Val
			100					105					110		
Asp	Gly	Val	Ser	Gln	Ile	Gln	Ser	Tyr	Thr	Ala	Gln	Ala	Ala	Leu	Gly
	115						120					125			
Gly	Thr	Arg	Thr	Ala	Gly	Ser	Ser	Gly	Ala	Ile	Asn	Glu	Ile	Glu	Tyr
	130					135					140				
Glu	Asn	Val	Lys	Ala	Val	Glu	Ile	Ser	Lys	Gly	Ser	Asn	Ser	Ser	Glu
145					150					155					160
Tyr	Gly	Asn	Gly	Ala	Leu	Ala	Gly	Ser	Val	Ala	Phe	Gln	Thr	Lys	Thr
				165					170					175	

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Ala	Ala	Asp	Ile	Ile	Gly	Glu	Gly	Lys	Gln	Trp	Gly	Ile	Gln	Ser	Lys
			180					185					190		
Thr	Ala	Tyr	Ser	Gly	Lys	Asp	His	Ala	Leu	Thr	Gln	Ser	Leu	Ala	Leu
		195					200					205			
Ala	Gly	Arg	Ser	Gly	Gly	Ala	Glu	Ala	Leu	Leu	Ile	Tyr	Thr	Lys	Arg
	210					215					220				
Arg	Gly	Arg	Glu	Ile	His	Ala	His	Lys	Asp	Ala	Gly	Lys	Gly	Val	Gln
225					230					235					240
Ser	Phe	Asn	Arg	Leu	Pro	Ile	Cys	Arg	Phe	Gly	Asn	Asn	Thr	Tyr	Thr
				245					250						255
Asp	Cys	Thr	Pro	Arg	Asn	Ile	Gly	Gly	Asn	Gly	Tyr	Tyr	Ala	Ala	Val
			260					265					270		
Gln	Asp	Asn	Val	Arg	Leu	Gly	Arg	Trp	Ala	Asp	Val	Gly	Ala	Gly	Ile
	275						280					285			
Arg	Tyr	Asp	Tyr	Arg	Ser	Thr	His	Ser	Glu	Asp	Lys	Ser	Val	Ser	Thr
	290					295					300				
Gly	Thr	His	Arg	Asn	Leu	Ser	Trp	Asn	Ala	Gly	Val	Val	Leu	Lys	Pro
305					310					315					320
Phe	Thr	Trp	Met	Asp	Leu	Thr	Tyr	Arg	Ala	Ser	Thr	Gly	Phe	Arg	Leu
				325					330					335	
Pro	Ser	Phe	Ala	Glu	Met	Tyr	Gly	Trp	Arg	Ala	Gly	Glu	Ser	Leu	Lys
			340					345					350		
Thr	Leu	Asp	Leu	Lys	Pro	Glu	Lys	Ser	Phe	Asn	Arg	Glu	Ala	Gly	Ile
		355					360					365			
Val	Phe	Lys	Gly	Asp	Phe	Gly	Asn	Leu	Glu	Ala	Ser	Tyr	Phe	Asn	Asn
	370					375					380				
Ala	Tyr	Arg	Asp	Leu	Ile	Ala	Phe	Gly	Tyr	Glu	Thr	Arg	Thr	Gln	Asn
385					390					395					400
Gly	Gln	Thr	Ser	Ala	Ser	Gly	Asp	Pro	Gly	Tyr	Arg	Asn	Ala	Gln	Asn
				405					410					415	
Ala	Arg	Ile	Ala	Gly	Ile	Asn	Ile	Leu	Gly	Lys	Ile	Asp	Trp	His	Gly
			420					425					430		
Val	Trp	Gly	Gly	Leu	Pro	Asp	Gly	Leu	Tyr	Ser	Thr	Leu	Ala	Tyr	Asn
		435					440					445			
Arg	Ile	Lys	Val	Lys	Asp	Ala	Asp	Arg	Ala	Asp	Arg	Thr	Phe	Val	Thr
	450					455					460				
Ser	Tyr	Leu	Phe	Asp	Ala	Val	Gln	Pro	Ser	Arg	Tyr	Val	Leu	Gly	Leu
465					470					475					480
Gly	Tyr	Asp	His	Pro	Asp	Gly	Ile	Trp	Gly	Ile	Asn	Thr	Met	Phe	Thr
				485					490					495	

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Tyr Ser Lys Ala Lys Ser Val Asp Glu Leu Leu Gly Ser Gln Ala Leu
500 505 510

Leu Asn Gly Asn Ala Asn Ala Lys Lys Ala Ala Ser Arg Arg Thr Arg
515 520 525

Pro Trp Tyr Val Thr Asp Val Ser Gly Tyr Tyr Asn Ile Lys Lys His
530 535 540

Leu Thr Leu Arg Ala Gly Val Tyr Asn Leu Leu Asn Tyr Arg Tyr Val
545 550 555 560

Thr Trp Glu Asn Val Arg Gln Thr Ala Gly Gly Ala Val Asn Gln His
565 570 575

Lys Asn Val Gly Val Tyr Asn Arg Tyr Ala Ala Pro Gly Arg Asn Tyr
580 585 590

Thr Phe Ser Leu Glu Met Lys Phe
595 600

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 607 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Met Asn Lys Lys His Gly Phe Gln Leu Thr Leu Thr Ala Leu Ala Val
1 5 10 15

Ala Ala Ala Phe Pro Ser Tyr Ala Ala Asn Pro Glu Thr Ala Ala Pro
20 25 30

Asp Ala Ala Gln Thr Gln Ser Leu Lys Glu Val Thr Val Arg Ala Ala
35 40 45

Lys Val Gly Arg Arg Ser Lys Glu Ala Thr Gly Leu Gly Lys Ile Ala
50 55 60

Lys Thr Ser Glu Thr Leu Asn Lys Glu Gln Val Leu Gly Ile Arg Asp
65 70 75 80

Leu Thr Arg Tyr Asp Pro Gly Val Ala Val Val Glu Gln Gly Asn Gly
85 90 95

Ala Ser Gly Glu Tyr Ser Ile Arg Gly Val Asp Lys Asn Arg Val Ala
100 105 110

Val Ser Val Asp Gly Val Ala Gln Ile Gln Ala Phe Thr Val Gln Gly
115 120 125

Ser Leu Ser Gly Tyr Gly Gly Arg Gly Gly Ser Gly Ala Ile Asn Glu

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130	135	140
Ile Glu Tyr Glu Asn Ile Ser Thr Val Glu Ile Asp Lys Gly Ala Gly		
145	150	155 160
Ser Ser Asp His Gly Ser Gly Ala Leu Gly Gly Ala Val Ala Phe Arg		
	165	170 175
Thr Lys Glu Ala Ala Asp Leu Ile Ser Asp Gly Lys Ser Trp Gly Ile		
	180	185 190
Gln Ala Lys Thr Ala Tyr Gly Ser Lys Asn Arg Gln Phe Met Lys Ser		
	195	200 205
Leu Gly Ala Gly Phe Ser Lys Asp Gly Trp Glu Gly Leu Leu Ile Arg		
	210	215 220
Thr Glu Arg Gln Gly Arg Glu Thr His Pro His Gly Asp Ile Ala Asp		
	225	230 235 240
Gly Val Ala Tyr Gly Ile Asn Arg Leu Ser Val Cys Gly Tyr Ile Glu		
	245	250 255
Thr Leu Arg Ser Arg Lys Cys Val Pro Arg Lys Ile Asn Gly Ser Asn		
	260	265 270
Ile His Ile Ser Leu Asn Asp Arg Phe Ser Ile Gly Lys Tyr Phe Asp		
	275	280 285
Phe Ser Leu Gly Gly Arg Tyr Asp Arg Lys Asn Phe Thr Thr Ser Glu		
	290	295 300
Glu Leu Val Arg Ser Gly Arg Tyr Val Asp Arg Ser Trp Asn Ser Gly		
	305	310 315 320
Ile Val Phe Lys Pro Asn Arg His Phe Ser Leu Ser Tyr Arg Ala Ser		
	325	330 335
Ser Gly Phe Arg Thr Pro Ser Phe Gln Glu Leu Phe Gly Ile Asp Ile		
	340	345 350
Tyr His Asp Tyr Pro Lys Gly Trp Gln Arg Pro Ala Leu Lys Ser Glu		
	355	360 365
Lys Ala Ala Asn Arg Glu Ile Gly Leu Gln Trp Lys Gly Asp Phe Gly		
	370	375 380
Phe Leu Glu Ile Ser Ser Phe Arg Asn Arg Tyr Thr Asp Met Ile Ala		
	385	390 395 400
Val Ala Asp His Lys Thr Lys Leu Pro Asn Gln Ala Gly Gln Leu Thr		
	405	410 415
Glu Ile Asp Ile Arg Asp Tyr Tyr Asn Ala Gln Asn Met Ser Leu Gln		
	420	425 430
Gly Val Asn Ile Leu Gly Lys Ile Asp Trp Asn Gly Val tyr Gly Lys		
	435	440 445

005760"83E9960

Leu Pro Glu Gly Leu Tyr Thr Thr Leu Ala Tyr Asn Arg Ile Lys Pro
 450 455 460

Lys Ser Val Ser Asn Arg Pro Gly Leu Ser Leu Arg Ser Tyr Ala Leu
 465 470 475 480

Asp Ala Val Gln Pro Ser Arg Tyr Val Leu Gly Phe Gly Tyr Asp Gln
 485 490 495

Pro Glu Gly Lys Trp Gly Ala Asn Ile Met Leu Thr Tyr Ser Lys Gly
 500 505 510

Lys Asn Pro Asp Glu Leu Ala Tyr Leu Ala Gly Asp Gln Lys Arg Tyr
 515 520 525

Ser Thr Lys Arg Ala Ser Ser Ser Trp Ser Thr Ala Asp Val Ser Ala
 530 535 540

Tyr Leu Asn Leu Lys Lys Arg Leu Thr Leu Arg Ala Ala Ile Tyr Asn
 545 550 555 560

Ile Gly Asn Tyr Arg Tyr Val Thr Trp Glu Ser Leu Arg Gln Thr Ala
 565 570 575

Glu Ser Thr Ala Asn Arg His Gly Gly Asp Ser Asn Tyr Gly Arg Tyr
 580 585 590

Ala Ala Pro Gly Arg Asn Phe Ser Leu Ala Leu Gly Met Lys Phe
 595 600 605

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 18 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

AAACAGGTCT CGGCATAG

18

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 27 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

0065330-04900

CGCGAATTCA AACAGGTCTC GGCATAG

27

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 33 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

CGCGAATTCA AAAACTTCCA TTCCAGCGAT ACG

33

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

TAAAACTTCC ATTCCAGCGA TACG

24

What is claimed is:

1. An isolated and purified recombinant nucleic acid encoding a hemoglobin receptor protein having an amino acid sequence that is the amino acid sequence depicted as Seq. I.D. No. 6.
2. A recombinant expression construct comprising a nucleic acid that encodes a hemoglobin receptor protein from a *Neisseria* species having an amino acid sequence that is the amino acid sequence depicted as Seq. I.D. No. 6.
3. A transformed cell culture comprising the recombinant expression construct of Claim 2.
4. An isolated and purified recombinant nucleic acid encoding a hemoglobin receptor protein having an amino acid sequence that is the amino acid sequence depicted as Seq. I.D. No. 8.
5. A recombinant expression construct comprising a nucleic acid that encodes a hemoglobin receptor protein from a *Neisseria* species having an amino acid sequence that is the amino acid sequence depicted as Seq. I.D. No. 8.
6. A transformed cell culture comprising the recombinant expression construct of Claim 5.

ABSTRACT OF THE DISCLOSURE

The present invention relates to novel bacterial hemoglobin receptor proteins and genes that encode such proteins. The invention is directed toward the isolation, characterization, diagnostic and therapeutic use of bacterial hemoglobin receptor proteins, nucleic acid encoding
5 such proteins, recombinant expression constructs comprising such nucleic acids and cells transformed therewith, and antibodies and epitopes of such hemoglobin receptor proteins. The invention relates particularly to hemoglobin receptor proteins and genes encoding such proteins from *Neisseria* species, especially *N. meningitidis* and serotypes thereof, and *N. gonorrhoeae*. Methods for the diagnostic and therapeutic use of the proteins, epitopes, antibodies and nucleic
10 acids of the invention are also provided, including the use of the proteins, epitopes, antibodies and nucleic acids of the invention for the production of vaccines effective in providing immunization of a human against infection by pathogenic bacteria of *Neisseria* species.

Figure 1

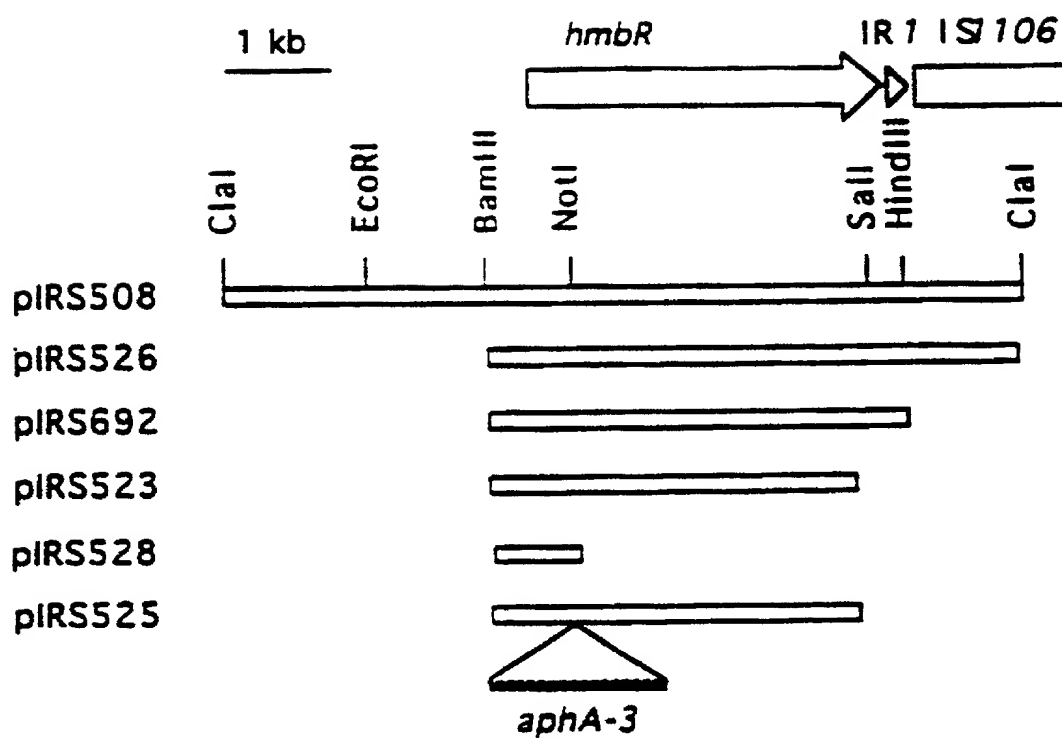


FIG. 2B

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	510	
ACAAATGCTCCCTATCGCCGCGCTGGTCCGGCAGTATTTTCGGCAATCCGGTCTTTGCGGC		
uGlnMetLeuProIleAlaAlaLeuValGlySerIlePheGlyAsnProValPheAlaAl		
560		
AGATGAAGCTGCAACTGAACCAACACCCCGTTAAGGCAGAGGTAAAGCAGTGCGCGTTAA		
aAspGluAlaAlaThrGluThrProValLysAlaGluValLysAlaValArgValLy		660
610		
AGGCCAGCGCAATGCGCCCTGCGGCTGTGGAACGCGTCAACCTTAACCGTATCAACAAGA		
sGlyGlnArgAsnAlaProAlaAlaValGluArgValAsnLeuAsnArgIleLysGlnGI		710
AATGATACGCGACAAACAAGACTTGGTGCGCTATTCCACCGATGTCGGCTTGAGCGACAG		
uMetIleArgAspAsnLysAspLeuValArgTyrSerThrAspValGlyLeuSerAspSe		760
CGGCCGCCCATCAAAAAGGCTTTGCTGTTCCGGCGGTGGAAAGCAACCGTGTCGGCGTGAG		
rGlyArgHisGlnLysGlyPheAlaValArgGlyValGluGlyAsnArgValGlyValSe		810
CATAGACGGCGTAACCTGCCTGATTCGGAAGAAACTCGCTGTACGCCCGTTATGGCAA		
rIleAspGlyValAsnLeuProAspSerGluGluAsnSerLeuTyrAlaArgTyrGlyAs		860
CTTCAACAGCTCGGCTCTGTCTATCGACCCCGAACTCGTGCGCAACATCGACATCGTAA		
nPheAsnSerSerArgLeuSerIleAspProGluLeuValArgAsnIleAspIleValLy		

FIG. 2D

TGAAGAGTCTTACAACCTGCTTGCTTCTTATTGGCGTGAAAGCTGACGATGTCAACAGACG IGluGluSerTyrAsnLeuLeuAlaSerTyrTrpArgGluAlaAspValAsnArgAr	1360	
GCGTAACACCAACCTCTTTACGAATGGACGCCGGAATCCGACCGGTTGTCTATGGTAAAGArgAsnThrAsnLeuPheTyrGluTrpThrProGluSerAspArgLeuSerMetValLy	1410	
AGCGGATGTCGATTATCAAAAACCAAAAGTATCTGCGGTCAACTACAAAGGTTTCGTCCCCsAlaAspValAspTyrGlnLysThrLysValSerAlaValAsnTyrLysGlySerPhePr	1460	1560
GATAGAGGATTCTTCACCTTGACACGTAACTACAATCAAAAGACTTGATGAAATCTA oIleGluAspSerSerThrLeuThrArgAsnTyrAsnGlnLysAspLeuAspGluIleTy		1610
CAACCGCAGTATGGATACCCGCTTCAAACGCATTACCCCTGCGTTTGGACAGCCATCCGTT rAsnArgSerMetAspThrArgPheLysArgIleThrLeuArgLeuAspSerHisProLe		1660
GCAACTCGGGGGGGGACACCGCCTGTTCGTTTAAAACTTTCGCCAGCCGCCGTGATTT uGlnLeuGlyGlyArgHisArgLeuSerPheLysThrPheAlaSerArgArgAspPh		1710
TGAAAAACCTAAACCGCGACGATTATTACTTCAGCGGCCGTGTGTTCGAACCAACAGCAG eGluAsnLeuAsnArgAspTyrTyrPheSerGlyArgValValArgThrThrSerSe		

FIG. 2E

1760
 TATCCAGCATCCGGTGAAACCAACCACTACGGTTTCTCACTGTCTGACCAAAATCAATG
 rIleGlnHisProValLysThrThrAsnTyrglyPheSerLeuSerAspGlnIleGlnTr
 1810
 GAACGACGTGTTCAGTAGCCGCGCAGGTATCCGTTACGATCATACCAAAATGACGCCTCA
 pAsnAspValPheSerSerArgAlaGlyIleArgTyraSpHisThrLysMETThrProGI
 1860
 1910
 GGAAATTGAATGCCGAGTGTGTCATGCTTGTGACAAACACCGCCTGCAGCCCAACTTATAA
 nGluLeuAsnAlaGluCysHisAlaCysAspLysThrProProAlaAlaAsnThrTyrlY
 1960
 AGGCTGGAGCGGTTTGTTCGGCTTGGCGGCGCAACTGAATCAGGCTTGGCGGTGTCGGTTA
 sGlyTrpSerGlyPheValGlyLeuAlaAlaGluLeuAsaGluAlaTrpArgValGlyTy
 2010
 CGACATTACTTCGGGTACCGTGTCCCCCAATGCGTCCGAAGTGTATTCTCACTTACAACCA
 rAspIleThrSerGlyTyraArgValProAsnAlaSerGluValTyraPheThrTyraSnHi
 2060
 CCGTTCCGGTAATTGGCTGCCCAATCCCAACCTGAAGCCGAGCGCACGACCCACAC
 sGlySerGlyAsnTrpLeuProAsnProAsnLeuLysAlaGluArgThrThrHisTh
 2110
 2160
 CCTCTCTCTGCAAGGCCGCGCAGCGAAAAAGGTACTTTGGATGCCAACCTGTATCAAAGCAA
 rLeuSerLeuGlnGlyArgSerGluLysGlyThrLeuAspAlaAsnLeuTyraGlnSerAs

FIG. 2G

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```

      2610      |      |      |      |
GCCTTGCAGAAAAGGTAAAGATTACCCGGTGGCTGAACAAAGTCGGCTTATGTGTTCCGA
rProLeuGlnLysLysValLysAspTyrProTrpLeuAsnLysSerAlaTyrValPheAs
      2660      |
TATGTACGGCTTCTACAAACCGGTGAAAAACCTGACTTTGCGTGCAGCGGTATATAATGT
pMetTyrGlyPheTyrLysProValLysAsnLeuThrLeuArgAlaGlyValTyrAsnVa
      2710      |      |      |      |
GTTCAACCGCAAAATACACCACTTGGGATTCCCTGCGCGCTGTATAGCTACAGCACCCAC
lPheAsnArgLysTyrThrTrpAspSerLeuArgGlyLeuTyrSerTyrSerThrTh
      2810      |      |      |      |
CAACTCGGTCGACCGCGATGGCAAAGGCTTAGACCGCTACCGCGCCCAAGCCGTAATTA
rAsnSerValAspArgAspGlyLysGlyLeuAspArgTyrArgAlaProSerArgAsnTy
      2860      |      |      |      |
CGCCGTATCGCTGGAAATGGAAAGTTTTTAATCTGGTATTATTGAATTAAATCGCCTTGTGAA
rAlaValSerLeuGluTrpLysPheSTOP
      2910      |      |      |      |
AATAAGCCGTCCGAATTGTGTTCAAGAACTCATTCGGACGGTTTTTACCGAATCTGTG
      2960      |      |      |      |
TGTGGGTTTATAGTGGATTAAACAAAAATCAGGACAAGGCCGACGAAGCCGCAGACAGTACA

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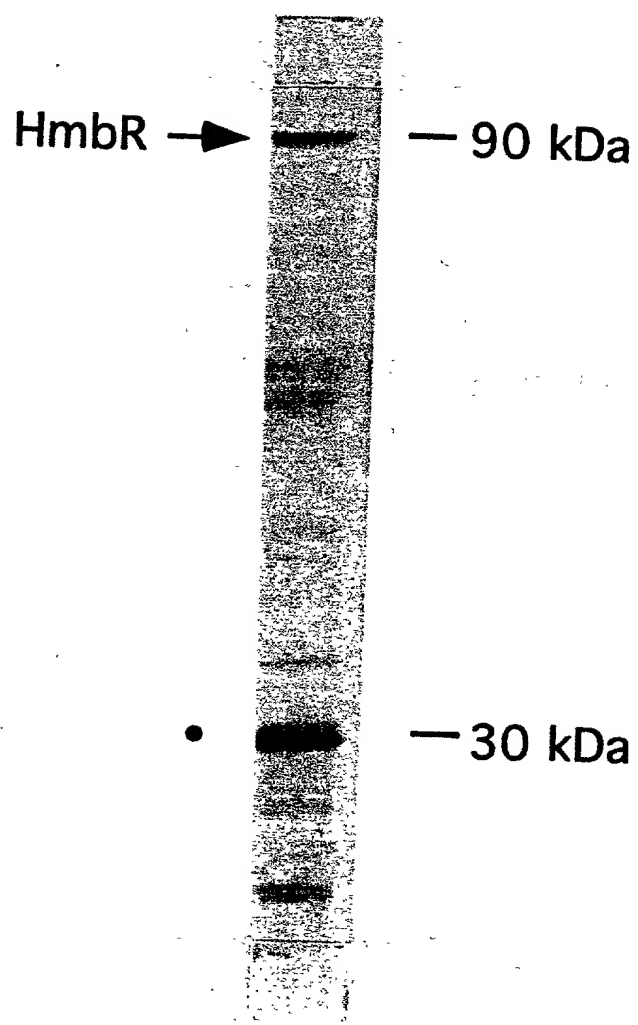

FIG. 2H

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3010 | | | | | 3060
GATAGTACGGAAACCGATTCACTTGGTGAGACCTTTGCAAAATTCCCTTCCCTCCCGACAG
-----> IS1106 3110
CCGAAACCCAAACACAGGTTTCGGCTGTTTCGCCCCCAAAATACCTCCTAATTCTACCCA
3160
AATACCCCTTAATCCTCCCCGATACCCGATAATCAGGCATCCGGCGCCTTTAGGCGGCA
3210
GCGGGCGCACTTAACCTGTTGGCGGCTTTCAAAAGGTTCAAAACACATCGCCTTCAGGTGC
3260
CTTTGCGCACTCACTTTAATCAGTCCGAAATAGGCCGCCCGGCATAGCAGAACTTACGG
3310
TGCAGCGTACCGAAGCTT
HindIII

Figure 3

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FIG. 4A

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TBP1M	MQQQHLFRLNILCLSLMTALPVYA--ENVQAEQAQEKQLDTIOVKAKKQ	47
LBPA	MNKKHGFQLTLTALAVAAAFPSYAANPETAAPDAAQTQSLKEVTVRAAKV	50
HMBR	MKPLQMLPIAALVGSIFGN-PVFAADEAATETTPVKAE-----VKAVR	43
	* * * *	
TBP1M	KTRRDNEVTGLGKLVKSSDTLSKEQVLNIRDLTRYDPGIAVVEQGRGASS	97
LBPA	-GRRSKEATGLGKIAKTSETLNKEQVLGIRDLTRYDPGVAVVEQNGASG	99
HMBR	KGQRNA-PAAVERV--NLNRIKQEMIRDNKKDLVRYSTDVGLSDSGRHQK-	89
	* * * *	
TBP1M	GYSIRGMDKNRVSLTVDGVSVQIQSYTAQAALGGTRTAGSSGAINIEIYEN	147
LBPA	GYSIRGVVDKNRVAVSVDGVAQIQAFTVQGSLSGYGGSGGAINIEIYEN	149
HMBR	GFAVRGVEGNRVGVSIDGVNLPDS--EENSLYARYGNFNSRSL- IDPEL	136
	* * * *	
TBP1M	VKAVEISKGSNSSEYNGALAGSVAFQTKTAADIIGEGKQWGIQSKTAYS	197
LBPA	I STVEIDKGAGSSDHSGGALGGAVAFRTKEAADLISDGKSWGIAKTAYG	199
HMBR	VRNIDI VKGADSFNTGSGALGGGVYNQTLQGRDLLPERQFGVMMKNGYS	186
	* * * *	
TBP1M	GKDHALTQSLALAGRSGGAELLIYTKRRGREIHAHKDAGKGVQ-SFNRL	246
LBPA	SKNRQFMKSLGAGFSKDGWEGLLIRTERQGRETHPHGDIADGVAYGINRL	249
HMBR	TRNREWTNTLFGFVSNDRVDAALLYSQRRGHETESAG-----	223
	* * * *	

FIG. 4B

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TBP1M	PICRFGNNTYT-DCTPRNIGGNGYYAAVQDNVRLGRWADVAGIRYDYRS	601
LBPA	SVCGYIETLSRCKVPRKINGSNIHISLNDRFSIGKYFDFSLGGRYDRKN	635
HMBR	-----SSIQHPVKTTNYGFSLSDIQWNDVFSSRAGIRYDHTK	460
*.....*	
TBP1M	THSED-----KSVSTGTHRNL SWNAGVVLKP--FTWMDLTYRSTGF	641
LBPA	FTTSE-----ELVRSGRYVDRSWNSGIVFKP--NRHFSLSYRASSGF	675
HMBR	MTPQELNAECHACDKTPPAANTYKGSFVGLAAQLNQAWRVGYDITSGY	510
*.....*	
TBP1M	RLPSFAEMYGWRA---GESLKTLDLKPESFNREAGIVFKGDFGNLEAS	687
LBPA	RTPSFQELFGIDIIYHDYPPKGWQRPALKSEKAAANREIGLQWKDFFGLEIS	725
HMBR	RVPNASEVY-FTYNHGSWNWLPNPNLKAERTTTHTL SLQGRSEKGTLDAN	559
	* .. * .. ** .. ** .. *	
TBP1M	YFNNA YRDLIAFGYET--RTQNGQTSASGDPGYR-----	719
LBPA	SFRNRYTDMIAVADHKTKLPNQAGQLTEIDIRDYY-----	760
HMBR	LYQSNYRNFLS--EEQKLT-SGDVSC TQMNYYYGMC SNPYSEKLEWQM	605
	... * .. * .. *	
TBP1M	-NAQNARIAGINILGKIDWHGVWGGLPDG--LYSTLAYNR IKVKDADIRA	766
LBPA	-NAQNMSLQGVN ILGKIDWNGVYGKLPEG--LYTTLAYNR IKPKSVSNRP	807
HMBR	QNIDKARIRGIELTGRNLNVDKVASFVPEGWKLFGSLGYAKSLSG----	650
	* .. * .. * .. * .. *	

FIG. 4C

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TBP1M	DRFTSYLFDVAVQPSRYVLGLGYDHPDGIWGINMTFTYSKAKSVDE - - -	813
LBPA	GLSL-RSYALDAVQPSRYVLGFQYDQPEGKWGANIMLTYSKGKNPDE - - -	853
HMBR	DNSLLST - - - - QPLKVIAGIDYESPSEKWGVFSRLTYLGAKKVKDAQY	694

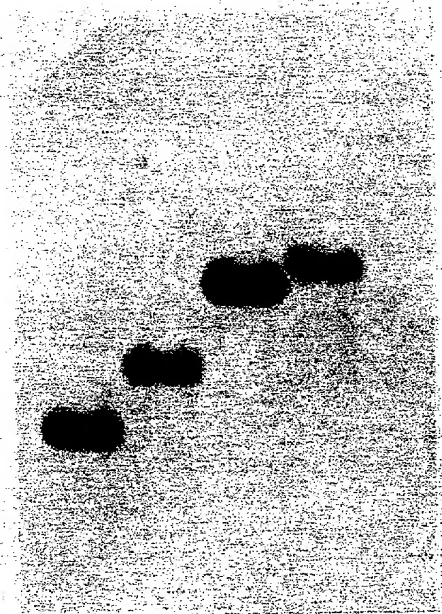
TBP1M	-LLGSQALLNGNANAKKAASRRTRPWYVTDVSGYVNIKKHLTLRAGVYNL	862
LBPA	-L - - - - AYLAGDQK - RYSTKRASSSWSTADVSAYLNKKRLTLRAAIYNI	897
HMBR	TVYENKGWGTPLQKKVKDYPWLNKSA YVFDMYGFYKPVKNLTLRAGVYNV	744

TBP1M	LNRYRYVTWENVVRQ - - TAGGAVNQHKNVGVYNNRYAAPGRNYTFSLMKF	908
LBPA	GNYRYVTWESLRQ - - TAESTANRRHGGDSNYGRYAAPGRNFSLALMKF	943
HMBR	FNRKYTTWDSLRGLYSYSTTNSVDRDGKGLDRYRAPSRNYAVSLEWKF	792

Figure 5

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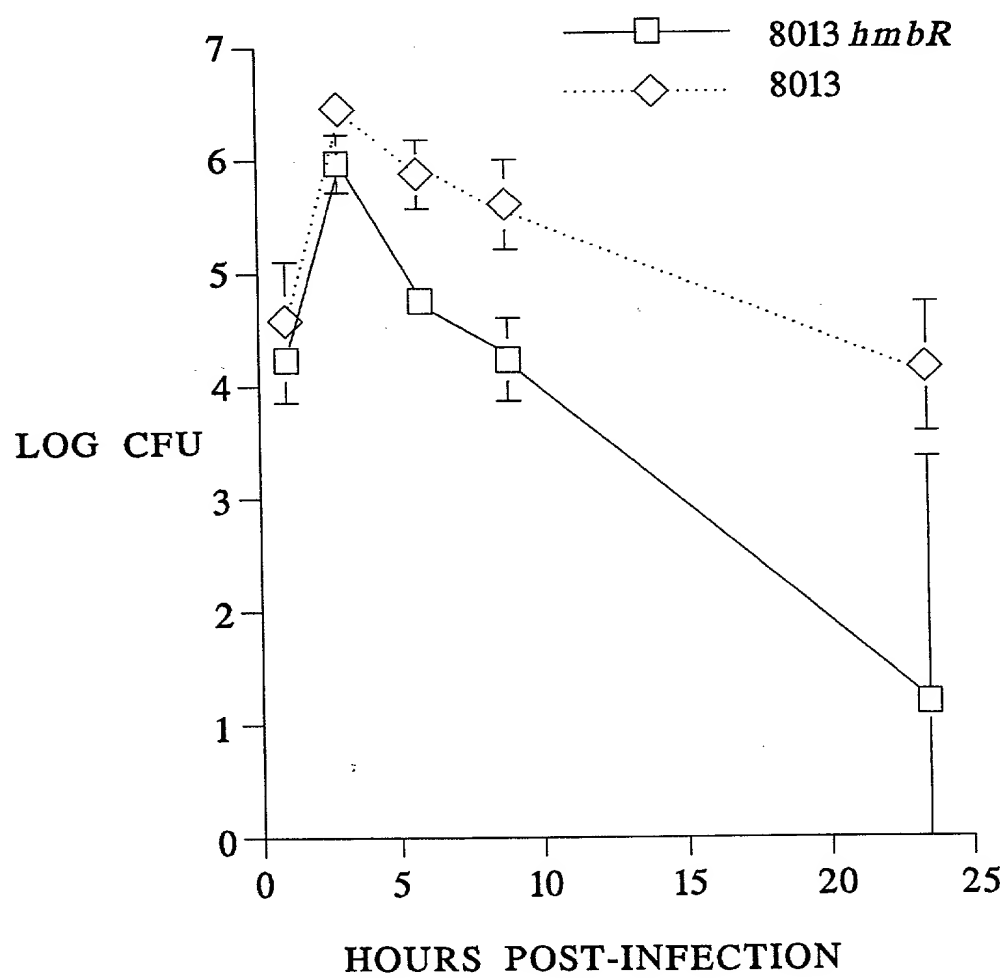
— 3 kb

— 2 kb

— 1 kb

FIG. 6

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FIG. 7A

ATG AAA CCA TTA CAA ATG CCC CCT ATC GCC GCG CTG CTC GGC AGT ATT	48
Met Lys Pro Leu Gln Met Pro Pro Ile Ala Ala Leu Leu Ser Ile	15
1	5
TTC GGC AAT CCG GTC TTT GCG GCA GAT GAA GCT GCA ACT GAA ACC ACA	96
Phe Gly Asn Pro Val Phe Ala Ala Asp Glu Ala Ala Thr Glu Thr Thr	30
20	25
CCC GTT AAG GCA GAG GTA AAA GCA GTG CGC GTT AAA GGT CAG CGC AAT	144
Pro Val Lys Ala Glu Val Lys Ala Val Arg Val Lys Gly Glu Arg Asn	45
35	40
GCG CCT GCG GCT GTG GAA GAG GTC AAC CTT AAC CGT ATC AAA CAA GAA	192
Ala Pro Ala Ala Val Glu Arg Val Asn Leu Asn Arg Ile Lys Gln Glu	60
50	55
ATG ATA CGC GAC AAT AAA GAC TTG GTG CGC TAT TCC ACC GAT GTC GGC	240
Met Ile Arg Asp Asn Lys Asp Leu Val Arg Tyr Ser Thr Asp Val Gly	80
65	70
TTG AGC GAC AGG AGC CGT CAT CAA AAA GGC TTT GCC ATT CGC GGC GTG	288
Leu Ser Asp Arg Ser Arg His Gln Lys Gly Phe Ala Ile Arg Gly Val	95
85	90

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FIG. 7B

GAA GGC GAC CGT GTC GGC GTT AGT ATT GAC GGC GTA AAC CTG CCT GAT	336
Glu Gly Asp Arg Val Ser Gly Val Ser Ile Asp Gly Val Asn Leu Pro Asp	110
100	105
TCC GAA GAA AAC TCG CTG TAC GCC CGT TAT GGC AAC TTC AAC AGC TCG	384
Ser Glu Glu Asn Ser Leu Tyr Ala Arg Tyr Gly Asn Phe Asn Ser Ser	125
115	120
CGT CTG TCT ATC GAC CCC GAA CTC GTG CGC AAC ATC GAC ATA GTA AAA	432
Arg Leu Ser Ile Asp Pro Glu Leu Val Arg Asn Ile Asp Ile Val Lys	140
130	135
GGG GCG GAC TCT TTC AAT ACC GGC AGC GGC GCC TTG GGC GGT GTG	480
Gly Ala Asp Ser Phe Asn Thr Gly Ser Gly Ala Leu Gly Gly Gly Val	150
145	155
AAT TAC CAA ACC CTG CAA GGA CGT GAC TTA CTG TTG CCT GAA CGG CAG	528
Asn Tyr Gln Thr Leu Gln Gly Arg Asp Leu Leu Pro Glu Arg Gln	165
160	170
TTC GGC GTG ATG ATG AAA AAC GGT TAC AGC ACG CGT AAC CGT GAA TGG	576
Phe Gly Val Met Met Lys Asn Gly Tyr Ser Thr Arg Asn Arg Glu Trp	180
185	190

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FIG. 7C

ACA AAT ACC CTC GGT TTC GGC GTG AGC AAC GAC CGC GTG GAT GCC GCT	624
Thr Asn Thr Leu Gly Phe Gly Val Ser Asn Asp Arg Val Asp Ala Ala	
195	200
TTG CTG TAT TCG CAA CGG CGC GGC CAT GAA ACT GAA AGC GCG GGC AAG	672
Leu Leu Tyr Ser Gln Arg Arg Gly His Glu Thr Glu Ser Ala Gly Lys	
210	215
CGT GGT TAT CCG GTA GAG GGT GCT GGT AGC GGA GCG AAT ATC CGT GGT	720
Arg Gly Tyr Pro Val Glu Gly Ala Gly Ser Gly Ala Asn Ile Arg Gly	240
225	235
TCT GCG CGC GGT ATT CCT GAT CCG TCC CAA CAC AAA TAC CAC AGC TTC	768
Ser Ala Arg Gly Ile Pro Asp Gln His Lys Tyr His Ser Phe	255
245	250
TTG GGT AAG ATT GCT TAT CAA ATC AAC GAC AAC CAC CAC CGC ATC GGC GCA	816
Leu Gly Lys Ile Ala Tyr Gln Ile Asn Asp Asn His Arg Ile Gly Ala	260
265	270
TCG CTC AAC GGT CAG CAG GGG CAT AAT TAC ACG GTT GAA GAG TCT TAC	864
Ser Leu Asn Gly Gln Gln Gly His Asn Tyr Thr Val Glu Ser Tyr	275
280	285

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FIG. 7D

AAC CTG CTT GCT TCT TAT TGG CGT GAA GCT GAC GAT GTC AAC AGA CCG	912
Asn Leu Leu Ala Ser Tyr Trp Arg Glu Ala Asp Asp Val Asn Arg Arg	
290	300
CGT AAC ACC AAC CTC TTT TAC GAA TGG ACG CCG GAA TCC GAC CCG TTG	960
Arg Asn Thr Asn Leu Phe Tyr Glu Trp Thr Pro Glu Ser Asp Arg Leu	
305	310
TCT ATG GTA AAA GCG GAT GTC GAT TAT CAA AAA ACC AAA TCT GCG	1008
Ser Met Val Lys Ala Asp Val Asp Tyr Gln Lys Thr Lys Val Ser Ala	
325	330
GTC AAC TAC AAA GGT TCG TTC CCG ACG AAT TAC ACC ACC ACA TGG GAA ACC	1056
Val Asn Tyr Lys Gly Ser Phe Pro Thr Asn Tyr Thr Thr Trp Glu Thr	
340	345
GAG TAC CAT AAA AAG GAA GTT GGC GAA ATC TAT AAC CGC AGC ATG GAT	1104
Glu Tyr His Lys Lys Glu Val Gly Glu Ile Tyr Asn Arg Ser Met Asp	
355	360
ACA ACC TTC AAA CGT ATT ACG CTG CGT ATG GAC AGC CAT CCG TTG CAA	1152
Thr Thr Phe Lys Arg Ile Thr Leu Arg Met Asp Ser His Pro Leu Gln	
370	375
	380
	385

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FIG. 7E

CTC	GGG	GGG	GGG	CGA	CAC	CGC	CTG	TCG	TTC	AAA	ACC	TTT	GCC	GGG	CAG	1200
Leu	Gly	Gly	Gly	Arg	His	Arg	Leu	Ser	Phe	Lys	Thr	Phe	Ala	Gly	Gln	
385				390						395					400	
CGT	GAT	TTT	GAA	AAC	TTA	AAC	CGC	GAC	GAT	TAC	TAC	TTC	AGC	GGC	CGT	1248
Arg	Asp	Phe	Glu	Asn	Leu	Asn	Arg	Asp	Asp	Tyr	Tyr	Phe	Ser	Gly	Arg	
				405					410					415		
GTT	GTT	CGA	ACC	ACC	AAC	AGT	ATC	CAG	CAT	CCG	GTG	AAA	ACC	ACC	AAC	1296
Val	Val	Arg	Thr	Thr	Asn	Ser	Ile	Gln	His	Pro	Val	Lys	Thr	Thr	Asn	
			420					425					430			
TAC	GGT	TTC	TCG	CTG	TCC	GAC	CAA	ATC	CAA	TGG	AAC	GAC	GTG	TTC	AGT	1344
Tyr	Gly	Phe	Ser	Leu	Ser	Asp	Gln	Ile	Gln	Trp	Asn	Asp	Val	Phe	Ser	
		435				440						445				
AGC	CGC	GCA	GGT	ATC	CGT	TAC	GAC	CAC	ACC	AAA	ATG	ACG	CCT	CAG	GAA	1392
Ser	Arg	Ala	Gly	Ile	Arg	Tyr	Asp	His	Thr	Lys	Met	Thr	Pro	Gln	Glu	
	450					455					460					
TTG	AAT	GCC	GAC	TGT	CAT	GCT	TGT	GAC	AAA	ACA	CCG	CCT	GCA	GCC	AAC	1440
Leu	Asn	Ala	Asp	Cys	His	Ala	Cys	Asp	Lys	Thr	Pro	Pro	Ala	Ala	Asn	
465					470					475					480	

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FIG. 7F

ACT TAT AAA GGC TGG AGC TTT GTC GGC TTG GCG GCG CAG CTG AGC	1488
Thr Tyr Lys Gly Trp Ser Gly Phe Val Gly Leu Ala Gln Leu Ser	
	485
CAA ACA TGG CGT TTG GGT TAC GAT GTG ACC TCA GGT TTC CGC GTG CCG	1536
Gln Thr Trp Arg Leu Gly Tyr Asp Val Thr Ser Gly Phe Arg Val Pro	
	500
	510
AAT GCG TCT GAA GTG TAT TTC ACT TAC AAC CAC GGT TCG GGC ACT TGG	1584
Asn Ala Ser Glu Val Tyr Phe Thr Tyr Asn His Gly Ser Gly Thr Trp	
	515
	520
	525
AAG CCT AAT CCT AAT TTG AAG GCA GAA CGC AGC ACC ACC CAC ACC CTG	1632
Lys Pro Asn Pro Asn Leu Lys Ala Glu Arg Ser Thr Thr His Thr Leu	
	530
	535
	540
TCC TTG CAG GGG CGC GGC GAC AAA GGG ACA CTG GAT GCC AAC CTG TAT	1680
Ser Leu Gln Gly Arg Gly Asp Lys Gly Thr Leu Asp Ala Asn Leu Tyr	
	545
	550
	555
CAA AGC AAT TAC CGA AAC TTC CTG TCG GAA GAG CAG AAT CTG ACT GTC	1728
Gln Ser Asn Tyr Arg Asn Phe Leu Ser Glu Glu Gln Asn Leu Thr Val	
	565
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	935
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	945
	950
	955
	960
	965
	970
	975
	980
	985
	990
	995

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FIG. 7G

AGC GGC ACA CCC GGC TGT ACT GAG GAG GAT GCT TAC TAC TAT AGA TGC	1776
Ser Gly Thr 580	
Pro Gly Cys Thr Glu Glu Asp Ala Tyr Tyr Arg Cys	590
585	
AGC GAC CCC TAC AAA GAA GAA AAA CTG GAT TGG CAG ATG AAA AAT ATC GAC	1824
Ser Asp 595	
Pro Tyr Lys Glu Lys Leu Asp Trp Gln Met Lys Asn Ile Asp	605
600	
AAG GCC AGA ATC CGC GGT GGT ATC GAG TTG ACA GGC CGT CTG AAT GTG GAC	1872
Lys Ala Arg Ile Arg Gly 615	
610	
Leu Thr Gly Arg Leu Asn Val Asp	620
AAA GTA GCG TCT TTT GTT GGT GAG GGT TGG AAA CTG TTC GGC TCG CTG	1920
Lys Val Ala Ser Phe 630	
625	
Leu Gly Trp Lys Leu Phe Gly Ser Leu 640	
GGT TAT GCG AAA AGC AAA CTG TCG GGC GAC AAC AGC CTG CTG TCC ACA	1968
Gly Tyr Ala Lys 645	
Ser Lys Leu Ser Gly Asp Asn Ser Leu Leu Ser Thr 655	
650	
CAG CCG CTG AAA GTG ATT GCC GGT ATC GAC TAT GAA AGT CCG AGC GAA	2016
Gln Pro Leu Lys Val Ile Ala Gly Ile Asp Tyr Glu Ser Pro Ser Glu	660
665	
670	

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FIG. 7H

AAA	TGG	GGC	GTA	TTC	TCC	CGC	CTG	ACC	TAT	CTA	GGC	GCG	AAA	AAG	GTC	2064
Lys	Trp	Gly	Val	Phe	Ser	Arg	Leu	Thr	Tyr	Leu	Gly	Ala	Lys	Lys	Val	
		675					680					685				
AAA	GAC	GGC	CAA	TAC	ACC	GTT	TAT	GAA	AAC	AAG	GGC	TGG	GGT	ACG	CCT	2112
Lys	Asp	Ala	Gln	Tyr	Thr	Val	Tyr	Glu	Asn	Lys	Gly	Trp	Gly	Thr	Pro	
		690				695					700					
TTG	CAG	AAA	AAG	GTA	AAA	GAT	TAC	CCG	TGG	CTG	AAC	AAG	TCG	GCT	TAT	2160
Leu	Gln	Lys	Lys	Val	Lys	Asp	Tyr	Pro	Trp	Leu	Asn	Lys	Ser	Ala	Tyr	
705					710					715					720	
GTG	TTT	GAT	ATG	TAC	GGC	TTC	TAC	AAA	CCG	GCT	AAA	AAC	CTG	ACT	TTG	2208
Val	Phe	Asp	Met	Tyr	Gly	Phe	Tyr	Lys	Pro	Ala	Lys	Asn	Leu	Thr	Leu	
				725					730					735		
CGT	GCA	GGC	GTG	TAC	AAC	CTG	TTC	AAC	CGC	AAA	TAC	ACC	ACT	TGG	GAT	2256
Arg	Ala	Gly	Val	Tyr	Asn	Leu	Phe	Asn	Arg	Lys	Tyr	Thr	Thr	Trp	Asp	
			740					745					750			
TCC	CTG	CGC	GGT	TTA	TAT	AGC	TAC	AGC	ACC	ACC	AAT	GCG	GTC	GAC	CGC	2204
Ser	Leu	Arg	Gly	Leu	Tyr	Ser	Tyr	Ser	Thr	Thr	Asn	Ala	val	Asp	Arg	
			755				760					765				

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FIG. 71

2352

GAT GGC AAA GGC TTA GAC CGC TAC CGC GCC CCA GGC CGC AAT TAC GCC
 Asp Gly Lys Gly Leu Asp Arg Tyr Arg Ala Pro Gly Arg Asn Tyr Ala
 770 775 780

2375

GTA TCG CTG GAA TGG AAG TTT TAA
 Val Ser Leu Glu Trp Lys Phe *
 785 790

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FIG. 8A

ATG AAA CCA TTA CAA ATG CTC CCT ATC GCC GCG CTG GTC GGC AGT ATT	48
Met Lys Pro Leu Gln Met Leu Pro Ile Ala Ala Leu Val Gly Ser Ile	
1	5
	10
	15
TTC GGC AAT CCG GTC TTT GCG GCA GAT GAA GCT GCA ACT GAA ACC ACA	96
Phe Gly Asn Pro Val Phe Ala Ala Asp Glu Ala Ala Thr Glu Thr Thr	
20	25
	30
CCC GTT AAG GCA GAG GTA AAA GCA GTG CGC GTT AAA GGC CAG CGC AAT	144
Pro Val Lys Ala Glu Val Lys Ala Val Arg Val Lys Gly Gln Arg Asn	
35	40
	45
GCG CCT GCG GCT GTG GAA CGC GTC AAC CTT AAC CGT ATC AAA CAA GAA	192
Ala Pro Ala Ala Val Glu Val Asn Leu Asn Arg Ile Lys Gln Glu	
50	55
	60
ATG ATA CGC GAC AAC AAC GAC TTT GCG TAT TCC ACC GAT GTC GGC	240
Met Ile Arg Asp Asn Lys Asp Leu Val Arg Tyr Ser Thr Asp Val Gly	
65	70
	75
	80
TTG AGC GAC AGC GGC GCG CAT CAA AAA GGC TTT GCT GTT CGC GGC GTG	288
Leu Ser Asp Ser Gly Arg His Gln Lys Gly Phe Ala Val Arg Gly Val	
85	90
	95

FIG. 8B

GAA GGC AAC CGT GTC GGC GTG AGC ATA GAC GGC GTA AAC CTG CCT GAT	336
Glu Gly Asn Arg Val Gly Val Ser Ile Asp Gly Val Asn Leu Pro Asp	
100	110
TCC GAA GAA AAC TCG CTG TAC GCC CGT TAT GGC AAC TTC AAC AGC TCG	384
Ser Glu Glu Asn Ser Leu Tyr Ala Arg Tyr Gly Asn Phe Asn Ser Ser	
115	125
CGT CTG TCT ATC GAC GAC CCC GAA CTC GTG CGC AAC ATC GAC ATC GTA AAA	432
Arg Leu Ser Ile Asp Pro Glu Leu Val Arg Asn Ile Asp Ile Val Lys	
130	140
GGG GCG GAC TCT TTC AAT ACC GGC AGC GGC GTG GGC GGC GGT GTG	480
Gly Ala Asp Ser Phe Asn Thr Gly Ser Gly Ala Leu Gly Gly Val	
145	150
AAT TAC CAA ACC CTG CAA GGA CGT GAC TTA CTG TTG CCT GAA CGG CAG	528
Asn Tyr Gln Thr Leu Gln Gly Arg Asp Leu Leu Pro Glu Arg Gln	
165	170
TTC GGC GTG ATG ATG AAA AAC GGT TAC AGC ACG CGT AAC CGT GAA TGG	576
Phe Gly Val Met Met Lys Asn Gly Tyr Ser Thr Arg Asn Arg Glu Trp	
180	185
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FIG. 8C

ACA AAT ACC CTC GGT TTC GGC GTG AGC AAC GAC CGC GTG GAT GCC GCT Thr Asn Thr Leu Gly Phe Gly Val Ser Asn Asp Arg Val Asp Ala Ala	195 200 205 624
TTG CTG TAT TCG CAA CGG CGC GGC CAT GAA ACT GAA AGC GCG GGC AAG Leu Leu Tyr Ser Gln Arg Arg Gly His Glu Thr Glu Ser Ala Gly Lys	210 215 220 672
CGT GGT TAT CCG GTA GAG GGT GCT GGT AGC GGA GCG AAT ATC CGT GGT Arg Gly Tyr Pro Val Glu Gly Ala Gly Ser Gly Ala Asn Ile Arg Gly	225 230 235 720
TCT GCG CGC GGT ATT CCT GAT CCG TCC CAA CAC AAA TAC CAC AGC TTC Ser Ala Arg Gly Ile Pro Asp Pro Ser Gln His Lys Tyr His Ser Phe	245 250 255 768
TTG GGT AAG ATT GCT TAT CAA ATC AAC GAC AAC CAC CGC ATC GGC GCA Leu Gly Lys Ile Ala Tyr Gln Ile Asn Asp Asn His Arg Ile Gly Ala	260 265 270 816
TCG CTC AAC GGT CAG CAG GGG CAT AAT TAC ACG GTT GAA GAG TCT TAC Ser Leu Asn Gly Gln Gln Gly His Asn Tyr Thr Val Glu Glu Ser Tyr	275 280 285 864

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FIG. 8D

AAC CTG CTT GCT TCT TAT TGG CGT GAA GCT GAC GAT GTC AAC AGA CGG	911
Asn Leu Leu Ala Ser Tyr Trp Arg Glu Ala Asp Val Asn Arg Arg	
290	300
CGT AAC ACC AAC CTC TTT TAC TAC GAA TGG ACG CCG GAA TCC GAC CGG TTG	960
Arg Asn Thr Asn Leu Phe Tyr Glu Trp Thr Pro Glu Ser Asp Arg Leu	
305	310
	315
TCT ATG GTA AAA GCG GAT GTC GAT TAT CAA AAA ACC AAA GTA TCT GCG	1008
Ser Met Val Lys Ala Asp Tyr Tyr Gln Lys Thr Lys Val Ser Ala	
325	330
	335
GTC AAC TAC AAA GGT TCG TTC CCG ATA GAG GAT TCT TCC ACC TTG ACA	1056
Val Asn Tyr Lys Gly Ser Phe Pro Ile Glu Asp Ser Thr Leu Thr	
340	345
	350
CGT AAC TAC AAT CAA AAG GAC TTG GAT GAA ATC TAC AAC CGC AGT ATG	1104
Arg Asn Tyr Asn Gln Lys Asp Leu Asp Glu Ile Tyr Asn Arg Ser Met	
355	360
	365
GAT ACC CGC TTC AAA CGC ATT ACC CTG CGT TTG GAC AGC CAT CCG TTG	1152
Asp Thr Arg Arg Phe Lys Arg Ile Thr Leu Arg Leu Asp Ser His Pro Leu	
370	375
	380

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FIG. 8E

CAA CTC GGG GGG GGG CGA CAC CGC CTG TCG TTT AAA ACT TTC GCC AGC	1200
Gln Leu Gly Gly Gly Arg His Arg Leu Ser Phe Lys Thr Phe Ala Ser	400
385	395
CGC CGT GAT TTT GAA AAC CTA AAC CGC GAC GAT TAT TAC TTC AGC GGC	1248
Arg Arg Asp Phe Glu Asn Leu Asn Arg Asp Tyr Tyr Phe Ser Gly	415
405	410
CGT GTT GTT CGA ACC ACC AGC AGT ATC CAG CAT CCG GTG AAC ACC ACC	1296
Arg Val Val Arg Thr Thr Thr Ser Ser Ile Gln His Pro Val Lys Thr Thr	430
420	425
AAC TAC GGT TTC TCA CTG TCT GAC CAA ATT CAA TGG AAC GAC GTG TTC	1344
Asn Tyr Gly Phe Ser Leu Ser Asp Gln Ile Gln Trp Asn Asp Val Phe	445
435	440
AGT AGC CGC GCA GGT ATC CGT TAC GAT CAT ACC AAA ATG ATG CCT CAG	1392
Ser Ser Arg Ala Gly Ile Arg Tyr Asp His Thr Lys Met Thr Pro Gln	455
450	460
GAA TTG AAT GCC GAG TGT CAT GCT TGT GAC AAA ACA CCG CCT GCA GCC	1440
Glu Leu Asn Ala Glu Cys His Ala Cys Asp Lys Thr Pro Pro Ala Ala	470
465	475
	480

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FIG. 8F

AAC	ACT	TAT	AAA	GGC	TGG	AGC	GGT	TTT	GTC	GGC	TTG	GCG	GCG	CAA	CTG	1488
Asn	Thr	Tyr	Lys	Gly	Trp	Ser	Gly	Phe	Val	Gly	Leu	Ala	Ala	Gln	Leu	
				485					490					495		
AAT	CAG	GCT	TGG	CGT	GTC	GGT	TAC	GAC	ATT	ACT	TCC	GGC	TAC	CGT	GTC	1536
Asn	Gln	Ala	Trp	Arg	Val	Gly	Tyr	Asp	Ile	Thr	Ser	Gly	Tyr	Arg	Val	
			500					505					510			
CCC	AAT	GCG	TCC	GAA	GTG	TAT	TTC	ACT	TAC	AAC	CAC	GGT	TCG	GGT	AAT	1584
Pro	Asn	Ala	Ser	Glu	Val	Tyr	Phe	Thr	Tyr	Asn	His	Gly	Ser	Gly	Asn	
			515				520					525				
TGG	CTG	CCC	AAT	CCC	AAC	CTG	AAA	GCC	GAG	CGC	ACG	ACC	ACC	CAC	ACC	1632
Trp	Leu	Pro	Asn	Pro	Asn	Leu	Lys	Ala	Glu	Arg	Thr	Thr	Thr	His	Thr	
	530					535					540					
CTC	TCT	CTG	CAA	GGC	CGC	AGC	GAA	AAA	GGT	ACT	TTG	GAT	GCC	AAC	CTG	1680
Leu	Ser	Leu	Gln	Gly	Arg	Ser	Glu	Lys	Gly	Thr	Leu	Asp	Ala	Asn	Leu	
545					550					555				560		
TAT	CAA	AGC	AAT	TAC	CGC	AAT	TTC	CTG	TCT	GAA	GAG	CAG	AAG	CTG	ACC	1728
Tyr	Gln	Ser	Asn	Tyr	Arg	Asn	Phe	Leu	Ser	Glu	Glu	Gln	Lys	Leu	Thr	
				565				570						575		

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FIG. 8G

ACC AGC GGC GAT GTC AGC TGT ACT CAG ATG AAT TAC TAC GGT ATG	1776
Thr Ser Gly Asp Val Ser Cys Thr Gln Met Asn Tyr Tyr Gly Met	
	580
	585
	590
TGT AGC AAT CCT TAT TCC GAA AAA CTG GAA TGG CAG ATG CAA AAT ATC	1824
Cys Ser Asn Pro Tyr Ser Glu Lys Leu Glu Trp Gln Met Gln Asn Ile	
	595
	600
	605
GAC AAG GCC AGA ATC CGC GGT ATC GAG CTG ACG GGC CGT CTG AAT GTG	1872
Asp Lys Ala Arg Ile Arg Gly Ile Glu Leu Thr Gly Arg Leu Asn Val	
	610
	615
	620
GAC AAA GTA GCG TCT TTT GTT CCT GAG GGC TGG AAA CTG TTC GGC TCG	1920
Asp Lys Val Ala Ser Phe Val Pro Glu Gly Trp Lys Leu Phe Gly Ser	
	625
	630
	635
	640
CTG GGT TAT GCG AAA AGC AAA CTG TCG GGC GAC AAC AGC CTG CTG TCC	1968
Leu Gly Tyr Ala Lys Ser Lys Leu Ser Gly Asp Asn Ser Leu Leu Ser	
	645
	650
	655
ACC CAG CCG TTG AAA GTG ATT GCC GGT ATC GAC TAT GAA AGT CCG AGC	2016
Thr Gln Pro Leu Lys Val Ile Ala Gly Ile Asp Tyr Glu Ser Pro Ser	
	660
	665
	670

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FIG. 8H

GAA AAA	TGG	GGC	GTG	TTC	TCC	CGC	CTG	ACC	TAT	CTG	GGC	GCG	AAA	AAG	2064
Glu Lys	Trp	Gly	Val	Phe	Ser	Arg	Leu	Thr	Tyr	Leu	Gly	Ala	Lys	Lys	
	675					680					685				
GTC AAA	GAC	GCG	CAA	TAC	ACC	GTT	TAT	GAA	AAC	AAG	GGC	TGG	GGT	ACG	2112
Val Lys	Asp	Ala	Gln	Tyr	Thr	Val	Tyr	Glu	Asn	Lys	Gly	Trp	Gly	Thr	
	690				695					700					
CCT TTG	CAG	AAA	AAG	GTA	AAA	GAT	TAC	CCG	TGG	CTG	AAC	AAG	TCG	GCT	2160
Pro Leu	Gln	Lys	Lys	Val	Lys	Asp	Tyr	Pro	Trp	Leu	Asn	Lys	Ser	Ala	
	705			710					715					720	
TAT GTG	TTC	GAT	ATG	TAC	GGC	TTC	TAC	AAA	CCG	GTG	AAA	AAC	CTG	ACT	2208
Tyr Val	Phe	Asp	Met	Tyr	Gly	Phe	Tyr	Lys	Pro	Val	Lys	Asn	Leu	Thr	
			725					730					735		
TTG CGT	GCA	GGC	GTA	TAT	AAT	GTG	TTC	AAC	CGC	AAA	TAC	ACC	ACT	TGG	2256
Leu Arg	Ala	Gly	Val	Tyr	Asn	Val	Phe	Asn	Arg	Lys	Tyr	Thr	Thr	Trp	
			740				745					750			
GAT TCC	CTG	CGC	GGC	CTG	TAT	AGC	TAC	AGC	ACC	ACC	AAC	TCG	GTC	GAC	2304
Asp Ser	Leu	Arg	Gly	Leu	Tyr	Ser	Tyr	Ser	Thr	Thr	Asn	Ser	Val	Asp	
			755			760						765			

FIG. 8I

[illegible]

GCC	GTA	TCG	CTG	GAA	TGG	AAG	TTT	TAA	
Ala	Val	Ser	Leu	Glu	Trp	Lys	Phe	*	
									785
									790
									2379

FIG. 9A

ATG AAA CCA TTA CAC ATG CTT CCT ATT GCC GCG CTG GTC GGC AGT ATT	48
Met Lys Pro Leu His Met Leu Pro Ile Ala Ala Leu Val Gly Ser Ile	15
1	5
TTC GGC AAT CCG GTC TTG GCA GCG GAT GAA GCT GCA ACC GAA ACC ACA	96
Phe Gly Asn Pro Val Leu Ala Ala Asp Glu Ala Ala Thr Glu Thr Thr	30
20	25
CCC GTT AAA GCA GAG ATA AAA GAA GTG CGC GTT AAA GAC CAG CTT AAT	144
Pro Val Lys Ala Glu Ile Lys Glu Val Arg Val Lys Asp Gln Leu Asn	45
35	40
GCG CCT GCA ACC GTG GAA CGT GTC AAC CTC GGC CGC ATT CAA CAG GAA	192
Ala Pro Ala Thr Val Glu Arg Val Asn Leu Gly Arg Ile Gln Gln Glu	60
50	55
ATG ATA CGC GAC AAC AAA GAC TTG GTG CGT TAC TCC ACC GAC GTC GGC	240
Met Ile Arg Asp Asn Lys Asp Leu Val Arg Tyr Ser Thr Asp Val Gly	80
65	70
TTG AGC GAT AGC GGC CGC CAT CAA AAA GGC TTT GCT GTG CGC GGC GTG	288
Leu Ser Asp Ser Gly Arg His Gln Lys Gly Phe Ala Val Arg Gly Val	95
85	90

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FIG. 9B

GAA GGC AAC CGT GTC GGT GTC AGC ATT GAC GGC GTG AGC CTG CCT GAT Glu Gly Asn Arg Val Gly Val Ser Ile Asp Gly Val Ser Leu Pro Asp 100 105 110 336
TCG GAA GAA AAC TCA CTG TAT GCA CGT TAT GGC AAC TTC AAC AGC TCG Ser Glu Glu Asn Ser Tyr Ala Arg Tyr Gly Asn Phe Asn Ser Ser 115 120 125 384
CGC CTG TCT ATC GKC CCC GAA CTC GTG CGC AAC ATC GAA ATC GCG AAG Arg Leu Ser Ile Asp Pro Glu Leu Val Arg Asn Ile Glu Ile Ala Lys 130 135 140 432
GGC GCT GAC TCT TTC AAT ACC GGT AGC GGC GCA TTG GGT GGC GGC GTG Gly Ala Asp Ser Phe Asn Thr Gly Ser Gly Ala Leu Gly Gly Gly Val 145 150 155 160 480
AAT TAC CAA ACC CTG CAA GGA CAT GAT TTG CTG TTG GAC GAC AGG CAA Asn Tyr Gln Thr Leu Gln Gly His Asp Leu Leu Asp Asp Arg Arg Gln 165 170 175 528
TTC GGC GTG ATG ATG AAA AAC GGT TAC AGC AGC CGC AAC CGC GAA TGG Phe Gly Val Met Met Lys Asn Gly Tyr Ser Ser Arg Asn Arg Glu Trp 180 185 190 576

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FIG. 9C

ACA AAT ACA CTC GGT TTC GGT GTG AGC AAC GAC CGC GTG GAT GCC GCT Thr Asn Thr Leu Gly Phe Gly Val Ser Asn Asp Arg Val Asp Ala Ala	195	200	205	624
TTG CTG TAT TCG CAA CGT CGC GGT CAT GAG ACC GAA AGC GCG GGC GAG Leu Leu Tyr Ser Gln Arg Arg Gly His Glu Thr Glu Ser Ala Gly Glu	210	215	220	672
CGT GGC TAT CCG GTA GAG GGT GCT GGC AGC GGA GCA ATT ATC CGT GGT Arg Gly Tyr Pro Val Glu Gly Ala Gly Ser Gly Ala Ile Ile Arg Gly	225	230	235	720
TCG TCA CGC GGT ATC CCT GAT CCG TCC AAA CAC AAA TAC CAC AAC TTC Ser Ser Arg Gly Ile Pro Asp Pro Ser Lys His Lys Tyr His Asn Phe	245	250	255	768
TTG GGT AAG ATT GCT TAT CAA ATC AAC GAC AAG CAC CAC CGC ATC GGC CCA Leu Gly Lys Ile Ala Tyr Gln Ile Asn Asp Lys His His Arg Ile Gly Pro	260	265	270	816
TCG TTT AAC GGC CAG CAG GGG CAT AAT TAC ACG ATT GAA GAG TCT TAT Ser Phe Asn Gly Gln Gln Gly His Asn Tyr Thr Ile Glu Glu Ser Tyr	275	280	285	864

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FIG. 9D

AAC CTG ACC GCT TCT TCC TGG CGC GAA GCC GAT GAC GTA AAC AGA CGG	912
Asn Leu Thr Ala Ser TCT TCC TGG Trp Arg Glu Ala Asp Asp Val Asn Arg Arg	
290	295
CGC AAT GCC AAC CTC TTT TAC GAA TGG ACG CCT GAT TCA AAT TGG CTG	960
Arg Asn Ala Asn Leu Phe Tyr Glu Trp Thr Pro Asp Ser Asn Trp Leu	
305	310
TCG TCT TTG AAG GCG GAC TTC GAT TAT CAG ACA ACC AAA GTG GCG GCG	1008
Ser Ser Leu Lys Ala Asp 325	330
	335
GTT AAC AAC AAA GGC TCG TTC CCG ACG GAT TAT TCC ACC TGG ACG CGC	1056
Val Asn Asn Lys Gly 340	345
	350
AAC TAT AAT CAG AAG GAT TTG GAG AAT ATA TAC AAC CGC AGC ATG GAC	1104
Asn Tyr Asn Gln Lys Asp Leu Glu Asn Ile Tyr Asn Arg Ser Met Asp	
355	360
	365
ACC CGA TTC AAA CGT TTT ACT TTG CGT ATG GAC AGC CAA CCG TTG CAA	1152
Thr Arg Phe Lys Arg Phe Thr Leu Arg Met Asp Ser Gln Pro Leu Gln	
370	375
	380

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FIG. 9E

CTG GGC GGC CAA CAT CGC TTG TCG CTT AAA ACT TTC GCC AGT CGG CGT	1200
Leu Gly Gly Gln His Arg Leu Ser Leu Lys Thr Phe Ala Ser Arg	400
385	395
GAG TTT GAA AAC TTA AAC CGC GAC GAT TAT TAC TTC AGC GAA AGA GTA	1248
Glu Phe Glu Asn Leu Asn Arg Asp Tyr Tyr Phe Ser Glu Arg Val	415
405	410
TCC CGT ACT ACC AGC TCG ATT CAA CAC CCC GTG AAA ACC ACT AAT TAT	1296
Ser Arg Thr Thr Ser Ser Ile Gln His Pro Val Lys Thr Thr Asn Tyr	430
420	425
GGT TTC TCA CTG TCT GAT CAA ATC CAA TGG AAC GAC GTG TTC AGC AGC	1344
Gly Phe Ser Leu Ser Asp Gln Ile Gln Trp Asn Asp Val Phe Ser Ser	445
435	440
CGT GCA GAT ATC CGT TAC GAT CAT ACC AAA ATG ACG CCT CAG GAA TTG	1392
Arg Ala Asp Ile Arg Tyr Asp His Thr Lys Met Thr Pro Gln Glu Leu	460
450	455
AAT GCC GAG TGT CAT GCT TGT GAC AAA ACA CCG CCT GCA GCC AAT ACT	1440
Asn Ala Glu Cys His Ala Cys Asp Lys Thr Pro Pro Ala Ala Asn Thr	480
465	470
	475

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FIG. 9F

TAT AAA GGC TGG AGC GGA TTT GTC GGT TTT GCG GCG CAA CTG AAT CAG	1488
Tyr Lys Gly Trp Ser Gly Phe Val Gly Leu Ala Ala Gln Leu Asn Gln	495
485	490
GCT TGG CAT GTC GGT TAC GAC ATT ACT TCC GGC TAC CGT GTC CCC AAT	1536
Ala Trp His Val Gly Tyr Asp Ile Thr Ser Gly Tyr Arg Val Pro Asn	510
500	505
GCG TCC GAA GTG TAT TTC ACT TAC AAC CAC GGT TCG GGT AAT TGG CTG	1584
Ala Ser Glu Val Tyr Phe Thr Tyr Asn His Gly Ser Gly Asn Trp Leu	525
515	520
CCC AAT CCC AAC CTG AAA GCC GAG CGC AGC ACC ACC CAC ACC CTG TCT	1632
Pro Asn Pro Asn Leu Lys Ala Thr Arg Ser Thr Thr His Thr Leu Ser	540
530	535
CTG CAA GGC CGC AGC GAA AAA GGT ACT TTT GAT GCC AAC CTG TAT CAA	1680
Leu Gln Gly Arg Ser Glu Lys Gly Thr Leu Asp Ala Asn Leu Tyr Gln	550
545	555
AAC AAT TAC CGC AAC TTC TTG TCT GAA GAG CAG AAG CTG ACC ACC AGC	1728
Asn Asn Tyr Arg Asn Phe Leu Ser Glu Gln Lys Leu Thr Thr Ser	570
565	575

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FIG. 9G

GCC GAT GTC GGC TGT ACT CAG ATG AAT TAC TAC TAC GGT ATG TGT AGC Gly Asp Val Gly Cys Thr Gln Met Asn Tyr Tyr Tyr Gly Met Cys Ser	580	585	590	1776
AAT CCT TAT TCC GAA AAA CCG GAA TGG CAG ATG CAA AAT ATC GAT AAG Asn Pro Tyr Ser Glu Lys Pro Glu Trp Gln Met Gln Asn Ile Asp Lys	595	600	605	1824
GCC CGA ATC CGT GGT CTT GAG CTG ACA GGC CGT CTG AAT GTG ACA AAA Ala Arg Ile Arg Arg Gly Leu Glu Leu Thr Gly Arg Leu Asn Val Thr Lys	610	615	620	1872
GTA GCG TCT TTT GTT CCT GAG GGC TGG AAA TTG TTC GGC TCG CTG GGT Val Ala Ser Phe Val Pro Pro Glu Gly Trp Lys Leu Phe Gly Ser Leu Gly	625	630	635	1920
TAT GCG AAA AGC AAA CTG TCG GGC GAC AAC AGC CTG CTG TCC ACA CAG Tyr Ala Lys Ser Lys Leu Ser Gly Asp Asn Ser Ser Leu Leu Thr Gln	645	650	655	1968
CCG CCG AAA GTG ATT GCC GGT GTC GAC TAC GAA AGC CCG AGC GAA AAA Pro Pro Lys Val Ile Ala Gly Val Asp Tyr Glu Ser Pro Ser Glu Lys	660	665	670	2016

FIG. 9H

2064

2112

2160

2208

2256

2304

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FIG. 9I

2352

GGC AAA GGC TTA GAC CGC TAC CGC GCC TCA GGC CGT AAT TAC GCC GTA
Gly Lys Gly Leu Asp Arg Tyr Arg Ala Ser Gly Arg Asn Tyr Ala Val

780

775

770

2378

TCG CTG GAT TGG AAG TTT TGA ATTCC
Ser Leu Asp Trp Lys Phe *
785 790

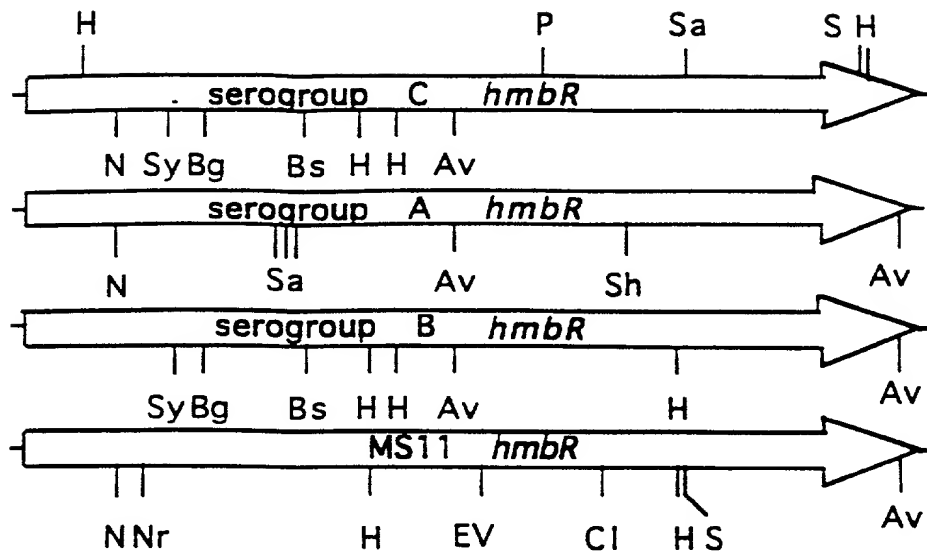
**Figure 10****SHEET 43/47**

FIG. 11A

HMBRA	MKPLQMLPIAALVGSIFGNPVLAADEAAATETTPVKAEIKAVRVKGQRNAP	50
HMBRB	MKPLQMPPIAALLGSI FGNPVFAXDEAAATETTPVKAEVKA V RVKGQRNAP	50
HMBRC	MKPLQMLPIAALVGSIFGNPVFAADEAAATETTPVKAEVKA V RVKGQRNAP	50
HMBRMS11	MKPLHMLPIAALVGSIFGNPVLAADEAAATETTPVKAEIKEVRVKDQLNAP	50
	**** * **** * **** * **** * **** * **** * **** *	

HMBRA	AAVERVNLNRIKQEMI RDNKDLVRYSTDVGLSDSGRHQKGF	AVRGVEG	NR	100
HMBRB	AAVERVNLNRIKQEMI RDNKDLVRYSTDVGLSDSGRHQKGF	AI RGV	EGDR	100
HMBRC	AAVERVNLNRIKQEMI RDNKDLVRYSTDVGLSDSGRHQKGF	AVRGVEG	NR	100
HMBRMS11	ATVERVNLGRIQQEMI RDNKDLVRYSTDVGLSDSGRHQKGF	AVRGVEG	NR	100

HMBRA	VGVSIDGVNLPDSEENSLYARYGNFNSSRLSIDPELVNIEIVKGADSFN	150
HMBRB	VGVSIDGVNLPDSEENSLYARYGNFNSSRLSIDPELVNIDIVKGADSFN	150
HMBRC	VGVSIDGVNLPDSEENSLYARYGNFNSSRLSIDPELVNIDIVKGADSFN	150
HMBRMS11	VGVSIDGVSLPDSEENSLYARYGNFNSSRLSIDPELVNIEIAKGADSFN	150
	*****	*****

HMBRA	TGSGALGGGVNYQT	LQGRDLLDD	RQFGVMMKNGYSTR	NREWTNTL	FGGV	200
HMBRB	TGSGALGGGVNYQT	LQGRDLLLP	RQFGVMMKNGYSTR	NREWTNTL	FGGV	200
HMBRC	TGSGALGGGVNYQT	LQGRDLLLP	RQFGVMMKNGYSTR	NREWTNTL	FGGV	200
HMBRMS11	TGSGALGGGVNYQT	LQGHDLLDD	RQFGVMMKNGYSSR	NREWTNTL	FGGV	200
	*****	*****	*****	*****	*****	*****

FIG. 11B

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HMBRA SDRVDAALLYQRRGHETESAGNRGYPVEGAGKETNIRGSARGIPDPSK 250
 HMBRB SDRVDAALLYQRRGHETESAGKRGPVEGAGSGANIRGSARGIPDPSQ 250
 HMBRC SDRVDAALLYQRRGHETESAGKRGPVEGAGSGANIRGSARGIPDPSQ 250
 HMBRMS11 SDRVDAALLYQRRGHETESAGERGYPVEGAGSGAIIRGSSRGIPDPSK 250

HMBRA HKYHNFELGKIAYQINDNHRIGASLNGQQGHNYTVEESYNLTASSWREADD 300
 HMBRB HKYHSFLGKIAYQINDNHRIGASLNGQQGHNYTVEESYNLLASYWREADD 300
 HMBRC HKYHSFLGKIAYQINDNHRIGASLNGQQGHNYTVEESYNLLASYWREADD 300
 HMBRMS11 HKYHNFELGKIAYQINDKHRIGPSFNGQQGHNYTIEESYNLTASSWREADD 300

HMBRA VNRRRNANLFYEWMPDSNWLSSLKADFYQKTKVAAIN-KGSFPT-NYTT 348
 HMBRB VNRRNTNLFYEWTPESDRLSMVKADVDYQKTKVSAVNYKGSFPT-NYTT 349
 HMBRC VNRRNTNLFYEWTPESDRLSMVKADVDYQKTKVSAVNYKGSFPIEDSST 350
 HMBRMS11 VNRRRNANLFYEWTPDSNWLSSLKADFYQTTKVAAVNKNKGSFPTD-YST 349

HMBRA WETEHKKKEVGEIYNRSMDTRFKRFTLRDLSHPLQLGGGRHRLSFKTFAS 398
 HMBRB WETEHKKKEVGEIYNRSMDTTFKRITLRMDSHPLQLGGGRHRLSFKTFAG 399
 HMBRC LTRNYNQKDLDEIYNRSMDTRFKRITLRDLSHPLQLGGGRHRLSFKTFAS 400
 HMBRMS11 WTRNYNQKDLNENIYNRSMDTRFKRFTLRMDSQPLQLGG-RHRLSLKTFAS 398

FIG. 11C

HMBRA	RRDFENLRDDYYFSGRVVRTTSSIQHPVKTTNYGFSLSQIQWNDVFSS	448
HMBRB	QRDFENLRDDYYFSGRVVRTTNSIQHPVKTTNYGFSLSQIQWNDVFSS	449
HMBRC	RRDFENLRDDYYFSGRVVRTTSSIQHPVKTTNYGFSLSQIQWNDVFSS	450
HMBRMS11	RREFENLRDDYYFSERVSTSSIQHPVKTTNYGFSLSQIQWNDVFSS	448

HMBRA	RAGIRYDHTKMTPQELNAECHACDKTPPAANTYKGWSGFVGLAAQLNQAW	498
HMBRB	RAGIRYDHTKMTPQELNADCHACDKTPPAANTYKGWSGFVGLAAQLSQTW	499
HMBRC	RAGIRYDHTKMTPQELNAECHACDKTPPAANTYKGWSGFVGLAAQLNQAW	500
HMBRMS11	RADIRYDHTKMTPQELNADCHACDKTPPAANTYKGWSGFVGLAAQLNQAW	498

HMBRA
RVGYDITSGYRVPNASEVYFTYNHGSGNWL PNP NLKAERSTTHTLSLQGR
548

HMBRB
RVGYDVTSGFRVPNASEVYFTYNHGSGTWKPNPNLKAERSTTHTLSLQGR
549

HMBRC
RVGYDITSGYRVPNASEVYFTYNHGSGNWL PNP NLKAERTTHTLSLQGR
550

HMBRMS11
HVGYDITSGYRVPNASEVYFTYNHGSGNWL PNP NLKAERSTTHTLSLQGR
548

HMBRA
 HMBRB
 HMBRC
 HMBRMS11

DECLARATION AND POWER OF ATTORNEY
(Case No. 94,784-A)

As below-named inventors, we hereby declare that:

Our residences, post office addresses and citizenship are as stated below next to our names.

We believe we are the original, first and joint inventors of the subject matter which is claimed and for which a patent is sought on the invention entitled:

NOVEL BACTERIAL HEMOGLOBIN RECEPTOR GENES AND USES

which specification was filed on October 2, 1995 and given Serial No. 08/537,361

We hereby state that we have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

We acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, Sec. 1.56(a).

We further acknowledge the duty to disclose such material information which occurred between the filing date of the prior application, U.S. Serial No. 08/326,670, filed October 18, 1994 and the filing of this continuation-in-part application.

The undersigned hereby appoint the following:

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as our Registered Patent Agents

the mailing address and telephone number of each of whom is BANNER & ALLEGRETTI, LTD., Ten South Wacker Drive, Chicago, Illinois 60606, and (312) 715-1000, with full power of substitution and revocation to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith.

We hereby declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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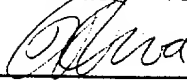
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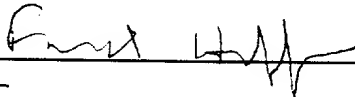
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